

Annual Review 2023/24

Redirect Recycling Wetherill Park

24 Davis Road, Wetherill Park NSW

Redirect Recycling

22 October 2024

Revision History

| Rev | Revision | Author / | Details | Authorised | |
|-----|------------|---------------|------------|-----------------|-----------|
| No. | Date | Position | | Name / Position | Signature |
| 1 | 22/10/2024 | James Sutton | For | James Sutton | *** |
| | | Environmental | submission | Environmental | Sulta |
| | | Manager | to DPHI | Manager | |



Table of Contents

| 1 | | Introduction | 4 |
|----|-------|--|----|
| | 1.1 | Scope | |
| | 1.2 | Background | 4 |
| | 1.3 | Consent | |
| | 1.4 | Annual Review Requirements | |
| | 1.5 | Environment Protection Licence | |
| | 1.6 | Water Licences | |
| | 1.7 | Trade Waste Licence | |
| | 1.8 | Environmental Management Plans | |
| | 1.9 | Contacts | |
| | 1.10 | Actions Required from Previous Annual Review | |
| 2 | | Operations during the Reporting Period | |
| | 2.1 | Production | |
| | 2.2 | Facility Improvements | |
| | 2.3 | Site Activities | |
| 3 | | Waste Management | |
| | 3.1 | Solid Waste | |
| _ | 3.2 | Trade Waste | |
| 4 | | Environmental Monitoring and Performance | |
| | 4.1 | Environmental Management System | |
| | 4.2 | Air Quality | |
| | 4.3 | Surface Water | |
| | 4.4 | Groundwater | |
| | 4.5 | Noise | |
| _ | 4.6 | Traffic | |
| 5 | | Community Relations | |
| | 5.1 | Environmental Complaints | |
| _ | 5.2 | Community Liaison | |
| 6 | | Independent Audit | |
| 7 | | Environmental Incidents & Non-compliances | |
| | | ncidents | |
| | 7.2 N | lon-conformances | |
| 8 | | Activities Proposed for the next Annual Review Period | 26 |
| Α | PPE | NDICIES | 27 |
| Α | pper | ndix A – Annual Surface Water and Baseline Groundwater | r |
| | | tion Report | |
| | | ndix B – Example Quarterly OEMP Checklist | |
| | | ndix C – Appendix D – Community Complaints | |
| ~ | hhai | idix 6 – Appendix D – Community Complaints | JU |
| Fi | aure | 1 Regional Context | |
| Fi | aure | 2 SSD 7401 Approved Development Area | 1 |
| | | 3 Groundwater Monitoring Locations | |



Annual Review Title Block

| Name of operation | Redirect Recycling |
|--|--------------------|
| Name of operator | Redirect Recycling |
| Development consent / project approval # | SSD 7401 |
| Name of holder of development consent / project approval | Bettergrow Pty Ltd |
| Mining lease # | N/A |
| Name of holder of mining lease | N/A |
| Water Access Licence # | N/A |
| Name of holder of water licence | N/A |
| MOP/RMP start date | N/A |
| MOP/RMP end date | N/A |

I, James Sutton, certify that this audit report is a true and accurate record of the compliance status of Redirect Recycling Pty Ltd for the period 23rd August 2023 to 22nd August 2024 and that I am authorised to make this statement on behalf of Redirect Recycling Pty Ltd Note.

- a) The Annual Review is an 'environmental audit' for the purposes of section 122B(2) of the Environmental Planning and Assessment Act 1979. Section 122E provides that a person must not include false or misleading information (or provide information for inclusion in) an audit report produced to the Minister in connection with an environmental audit if the person knows that the information is false or misleading in a material respect. The maximum penalty is, in the case of a corporation, \$1 million and for an individual, \$250,000.
- b) The Crimes Act 1900 contains other offences relating to false and misleading information: section 192G (Intention to defraud by false or misleading statement—maximum penalty 5 years imprisonment); sections 307A, 307B and 307C (False or misleading applications/information/documents—maximum penalty 2 years imprisonment, \$22,000, or both.)

| Name of authorised reporting officer | James Sutton |
|---|---------------------|
| Title of authorised reporting officer | Environment Manager |
| Signature of authorised reporting officer | J. Sutto |
| Date | 22/10/2024 |



1 Introduction

1.1 Scope

This Annual Review has been prepared for the Redirect Recycling Pty Ltd (reDirect) Wetherill Park site and covers the twelve-month reporting period from 23 August 2023 to 22 August 2024. This Annual Review has been prepared to satisfy condition C9 of Development Consent SSD 7401 issued by the Minister for Planning on 11 October 2017.

The reDirect facility is located at 24 Davis Road, Wetherill Park NSW and consists of a resource recovery facility purpose built for washing and processing of construction and liquid waste.

This Annual Review is submitted to NSW Department of Planning, Housing and Infrastructure (DPHI). The Annual Review is also made available on the reDirect website:

Redirect Recycling

reDirect maintained compliance with all necessary approvals and licenses (EPL 21092 & SSD-7401) during the reporting period recording nil non-compliance items.

Table 1 Compliance

| Relevant approval | Condition | Condition description (summary) | Compliance status | Comment | Where addressed in Annual Review |
|----------------------|-----------|---------------------------------|----------------------|---------|---|
| SSD 7401 | C9 | Annual review | Compliant | Nil | 1.4 Annual Review Requirements |
| EPL 21092 | L3.1 | Noise | Compliant | Nil | Section 4.5 Noise |
| EPL 21092 | O3.1 | Air Quality | Compliant | Nil | Section 4.2 Air Quality |
| EPL 21092 | O5 | Water Quality | Compliant | Nil | 4.3 Surface Water & 4.4 Groundwater |

1.2 Background

Consent for State Significant Development 7401 (SSD-7401) was initially granted by the NSW Department of Planning Housing and Infrastructure (DPHI) on 22 December 2017. The facility was commissioned in August 2022 and shortly after the licence was transferred to reDirect (a Borg Company) who currently operate the site (see Figure 1).



Figure 1 Regional Context



The development has been staged with only the wash plant operational at this time. The landscape supplies food and garden organics approvals are not operational. Stage 1 includes the wash plant processing area only. Facilities covered under Stage 1 include:

- A main administration building, office and carpark constructed at the fore of the property. Site amenities, including toilets and kitchen, contained in the main administration building.
- Partially enclosed shed space, containing:
 - o Two tier ground levels with external ramp to the west of the shed.
 - o Four hydro-tips, and one dry feed hopper.
 - One weighbridge located west of the shed for the weighing of trucks on entry and prior to departure from the facility.
 - Screening walls.
 - Drill mud processing plant and equipment.
 - Drill mud machinery control rooms and internal office space.
- An inground sand filter located under the hardstand on the lower level of the site, adjacent to the south-western corner of the existing approved shed, to be used for stormwater retention and treatment.
- Rainwater / raw water storage tanks.
- Main thoroughfare, including:
 - A combined ingress/egress access driveway, providing a 12.5 m width at the western property boundary and facilitating connectivity between the off-street parking and internal heavy vehicle circulation areas.
 - Off-street parking spaces designed in accordance with AS2890.1 and AS2890.6.



- A combined ingress / egress driveway, providing a 5.5 m width adjacent to the eastern property boundary facilitating service access to the office complex and emergency access for Fire NSW.
- Internal hardstand areas and roadways.

The main waste types and materials accepted at the site include:

- Hydro-excavation and drill muds;
- Concrete slurry;
- Stormwater:
- · Street sweepings; and
- General solid waste (soils that meet EPL conditions).

1.3 Consent

Consent for State Significant Development 7401 (SSD-7401) was initially granted by the then NSW Department of Planning and Environment (DPHI) on 22 December 2017. Consent for Modification 1 of SSD-7401 (SSD-7401-MOD-1) was approved by the NSW Department of Planning, Industry and Environment (DPIE) on 21 April 2021, with consent for Modification 2 (SSD-7401-MOD-2) granted on 30 November 2021. Consent for Modification 3 (SSD-7401-MOD-3) was granted by DPHI (name reverted from DPIE) on 1 April 2022. Consent for Modification 4 (SSD-7401-MOD-4) was granted by Department of Planning, Housing and Infrastructure (DPHI) (name reverted from DPHI).

Approval for SSD-7401 permitted the construction and operation of a resource recovery facility to process up to 160,000 tonnes per year of waste comprising of:

- 60,000 tonnes per annum (tpa) of hydro-excavation, drill muds and fluids.
- 70,000 tpa of food and garden organics.
- 30,000 tpa of packaged and bulk food and liquids.

In addition, the approval for SSD-7401 allowed for the operation of a landscaping material supplies facility for the storage and sale of up to 40,000 tpa of landscaping supplies.

Approval of SSD-7401-MOD-1 allowed for the increase of processing capacity to 350,000 tpa in conjunction with the following:

- Introduction of additional waste streams.
- Demolition of existing structures.
- · Construction of a partially enclosed shed.

SSD-7401-MOD-2 included the replacement of the 30, 000 L sediment basin and associated bioretention basin, located within the southwest corner of the subject site. In lieu of the detention and bioretention basins it was proposed to utilise an existing inground concrete pit that remains onsite as part of a decommissioned weighbridge. This pit was modified and improved to include a sand filter to treat onsite stormwater.

SSD-7401-MOD-3 included the following:

- Replacement of the five (5) approved weighbridges with one (1) 25 m by 4.2 m weighbridge located approximately 55 m from the Facility intersection with Davis Road.
- To facilitate weighbridge installation and improve site safety, vehicle parking spaces were reconfigured:
 - o Five (5) parking spaces immediately east of the existing site office.
 - Two (2) parking spaces located north of the inground sand filter, abutting the western facade of the drill muds processing shed.



- Five (5) parking spaces located on the hardstand area immediately north of the western parcel of retained Cumberland Plain Woodland.
- Remaining parking spaces were not altered.
- Relocation of proposed humeceptor water treatment device to the north-western corner of the central portion of Cumberland Plain Woodland onsite.
- Relocation of the 5,000 L rainwater tank to inside drill muds processing shed next to the control room. Rainwater from the existing office will now be captured via the Facility stormwater network.

SSD-7401-MOD-4 included an administrative amendment to reflect additional waste streams of concrete slurry and stormwater, originally assessed as appropriate in Modification 1, in the limits of consent.

This Annual Review covers facility operations conducted under Stage 1 of SSD-7401 (including modifications). Stage 2 (bulk landscape area and the organics processing area) is not operational, therefore assessment of conditions specific to Stage 2 have not been triggered or included within this report.

A summary of development consents including modifications currently held by Bettergrow Pty limited (original applicant) is presented in Table 2.

Table 2 Development Consents

| Consent Description | Approval Date | Approval Authority | Approved Development |
|--|---------------------|------------------------------|---|
| Development Consent SSD 7401 | 22 December 2017 | NSW Minister for Planning | The construction and operation of a resource recovery facility to process up to |
| | | | 160,000 tonnes per year of waste comprising of: |
| | | | 60,000 tpa of hydro-excavation, drill muds and fluids; |
| | | | 70,000 tpa of food and garden organics; and |
| | | | • 30,000 tpa of packaged and bulk food and liquids. |
| | | | The operation of a landscaping material supplies facility for the storage and |
| | | | sale of up to 40,000 tpa of landscaping supplies. |
| Development Consent SSD 7401 MOD 1 | 21 April 2021 | NSW Minister for Planning | Increase the processing capacity to 350,000 tpa of waste; introduce additional waste streams; demolish existing structures; construct a partially |
| | | | enclosed shed; and increase the hours of operation to 24/7. |
| Development Consent SSD 7401 MOD 2 | 30 November 2021 | NSW Minister for Planning | Amend the stormwater management system to include the use of an in-ground concrete pit with sand filter. |



| Consent Description | Approval Date | Approval Authority | Approved Development |
|--|-----------------|------------------------------|--|
| Development Consent SSD 7401 MOD 3 | 31 March 2022 | NSW Minister for Planning | Amend the carparking configuration, replace the five on-site weighbridges with one weighbridge, relocate the 5 kilolitre underground rainwater tank to an above ground tank inside the drill muds processing shed and replace and relocate the Humeceptor with an Ecoceptor. |
| Development Consent SSD 7401 MOD 4 | 25 January 2024 | NSW Minister for Planning | Administrative amendment to reflect additional waste streams of concrete slurry and stormwater, originally assessed as appropriate in Modification 1, in the limits of consent. |

1.4 Annual Review Requirements

In accordance with condition C9 of Development Consent SSD 7401, annual review requirements and the sections within this review where these are addressed have been summarised in Table 3.

Table 3 Annual Review Requirements

| Develo | oment Consent SSD 7401 – Condition C9 | Section of Annual Review | | |
|--------|--|-----------------------------|--|--|
| | ar, the Applicant must review the environmental performance of the ment to the satisfaction of the Planning Secretary. This review must: | This Report | | |
| (a) | describe the development that was carried out in the previous calendar year, and the Development that is proposed to be carried out over the next year; | Section 2 Section 8 | | |
| (b) | include a comprehensive review of the monitoring results and complaints records of the Development over the previous reporting period, which includes a comparison of these results against the: i. the relevant statutory requirements, limits or performance measures/criteria: | Section 4 Section 5 | | |
| | ii. requirements of any plan or program required under this consent; iii. the monitoring results of previous years; and iv. the relevant predictions in the EIS; | | | |
| (c) | identify any non-compliance during the reporting period, and describe what actions were (or are being) taken to ensure compliance; | Section 4 Section 7 | | |
| (d) | identify any trends in the monitoring data over the life of the Development; | Section 4 | | |
| (e) | se) identify any discrepancies between the predicted and actual impacts of the Development, and analyse the potential cause of any significant discrepancies; and | | | |
| (f) | describe what measures will be implemented over the next reporting period to improve the environmental performance of the Development. | Section 8 | | |

1.5 Environment Protection Licence

reDirect operates in accordance with Environment Protection Licence 21092 (EPL 21092), issued by the NSW Environment Protection Authority (EPA) under Section 55 of the *Protection of the Environment Operations Act 1997*. The current Licence version date is 01 June 2023.



1.6 Water Licences

reDirect does not hold any water licences.

1.7 Trade Waste Licence

reDirect's Trade Waste Service Contract with Sydney Water for the discharge of liquid trade waste into Sydney Water's sewerage system was initially approved on 01 August 2022 prior to the site's operational start date (23 August 2022). Throughout the reporting period, trade wastewater sampling was conducted every 60 days or on the day the trade waste was discharged thereafter. Substance characteristics analysed included:

- Biochemical Oxygen Demand
- Ammonia (As N)
- Sulphate
- Suspended Solids
- Total Dissolved Solids

reDirect have not had any trade waste sampling results exceed the criteria. Results were consistent with the previous reporting year (2022/2023).

1.8 Environmental Management Plans

As per Schedule 2 Part C of SSD 7401, the existing development is carried out in accordance with the Operational Environmental Management Plan (OEMP) and associated sub-plans.

In accordance with C8 Revision of Strategies, Plans and Programs, environmental management plans are required to be reviewed within three months of completion of an audit under C14 and/or approval of an annual report review under C9.

reDirect received correspondence from DPHI (8 December 2023) determining the 2023 Independent Environmental Audit to generally satisfy the reporting requirements of the consent and the NSW Planning *Independent Audit Post Approval Requirements* (2020). Additional correspondence was also received from DPHI (10 January 2024) determining the Annual review undertaken for the period 23 August 2022 to 22 August 2023 to generally satisfy the reporting requirements of the consent.

In accordance with C8 reDirect conducted a review of all management plans accordingly. The following management plans were reviewed.

- Operational Environmental Management Plan
- Air Quality and Odour Management Plan
- Stormwater Management Plan
- Operational Waste Management Plan
- Flood Emergency Plan
- Water Management Plan
- Emergency Plan
- Operational Traffic Management Plan
- Conceptual Decommissioning management Plan

Based on the findings of the previous Independent environmental Audit and Annual Review, no changes were deemed required to any of the management plans. A record of the review



was recorded in the relevant document control section of each management plan and the plans re-published on the reDirect website on 10 January 2024.

1.9 Contacts

Table 4 outlines the contact details for site personnel responsible for managing environmental operations at the reDirect facility.

Table 4 Site Personnel

| Name | Title | Contact Details |
|---------------|-----------------------|-----------------|
| Neale Hogarth | Manager | 0498 692 443 |
| James Sutton | Environmental Manager | 0414 987 168 |

1.10 Actions Required from Previous Annual Review

Table 5 represents activities proposed in Section 8 of Annual Review 22/23 and corresponding comments regarding outcomes of those proposed activities.

Table 5 Proposed Activities in 2023/24 Reporting Period

| Activities Proposed in Reporting Period | Results achieved in Reporting Period |
|---|---|
| Ongoing implementation of Environmental Management Plans for the existing development and the project. | Operational staff have continued to implement daily inspection checklists (as required under OEMP). No non-compliance or notifiable incidents have occurred. |
| Complete installation of new centrifuge to increase efficiency in material processing. | New centrifuge was installed and commissioned in November 2023. |
| Attain new site-specific resource recovery order and exemption (SSRRO/E) for processed materials allowing new uses and increased efficiency for resource recovery activities. | Approved SSRRO/E's: The reDirect washed clay fines order July 2024 The reDirect washed sand order July 2024 The reDirect washed aggregate order July 2024 The reDirect washed clay fines exemption July 2024 The reDirect washed sand exemption July 2024 The reDirect washed aggregate exemption July 2024 |
| Continue erosion and sediment control inspections and rectification works as necessary to manage stormwater discharge. | Operational staff have continued to implement daily inspection checklists (as required under OEMP). Checklist have documented regular sweeping of the site, inspection and maintenance of sediment control infrastructure. |
| Update current operational management plans to reflect recommendations from audit and findings from annual review. | Operational management plans were reviewed following the completion of the previous 2023 Independent Environment Audit and Annual Review. No changes were deemed required; the review was recorded in each respective document control table. |

2 Operations during the Reporting Period

2.1 Production

Development Consent SSD 7401 allows for the receival and processing of up to 350,000 tonnes of waste per year, including 100,000 tonnes of liquid waste and 150,000 tonnes of general solid waste. During the reporting period reDirect received and processed a total of 80,031.08 tonnes of combined liquid and general solid waste. A total of 48977.81 tonnes were



recovered and beneficially reused under applicable resource recovery orders, 184.02 tonnes were sent for lawful disposal, comprising of trash and light organics.

2.2 Facility Improvements

The following improvements were made to site infrastructure, plant and/or equipment during the reporting period:

 Installation of a second centrifuge for increased processing efficiency and ability to continue processing in case of breakdown or maintenance.

See Figure 2 for location of site infrastructure.

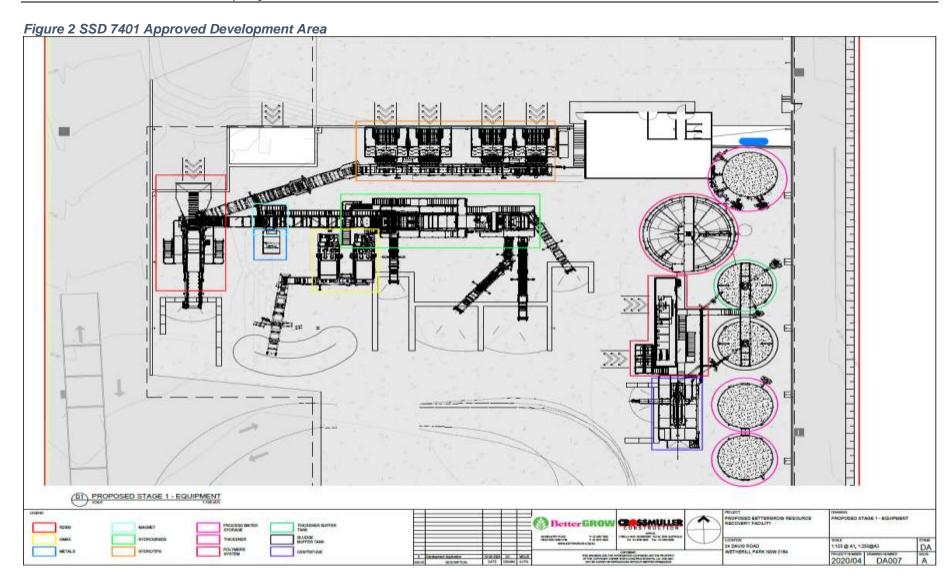
2.3 Site Activities

Environmental commitments and management/mitigation measures that were applied during the reporting period include the following:

- Operational works undertaken in accordance with the Operational Environmental Management Plan and sub-plans;
- Surface water sampling events;
- Background groundwater quality sampling events;
- Site environmental inspections; and
- Site wide communication of environmental requirements via inductions and Toolbox Talks.

No activities associated with additional construction were undertaken within the reporting period.







3 Waste Management

Waste generated at the reDirect site is managed in accordance with the Waste Management Plan that has been developed for the facility. The management process incorporates a system of recycling and reuse of waste materials where possible. Waste that cannot be incorporated into this system is removed from site and taken to landfill for lawful disposal.

3.1 Solid Waste

A summary of waste and resource recovery materials removed from reDirect Wetherill Park during the reporting period is provided in Table 6.

Table 6 Waste Management 2023/24

| Month | Destination | | | |
|---------------|-------------|-----------------------------|--------------------------------|--|
| | Tonnes | Waste | Reuse / Disposal | |
| September | 948.22 | Recovered aggregate 5-20mm | | |
| 2023 | 32.76 | Recovered aggregate 20-40mm | | |
| | 1402.54 | Treated drill mud | Resource recovery material | |
| | 609.02 | Washed sand | | |
| October 2023 | 445.58 | Recovered aggregate 5-20mm | | |
| | 543.96 | Recovered aggregate 20-40mm | December 1 | |
| | 1349.90 | Treated drill mud | Resource recovery material | |
| | 1476.82 | Washed sand | | |
| November 2023 | | Recovered aggregate 5-20mm | | |
| | 1361.26 | Recovered aggregate 20-40mm | | |
| | 470.24 | Recovered aggregate 40-80mm | Resource recovery material | |
| | 1817.18 | Treated drill mud | | |
| | 1232.44 | Washed sand | | |
| December 2023 | 37.48 | Recovered aggregate 5-20mm | | |
| | 1674.08 | Treated drill mud | Resource recovery material | |
| | 774.40 | Washed sand | | |
| January 2024 | 43.30 | Organics / light trash | Central Waste – Kurri Kurri | |
| | 144.84 | Recovered aggregate 5-20mm | | |
| | 202.72 | Recovered aggregate 40-80mm | December we serve we week wind | |
| | 2186.38 | Treated drill mud | Resource recovery material | |
| | 624.42 | Washed sand | | |
| February 2024 | 357.82 | Recovered aggregate 5-20mm | | |
| | 105.46 | Recovered aggregate 20-40mm | | |
| | 2373.38 | Treated drill mud | Resource recovery material | |
| | 407.60 | Washed sand | | |



| March 2024 | 404.00 | B | |
|---------------|----------|-----------------------------|---|
| IVIAIUII ZUZ4 | 134.32 | Recovered aggregate 5-20mm | |
| | 3236.43 | Treated drill mud | Resource recovery material |
| | 918.84 | Washed sand | |
| April 2024 | 247.10 | Recovered aggregate 5-20mm | |
| | 4051.19 | Treated drill mud | Resource recovery material |
| | 891.56 | Washed sand | |
| May 2024 | 139.74 | Recovered aggregate 5-20mm | |
| | 36.50 | Recovered aggregate 20-40mm | |
| | 73.10 | Recovered aggregate 40-80mm | Resource recovery material |
| | 3940.91 | Treated drill mud | |
| | 862.14 | Washed sand | |
| June 2024 | 140.72 | Organics / light trash | Central Waste – Kurri Kurri |
| | 188.14 | Recovered aggregate 5-20mm | |
| | 86.98 | Recovered aggregate 20-40mm | |
| | 33.38 | Recovered aggregate 40-80mm | Resource recovery material |
| | 2973.22 | Treated drill mud | |
| | 1203.06 | Washed sand | |
| July 2024 | 317.88 | Recovered aggregate 5-20mm | |
| | 2580.12 | Treated drill mud | Resource recovery material |
| | 1093.76 | Washed sand | |
| August 2024 | 41.86 | Recovered aggregate 5-20mm | Resource recovery material |
| | 3392.72 | Treated drill mud | |
| | 699.94 | Washed sand | |
| TOTAL | 184.02 | Organics / light trash | Wanless Waste Management Kemps Creek |
| | 4050.5 | Recovered Aggregate 05–20mm | Resource recovery material |
| | 2166.92 | Recovered Aggregate 20–40mm | Resource recovery material |
| | 779.44 | Recovered Aggregate 40–80mm | Resource recovery material |
| | 30978.05 | Treated Drilling Mud | Resource recovery material |
| | 10794 | Washed Sand | Resource recovery material |

Waste types in Table 6 are further described as:

- Organics and Light Trash: General waste including a mix of organics such as sticks, leaf litter and other organic matter mixed with light film plastic and other small anthropogenic inclusions.
- Resource Recovery Material: Material meeting a general or site-specific resource recovery order made under clause 93 of the 2014 Waste Regulation and/or section 286A of the Protection of the Environment Operations Act 1997.



There was no trackable waste generated during this reporting period.

3.2 Trade Waste

Redirect's current trade waste agreement (Consent no: 51950) allows for the following discharge rates to Sydney Water's wastewater system:

- Instantaneous maximum rate of pumped discharge 8,000 litres per second
- Maximum daily discharge 320 kilolitres
- Average daily discharge 200 kilolitres

The last sampling event conducted during the reporting period was completed on 24 July 2024, 29 days prior to the end of the reporting period. A total of 147 kilolitres were discharged during the sampling event, well below the average and maximum daily discharge limits. Additionally, sampling completed since the commencement of the agreement confirmed a total of 83515.33 kilolitres had been disposed as trade waste up to this date, equating to a daily average below 200 kilolitres.

Water discharged to trade waste has nearly doubled when compared to the readings comparable from the previous reporting year (23 August 2023 - 02 August 2024, 20,487 Kilolitres and 02 August 2023 - 24 July 2024, 55,028.33 Kilolitres). This increase is attributed to increased waste throughput in the facilities second full operational year.

4 Environmental Monitoring and Performance

4.1 Environmental Management System

ReDirect operates in accordance with the Operational Environmental Management Plan (OEMP) as documented in Section 1.8. This OEMP aims to ensure adequate management, monitoring and mitigation systems are in place to protect the surrounding environment. Similarly, construction activities are undertaken in accordance with the Construction Environmental Management Plan (CEMP).

Environmental performance and management are conducted in accordance with the requirements of SSD 7401, its subsequent modifications (MOD1, MOD2, MOD3 & MOD4), and EPL 21092. Environmental performance and monitoring are an integral part of environmental management system. The measurement and evaluation of monitoring results allows for the assessment of performance against quantitative and qualitative standards and assists in the identification of any non-conformances or areas that may require additional attention.

4.2 Air Quality

Air quality is monitored in accordance with the reDirect's Operational Air Quality and Odour Management Plan (AQOMP). Condition O3.1 of EPL 21092 states that:

Additionally, Condition L5.1 of EPL 21092 states that:

[&]quot;The premises must be maintained in a condition which minimises or prevents the emission of dust from the premises."

[&]quot;The licensee must not cause or permit the emission of offensive odour beyond the boundary of the premises."



EPL 21092 does not specify dust monitoring be undertaken, the AQOMP assessed material handling and processing in the drill mud processing plant to have minimal fugitive dust emissions due to the high moisture content of waste received and retained within recovered processed materials. Additionally, road surfaces at the Site are sealed and processing is undertaken within the partially enclosed shed, currently no other activities approved under SSD-7401 are undertaken as part of the development. All current dust management procedures undertaken as part of the AQOMP and OEMP are currently deemed suitable and effective.

As Stage 1 operations only involves the drill mud processing plant, dust emissions have been identified as the only air quality impact associated with these operations. Therefore, no management of odour generating activities was required during the reporting period.

4.3 Surface Water

Surface water is considered any water other than process water, leachate or wastewater being defined as:

- Process water is water used in the processing of drill muds.
- Leachate is water generated through rain coming into contact with soil stockpiles.
 Leachate is not anticipated to be generated onsite during Stage 1 of operations due to bulk storage bays being underneath the main processing shed.
- Wastewater is water generated through the processing of drill muds that require disposal or have no further use on site.

Surface water is, thus, principally stormwater runoff from building roofs and areas outside waste processing or handling areas.

Surface water discharges from operational areas of the site and areas with potential to discharge off-site are summarised in the following table. Surface water may also discharge from other areas of the site, but these areas are away from operational areas.

Table 7 Surface Water Sources and Management

| Site Feature | Purpose | Runoff Water Sources | Management |
|---------------------------------|--|--|---|
| Entrance Driveway | Site access | The driveway receives runoff from paved areas near the weighbridge and entrance areas. | Management under the surface water management plan – though this is considered a low risk of impact. |
| Drill Mud Processing Shed | Rainwater re- use | A portion of roof water runoff from the drill mud processing shed is to be directed by downpipes to an above-ground rainwater harvesting tank which has been sized to meet the facility's reuse demand for non-potable water of 5 kL. The harvested volume from the shed roof is reused internally through the amenities connections with tank overflows being diverted directly to the stormwater system. The remainder of the roof water collected is to be directed to the stormwater system. | Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. Regularly check the structural integrity of the tanks. Check for any accumulated litter, sediment, or debris on or within the tanks. |
| Stormwater System | Collection, treatment and transportation | Runoff from majority of sealed surfaces on the site, all roof areas not connected to the rainwater tank | Management under the stormwater management plan (Eclipse 2021) and the WMP. |



| of stor | , | d rainwater tank overflow rted into the stormwater | Remove deposited sediment and debris from the sand filter bed/detention pit and Ecoceptor inlet/outlet areas. Regularly check the structural integrity of |
|---------|---|--|---|
| | | | hydraulic structures. |

In accordance with the reDirect Water Management Plan, six-monthly (following a rainfall event) sampling of two sampling points on-site (SW1 in the sand filter and SW2 in the ecoceptor outflow sampling point) was undertaken during this reporting period.

A summary of the results is presented in Table 8 and Table 9.

Table 8 Surface Water Observations and Geotechnical Requirements

| Location | Event | Dissolved Oxygen (mg/L) | Electrical Conductivity (μs/cm) | рН | Redox (Eh) (mV) | Temperature (°C) | Observations |
|-----------------------------|------------------|-------------------------------|---------------------------------------|------|-----------------------|---------------------|--|
| SW1 (untreated water) | February 2024 | 3.07 | 574 | 7.51 | 307 | 23.9 | Colourless, no odour, no sheen, suspended sediments |
| | July 2024 | 1.06 | 334 | 7.08 | 243 | 13.6 | Light grey brown, no odour, no sheen, moderately turbid |
| Location | Event | Dissolved Oxygen (mg/L) | Electrical Conductivity (μs/cm) | рН | Redox (Eh) (mV) | Temperature (°C) | Observations |
| SW2 (treated water) | February 2024 | 4.19 | 656 | 7.70 | 286 | 25.0 | Colourless, no odour, no sheen, suspended sediments |
| | July 2024 | 6.09 | 370 | 8.48 | 187 | 13.7 | Light grey, no odour, no sheen, slightly turbid |
| | • | | • | | • | · | • |



| Table 9 Surface Water Analytical Summ |
|---------------------------------------|
|---------------------------------------|

| Analyte / Value | Screening Criter | ia Exceedances | | Comment | | |
|--------------------------------|------------------|---|-----------------|---|--|--|
| | Health-Risk | Ecological Risk | Aesthetics | | | |
| Heavy metals and metalloids | None identified | Heavy metal concentrations were reported at low levels, less than | Fig. | Metal concentrations were generally less than or similar to relevant screening criteria for disturbed ecosystems consistent with the WMP. | | |
| | | relevant screening criteria for highly disturbed environments, with the exceptions of: | | Reported copper concentrations were slightly elevated above ecological screening criteria at both SW1 (untreated) and SW2 (treated), indicating that the treatment train has not reduced copper concentrations in the stormwater. | | |
| | | Copper (at SW1 and SW2 in February 2024 and July 2024). Zinc (at SW2 in July 2024). | | The reported zinc concentrations at SW2 in February 2024 and July 2024 were higher than reported in SW1. This scenario was also noted in August 2023. The cause for the increase in zinc concentrations is unclear, but may be related to the treatment train and should be reviewed. The average concentration of zinc in SW2 over the four sampling events in 2023 and 2024 (0.02 mg/L) was also above the ecological criterion. | | |
| Nutrients | None identified | No exceedances for toxicants. Exceedances of conservative physical stressor values for total oxidised nitrogen (as N), TN and TP at SWM and SWD in both | ×25 | Concentrations are similar to the median values for TN (1.7 mg/L) and TP (0.31 mg/L) in stormwater runoff in urban or commercial/industrial areas in east coast Australia reported by Drapper et al (2022) and Fletcher et al (2004). TN concentrations are generally less than DGV (90% spp) for nitrate (as N) (5.6 mg/L). The concentrations of TN and TP were lower in SW2 (treated) than in SW1 (untreated). This is further | | |
| | | February 2024 and July 2024. | | discussed below. | | |
| Organic CoPC | None identified | Exceedance of TRH >C10-C16 | None identified | BTEX, PAHs and phenois were not detected in water samples. | | |
| | | fraction minus naphthalene (F2) at SW1 in February 2024. | | TRH >C10-C16 fraction minus naphthalene (F2) exceeded the ecological criteria at SW1 in February 2024, but was reported below the laboratory limit of reporting (LOR) in SW2, indicating that the treatment train was effective in removing the petroleum contamination. The concentrations of TRH in both SW1 and SW2 were below the LOR in July 2024, potentially indicating that there may have been an unreported or undetected spill or leak of petroleum products during February 2024. | | |
| Physico-chemical Parameters | None identified | None identified | None identified | The TSS was almost two orders of magnitude greater in July 2024 compared to February 2024 at SW1, which may be related to the rainfall preceding both events. The concentrations reported in SW2 were lower (by at least one order of magnitude), indicating that the treatment train is effective in reducing the TSS concentration under a range of furbidity conditions. | | |

There are indications that concentrations of key parameters (TSS, TN and TP) are lower at SW2 (downstream of treatment train) than SW1 (upstream of system), which was also noted during the 2023 sampling events. The water, sediment and erosion controls in the WMP should continue to be followed to minimise migration of sediments and fines into the stormwater system. Annual surface water monitoring should continue from both SW1 and SW2.

A copy of the *Annual Surface Water and Baseline Groundwater Condition Report - 2024* (Senversa, 2024) has been included in Appendix A.

4.4 Groundwater

In accordance with the reDirect Water Management Plan, a monitoring network was established, including the installation of 6 shallow groundwater monitoring wells that intersect the water table located within the shale bedrock.

These new wells were installed as part of the site infrastructure upgrades. Senversa (engaged by reDirect) designed a groundwater monitoring network that seeks to characterise groundwater both hydraulically up-gradient and down-gradient of the site. The location of the groundwater monitoring wells is presented on **Figure 3**. The groundwater monitoring network comprises:

 One well (MW06) that captures the quality of background groundwater migrating onto the site from the north.



- Five wells (MW01, MW02, MW03, MW04, MW05) placed in targeted locations with the following rationale.
- MW01 Down gradient of the stormwater treatment sand filter box.
- MW02 Down gradient of the Ecoceptor.
- MW03 Western site boundary down gradient of neighbouring property.
- MW04 Down gradient of the drill mud processing facility on eastern boundary.
- MW05 Middle level of site in the vicinity of the historic aboveground storage tanks (ASTs).

The wells target the shallow groundwater as this is most susceptible to impact.



Figure 3 Groundwater Monitoring Locations



A baseline monitoring event is required, with ongoing groundwater monitoring conducted on a periodic basis. Additional monitoring will likely be required - triggered as a response to changes in site activities such as the commencement of Stage 2 operations. The monitoring locations, and sampling, analytical and reporting schedules are provided in Table 10.



Table 10 Groundwater Monitoring Frequency

| Frequency | Monitoring Aspect | Locations | Analytical Schedule | Reporting Schedule |
|---|--|--|--|---|
| Sampling every 6 months for a two year period | Gauging, <u>sampling</u> and analysis | MW01, MW02, MW03, MW04, MW05, MW06 | Field: pH, electrical conductivity (EC), dissolved oxygen (DO) and redox potential. Laboratory: Ammonia (as N), nitrate, TN, TP, dissolved metals, TPH, BTEX, PAH. | Interpretive baseline report |
| Annual, then reviewed after three years | Gauging, <u>sampling</u> and analysis | MW01, MW02, MW03, MW04, MW05, MW06 | Field: pH, EC, DO and redox potential. Laboratory: TRH, TN, JP and dissolved metals. Additional contaminants based on the findings of the baseline assessment. | Annual data report, then 3- year interpretative report |
| Triggered | Sampling and analysis* | As required* | As required* | Reporting as above |
| | Sampling every 6 months for a two year period Annual, then reviewed after three years | Sampling every 6 months for a two year period Annual, then reviewed after three years Gauging, sampling and analysis Gauging, sampling and analysis | Sampling every 6 months for a two year period Annual, then reviewed after three years Gauging, sampling and analysis Gauging, sampling and analysis MW01, MW02, MW05, MW06 MW01, MW02, MW06 MW01, MW02, MW06 | Sampling every 6 months for a two year period Annual, then reviewed after three years Gauging, sampling and analysis MW01, MW02, MW04, MW05, MW06 MW01, MW02, MW04, MW05, MW06 MW01, MW02, MW04, MW05, MW06 MW01, MW02, MW04, MW02, MW04, MW03, MW04, MW03, MW04, MW05, MW06 MW01, MW02, MW06 MW01, MW02, MW02, MW01, MW02, MW03, MW04, MW03, MW04, MW05, MW06 MW01, MW02, MW06 MW01, MW02, MW02, MW04, MW03, MW04, MW03, MW04, MW05, MW06 MW01, MW02, MW06 MW03, MW04, MW06 MW01, MW02, MW06 MW03, MW04, MW06 MW01, MW02, MW06 MW03, MW04, MW06 MW01, MW02, MW06 MW01, MW02, MW06 MW01, MW02, MW06 MW03, MW04, MW06 MW03, MW06 MW03, MW06 MW03, MW06 MW03, |

All sampling was undertaken by a suitably qualified and experienced person consistent with guidance in:

- DEC (2004). Approved Methods for Sampling and Analysis of Water Pollutants in NSW. March 2004.
- AS/NZS 5667.1:1998, Water Quality Sampling series.
- NEPC (2013). Schedule B (2) Guideline on Site Characterisation.

Appropriate data QA/QC procedures consistent with the above guidance were implemented and assessed as part of the program.

All analyses was conducted by a NATA accredited laboratory.

Groundwater management reporting requirements are outlined in Table 11.



Table 11 Groundwater Reporting Requirements

Report Type Content Details of monitoring scope and methods, and any non-conformances with this WMP. Baseline Groundwater Assessment Report Digitisation and analysis of historic groundwater monitoring results. (following completion of A plan showing monitoring locations. sampling) A plan showing groundwater elevations and inferred flow. Field records, calibration certificates and laboratory analytical certificates. Combined results for the first four monitoring events, including summary tables of gauging, field measurements and analytical data. Comparison of analytical results against performance criteria and historic results. Review of QA/QC. Statistical analysis of historical data for key chemicals of concern, including the mean, minimum, maximum, 80th percentile of site background groundwater quality (MW06) and baseline groundwater quality (at newly installed wells) to allow future comparison to Reporting shall be conducted in accordance with NSW EPA made or approved guidance. Details of monitoring scope and methods, and any non-conformances with this WMP. Data Report (annual) A plan showing monitoring locations. Field records, calibration certificates and laboratory analytical certificates. Tabulated results (gauging, field measurements and analytical data). Comparison of analytical results against performance criteria and baseline.

Condition L1 of the EPL states that the licensee must comply with section 120 of the POEO Act, which prohibits the pollution of waters. Assessment of groundwater quality will principally be via comparison against baseline and site background conditions. Table 12 below summarises the groundwater quality criteria to be adopted to assess whether pollution of waters may have occurred.

Table 12 Groundwater Reporting Requirements

| Receptor | Adopted Assessment Criteria |
|---|---|
| Change to baseline / background conditions | No statistically significant increasing trend or 20% increase over baseline / background concentrations or field parameters. |
| | Relevant criteria in NEPC (2013) for the commercial/industrial land use setting should be adopted as a screening levels. This includes: |
| Human Health | Direct contact criteria have also been considered due to the relatively shallow depth to groundwater in some locations. The presence of concrete and asphalt hardstand however indicates that groundwater will be predominantly inaccessible to humans. Drinking water guidelines will not be considered, given the site geology, land use and provision of a reticulated drinking water supply. |
| | Health Screening Level (HSL) for commercial/industrial land use (HSL-D) for vapour intrusion, sand aquifer, 2-<4 m based on the presence of fill and clay in the subsurface the most conservative soil type of sand has been selected. |
| | No gross aesthetic impacts such as non-aqueous phase liquids. |
| Ecological | Groundwater may migrate and discharge into Prospect Creek, which is the nearest surface water body down gradient of the site, though the ultimate receiving environment is the George's River and Botany Bay (marine). Northrop Pty Ltd (2017) indicate the local receiving waterways are heavily disturbed. The relevant ecological guidelines for toxicants, are therefore, the <u>fresh water</u> default guidelines values for heavily disturbed environments from ANZG (2018). |
| ns and the Table 1 12 | ANZG (2018) notes that exceedance of a DGV does not necessarily imply that there is an inherent risk, rather that further assessment and monitoring may be required prior to implementing appropriate management actions. These values should be used as 'triggers' for further assessment. |
| | |

These may be applied for screening purposes for groundwater that has the potential to migrate from the site.



The analytical results for the groundwater samples are presented in Table 4, with exceedances of the adopted site assessment criteria displayed on Figure 3 (Senversa, 2024). Table 13 below outlines the minimum, maximum and mean concentration of key CoPCs in each groundwater monitoring well during the four biannual sampling events, thus establishing the baseline groundwater conditions as required by the Surface Water Management Plan.

Table 13 Groundwater Statistical Analysis of Analytical Data

| | | Arsenic | Chromium | Copper | Manganese | Nickel | Zinc | Ammonia (as N) | Nitrate (as N) | BTEX | TRH (C6-C10) | TRH (>C10-C40) | PAH / Phenol |
|----------|-----------|---------|----------|--------|-----------|--------|--------|-------------------|-------------------|---------|-----------------|-------------------|-----------------|
| Criteria | tos. | 0.042 | 0.0033 | 0.0018 | 2.5 | 0.013 | 9.015 | 1.43 | 3.8 | 0.11* | 0.44° | 0.645 | 4 |
| (mg/t) | Health | 0.1 | 0.5 | 20 | 5 | 0.2 | 60 | * | 110 | 0.01* | 0.9h | 9.9° | 4 |
| Well ID | Statistic | | | | | | | | | | | | |
| MW1 | Min | 0.007 | -0.001 | +0.001 | 0.59 | 0.015 | 0.012 | 0.19 | -0.01 | -0.001 | -0.112 | -0.1 | =LOR |
| | Max | 0.012 | -0.001 | 0.015 | 4.84 | 0.156 | 0.174 | 0.71 | 0.02 | 40.001 | -0.02 | 0.52 | -LOR |
| | Mean | 0.010 | +0.001 | 0.005 | 2.15 | 0.058 | 0.062 | 0.47 | 0.01 | +0.001 | -0.02 | 0.2 | +LOR: |
| MW2 | Min | 0.004 | -0.001 | +0.001 | 0.96 | 0.005 | <0.005 | 0.26 | <0.01 | -0.001 | +0.02 | 10.1 | +LOR |
| | Max | 0.008 | +0.001 | 0.011 | 3.28 | 0.006 | 0.009 | 0.52 | 0.03 | +0.001 | +0.02 | +0.1 | +(.0H |
| | Mean | 0.006 | +0.001 | 0.003 | 1.75 | 0.006 | 0.007 | 0.44 | 0.02 | <0.001 | +0.02 | +0.1 | 4.08 |
| MW3 | Min | 0.002 | +0.001 | <0.010 | 6.15 | 0.191 | 0.122 | 0.22 | =0.01 | +0.001 | 10.02 | 19.1 | =0.0E |
| | Max | 0.011 | 0.005 | 0.006 | 7.4 | 9.207 | 0.253 | 0.29 | 0.10 | -0.001 | +0.02 | 0.91 | 40R |
| | Mean | 0.005 | 0.004 | 0.003 | 6.8 | 0.199 | 0.214 | 0.27 | 0.03 | <0.001 | -0.02 | 0.31 | -1.0R |
| MW4 | Min | 0.005 | <0.1101 | 40.001 | 4.00 | 0.011 | <0.005 | 0.28 | <0.01 | <0.901 | <0.02 | -0.1 | 4L06 |
| | Max | 0.00H | -0.001 | 0.005 | 5.04 | 0.021 | 0.006 | 0.34 | 0.01 | 0.002 | <0.02 | -0.1 | *L0R |
| | Mean | 0.007 | 40.001 | 0.002 | 5.12 | 0.017 | 0.002 | 0.31 | <0.01 | +0.001 | -0.02 | +0.1 | +LOR |
| MW6 | Min | -0.001 | -0.001 | <0.004 | -0.01 | 0.001 | =0.005 | =0.01 | 0.18 | +0.001 | -0.02 | 10.1 | -L08 |
| | Max | 0.002 | +0.001 | 0.003 | 0.225 | 0.002 | 0.006 | 0.09 | 1.93 | +0.001 | <0.02 | -0.1 | -4.0R |
| | Mean | 0.002 | +0.001 | 0.001 | 0.81 | 0.601 | 0.002 | 0.03 | 1.19 | < 0.001 | y0.02 | +0.1 | 408 |

Most conservative assessment criteria for BTEX displayed. Table 4 presents the applicable criteria for each BTEX compound.

Although the WMP outlined that annual groundwater monitoring be conducted for three years post completion of baseline monitoring, Senversa (2024) recommends that this is no longer required.

Baseline groundwater monitoring commenced at the same time as commencement of operations at the site and no detrimental statistical trends, considered to be associated with site operations, have been noted in groundwater quality during this time. It is considered unlikely that additional changes in groundwater quality would be noted after a further one year of groundwater monitoring given the sealed nature of the operational portion of the site, the low hydraulic conductivity of the underlying aquifer and adherence to the Applicant's Management and Mitigation Measures that form Appendix B of the Development Consent. Further triggers, in accordance with Table 5.2 of the WMP, for additional groundwater monitoring should include if additional processes commence at the site (e.g. food and garden organics [FGO], food and liquid depackaging [FLD], or other trackable liquid waste), if a potentially contaminating substance is to be stored or used/processed on the site, or a major incident occurs at the site (e.g. spill or leak of liquid substance/leachate, fire, etc). A review of the requirement for groundwater monitoring should be conducted every three years and when additional processes commence at the site.

4.5 Noise

In accordance with EPL 21092, noise from the premises must not exceed the limits noted in Table 13. In accordance with Development Consent SSD-7401 all construction activities related to the development must also comply with the limits in Table 13.

Assessment criteria for TRH OS-C10 fraction minus BTEX (F1) displayed

Adopted assessment criteria for TRH >C34-C40 displayed. Table 4 presents the applicable criteria for each TRH fraction.

^{*} Table 4 presents the adopted assessment criteria for each PAHIghenol compound, where available



Table 14 Noise Limits dB(A)

| Location | Day | Evening | Night | Night | |
|-------------------------|-----------------|-----------------|-----------------|----------------|--|
| | LAeq(15 minute) | LAeq(15 minute) | LAeq(15 minute) | LAeq(1 minute) | |
| All sensitive receivers | 35 | 35 | 35 | 45 | |

Day - The period from 7:00am to 6:00pm Note:

Evening – The period from 6:00pm to 10:00pm Night - The period from 10:00pm to 7:00am

LAeq means the equivalent continuous noise level - the level of noise equivalent the energy-average

of noise levels occurring over a measurement period.

4.5.1 Operational Noise

EPL 21092 stipulates that noise monitoring is to be carried out upon the request of an authorised NSW EPA officer. If requested, noise monitoring must be undertaken in accordance with Australian Standard AS 1055: 2018 Acoustics - Description and measurement of environmental noise, and the compliance monitoring guidance provided in the NSW Noise Policy for Industry (EPA 2017).

During the 2023/24 reporting period, reDirect was not requested to complete any noise monitoring.

Traffic 4.6

In accordance with the reDirect Operational Traffic Management Plan (OTMP), observations of compliance are to be undertaken at three monthly intervals, to document any remedial actions required with employees, heavy vehicle drivers or haulage companies.

reDirect carry out daily observations of traffic management and compliance against mitigation measures included within the OTMP. Observations are recorded on the Operational Environmental Management Plan – Wetherill Park Inspection Checklist. No breaches of traffic management procedures were recorded during the reporting period. Refer to Appendix C -Example Quarterly OEMP Checklist for example records.

Community Relations 5

5.1 **Environmental Complaints**

No community complaints were received during the 2023/24 reporting period.

5.2 **Community Liaison**

5.2.1 Information Exchange

In accordance with EPL 21092 condition M3.1 and M3.2, reDirect operate a telephone complaints line for the purpose of receiving any complaints from the members of the public in relation to activities conducted at the premises or by any vehicle or mobile plant. The complaints line is published on the reDirect recycling website, so the public know how to contact reDirect should a scenario trigger a complaint.



6 Independent Audit

Development Consent SSD 7401 condition C13 sets out requirements for independent environmental audits of the Development. reDirect commissioned environmental consultants RPS AAP Consulting Pty Ltd (RPS) to conduct an Independent Environmental Audit (IEA) of the site for operations audit period 23 August 2022 to 14 September 2023 (site inspection date) and construction period preceding operation of the site.

RPS noted good environmental management practices occurring at Redirect Recycling Wetherill Park. reDirect's compliance management consists of daily site inspection checklists, inspection of incoming loads and staff training. Overall, reDirect's general environmental management was commended. Appropriate shed layout, bunding and storage of materials, regular cleaning of the shed floor and other environmental management processes contribute to effective minimisation of the development's environmental impacts. The IEA concluded that the Development was undertaken generally in accordance with SSD 7016, the EIS and RTS, development layout plans and drawings, management and mitigation measures, and documents and drawings of the Existing Development.

There were 2 non-compliances (items) with 2 associated corrective actions raised. Corrective actions associated with Condition A27 have been completed by reDirect and required the provision of existing dilapidation to DPHI. Corrective actions associated with Condition B14 will not be triggered until further construction is planned and/or determined, due to the nature of specific detail required to be incorporated into the Construction and Demolition Waste Management Plan.

The IEA Report was submitted to DPIE on 17 October 2023.

In accordance with SSD 7401 condition C13 the next IEA is scheduled for August 2026.

7 Environmental Incidents & Non-compliances

Environmental incidents are managed through reDirect's Pollution Incident Response Management Plan (PIRMP) and are logged in DataStation, reDirect's incident management system. Each incident report details the issue, the corrective and preventative actions taken, and the responsibilities and timing for completion of the actions. The report also includes any additional comments relevant to the incident and the completion date of corrective actions.

7.1 Incidents

A pollution incident that requires notification is defined in section 147 of the Protection of the Environment Operations Act 1997 as:

- (a) Harm to the environment is material if:
 - i. It involves actual of potential harm to the health or safety of human beings or the ecosystems that is not trivial, or
 - ii. If results in actual or potential loss or property damage of an amount, or amounts in aggregate, exceeding \$10,000 (or such other amount as is prescribed by the regulations),
- (b) Loss includes the reasonable costs and expenses that would be incurred in taking all reasonable and practicable measures to prevent, mitigate or make good harm to the environment.

During this reporting period, there were no reportable environmental pollution incidents at the reDirect facility.



7.2 Non-conformances

reDirect Recycling have not been issued with any non-conformance or breach of licence correspondence from NSW DPHI or NSW EPA, respectively. Additionally, reDirect have not determined any non-compliances regarding operation of the site during the reporting period.

8 Activities Proposed for the next Annual Review Period

reDirect will endeavour to carry out the activities listed in Table 14 during the 2024/25 reporting period to assist with improving the environmental performance of the existing development and the project.

Table 15 Proposed activities for 2023/2024 reporting period

Ongoing implementation of Environmental Management Plans for the existing development and the project.

Complete installation of new hydro tip controls on top tier of the site.

Continue erosion and sediment control inspections and rectification works as necessary to manage stormwater discharge.

Update current operational management plans to reflect recommendations from findings of the annual review and any relevant monitoring results.



APPENDICIES



Appendix A – Annual Surface Water and Baseline Groundwater Condition Report



ReDirect Resource Recovery Facility, 24 Davis Road, Wetherill Park, NSW

24 September 2024

Annual Surface Water and Baseline Groundwater Condition Report – 2024





Document Information

Annual Surface Water and Baseline Groundwater Condition Report – 2024, 24 Davis Road, Wetherill Park, NSW

Prepared by:

Senversa Pty Ltd

ABN: 89 132 231 380

Level 24, 1 Market St, Sydney, NSW 2000

tel:+61 2 8252 0000 www.senversa.com.au

Prepared for:

ReDirect Recycling

2 Wella Way Somersby, NSW

| Revision | Date | Author | Reviewed | Approved | Detail |
|----------|-------------------|-------------|------------|------------------|--------|
| 0 | 18 September 2024 | Bec Chapple | Emma Walsh | Andrei Woinarski | Rev0 |
| 1 | 24 September 2024 | Bec Chapple | Emma Walsh | Andrei Woinarski | Rev1 |

Project Manager: Bec Chapple
Project Director: Emma Walsh

Disclaimer and Limitations:

This document is confidential and has been prepared by Senversa for use only by its client and for the specific purpose described in our proposal which is subject to limitations. No party other than Senversa's client may rely on this document without the prior written consent of Senversa, and no responsibility is accepted for any damages suffered by any third party arising from decisions or actions based on this document. Matters of possible interest to third parties may not have been specifically addressed for the purposes of preparing this document and the use of professional judgement for the purposes of Senversa's work means that matters may have existed that would have been assessed differently on behalf of third parties.

Senversa prepared this document in a manner consistent with the level of care and skill ordinarily exercised by members of Senversa's profession practising in the same locality under similar circumstances at the time the services were performed.

Permission should be sought before any reference (written or otherwise) is made public that identifies any people, person, address or location named within or involved in the preparation of this report. Senversa requires that this document be considered only in its entirety and reserves the right to amend this report if further information becomes available. This document is issued subject to the technical principles, limitations and assumptions provided herein in **Section 5.0**.

©2024 Senversa Pty Ltd

Senversa acknowledges the traditional custodians of the land on which this work was created and pay our respect to Elders past and present.



Executive Summary

Senversa Pty Ltd (Senversa) was engaged by reDirect Recycling Pty Ltd (reDirect) to conduct four biannual groundwater and surface water monitoring events (WME), along with associated reporting, over a two-year period at the reDirect Resource Recovery Facility located at 24 Davis Road, Wetherill Park, New South Wales (NSW) (the site).

The Water Management Plan (WMP) for the site (Senversa, 2022. *Water Management Plan, reDirect Resource Recovery Facility* – 24 Davis Road, Wetherill Park, NSW) outlines the initial requirement for six-monthly (bi-annual) monitoring of surface water and groundwater across and under the site for a two-year period following commencement of operations (August 2022).

This report documents the surface water monitoring conducted in February and July 2024, as well as the baseline groundwater conditions established over a two-year period from February 2023 to July 2024.

Objectives

The objectives of surface water and groundwater monitoring were to:

- Comply with the requirements of the WMP.
- Verify whether surface water/stormwater controls are adequately maintained and performing to meet the performance targets set out in the State Significant Development (SSD) Conditions of Approval (COA) and Fairfield City Council (FCC) (2017) Stormwater Management Policy.
- Assess surface water/stormwater quality with respect to the conditions of the site's environmental protection licence (EPL 21092).
- Assess geochemical parameters and target analytes in groundwater to form a baseline characterisation of the groundwater on site.

Scope of Work

The scope of work was in accordance with the WMP and included the following:

- Ongoing inspections by reDirect of the site areas outside of the covered and controlled processing areas (e.g. driveway, car park area, ramp) and all surface water sampling points and subsurface drains.
- Six-monthly (following a rainfall event) sampling and analysis of two surface water sampling points on-site (SW1 in the sand filter and SW2 in the Ecoceptor outflow sampling point).
- Six-monthly gauging, sampling and analysis of five on-site groundwater monitoring wells (MW1, MW2, MW3, MW4 and MW6).
- Preparation of this report.



Conclusions

Based on the available data and with respect to the objectives, the following conclusions are made:

Compliance with WMP:

Surface water monitoring was conducted consistent with requirements in the WMP during 2024.

Groundwater monitoring was generally conducted consistent with requirements in the WMP during 2023 and 2024, with no material deviations that were considered to impact the outcomes of the assessment.

Ensure surface water/stormwater controls are adequately maintained and performing to meet the performance targets set out in the SSD COA and FCC (2017):

No repairs were identified to be required. The quarterly inspections reported that there were no outstanding factors that needed addressing during the monitoring period.

Through comparison of analytical results at SW1 (untreated stormwater) vs SW2 (treated stormwater), the stormwater treatment train met the stormwater pollutant reduction targets outlined in FCC (2017) for total suspended solids (TSS), total phosphorus (TP) and total nitrogen (TN).

Assess surface water/stormwater quality with respect to Condition L1.1 of EPL 21092:

During the 2024 stormwater monitoring events:

- Concentrations of all analytes tested in surface water were reported below the adopted healthbased assessment criteria.
- Concentrations of copper, zinc and nutrients were reported above the conservative ecological screening criteria at the Ecoceptor discharge point (SW2), indicative of the quality of stormwater being discharged from site. However, given the low concentrations reported (within the range of expected concentrations provided in literature of discharge from industrial sites), the intermittent nature of stormwater flows (rainfall dependent), mixing with downstream stormwater discharges and the distance to the nearest ecological receptor (1.5 kilometres), this is not considered to pose an unacceptable risk to the receptor or constitute a pollution event.

Assess geochemical parameters and target analytes in groundwater to form a baseline characterisation of the groundwater on site.

During the 2023 - 2024 WMEs:

- Baseline groundwater conditions were established, which can be used for assessing potential impacts to groundwater from operations at the site in the future.
- The laboratory reported either low concentrations, or concentrations below the laboratory limit of reporting, for benzene, toluene, ethylbenzene and xylene (BTEX), total recoverable hydrocarbons (TRH), polycyclic aromatic hydrocarbons (PAH) and phenols, which are the primary contaminants associated with the historical use of the site as an asphalt batching plant.
- Concentrations of analytes tested in groundwater were below the adopted health-based assessment criteria, with the exception of manganese and nickel, which are considered indicative of regional groundwater conditions.
- Concentrations of chromium, copper, manganese, nickel and zinc were reported above the
 conservative ecological screening criteria. Metal concentrations were generally highest at MW3,
 which may be indicative of the quality of groundwater migrating onto the site from the
 neighbouring property to the west and/or regional groundwater conditions. The potential risk to
 ecological receptors is considered low given the distance to the nearest receptor and the low
 hydraulic conductivity of the groundwater.

No incidents were reported that may have resulted in an impact to groundwater.



Recommendations

On the basis of the results from this investigation, Senversa recommends the following:

- reDirect undertake scheduled maintenance on the site's stormwater treatment train, including the sand filter detention pit and Ecoceptor, to remove possible sediment build-up that may be causing zinc concentrations to increase through the treatment process.
- Per the WMP, the following monitoring should continue:
 - Ongoing weekly, quarterly and biannual environmental inspections and maintenance of the stormwater system by reDirect in accordance with the site's operational environmental management plan (OEMP).
 - Annual surface water sampling (per Table 4.2 of the WMP) from both SW1 and SW2 for the following analytes: pH, TSS, TP, TN and copper & zinc.
- Although the WMP outlined that annual groundwater monitoring be conducted for three years post completion of baseline monitoring, Senversa recommends that this is no longer required. Baseline groundwater monitoring commenced at the same time as commencement of operations at the site and no detrimental statistical trends, considered to be associated with site operations, have been noted in groundwater quality during this time. It is considered unlikley that additional changes in groundwater quality would be noted after a further one year of groundwater monitoring given the sealed nature of the operational portion of the site, the low hydraulic conductivity of the underlying aquifer and adherence to the Applicant's Management and Mitigation Measures that form Appendix B of the Development Consent. Further triggers, in accordance with Table 5.2 of the WMP, for additional groundwater monitoring should include if additional processes commence at the site (e.g. food and garden organics [FGO], food and liquid depackaging [FLD] or other trackable liquid wastes), if a potentially contaminating substance is to be stored or used/processed on the site, or a major incident occurs at the site (e.g. spill or leak of liquid substance/leachate, fire, etc). A review of the requirement for groundwater monitoring should be conducted every three years and when additional processes commence at the site.
- The potential risk to intrusive maintenance workers from elevated concentrations of manganese
 and nickel in the groundwater is considered to be low, given that exposure is unlikely due to no
 identified extraction/use and the average depth to water exceeds 2 metres below ground level
 (m bgl). However, it may be prudent to reduce potential risks during any deep intrusive works via
 minimising contact with groundwater and implementing good hygiene practices. These potential
 risks and management controls should be documented in safe work method statements (SWMS).
- Nutrient concentrations should no longer be compared against the overly conservative ANZECC & ARMCANZ (2000)¹ physical stressor trigger levels. Reductions in nutrient levels during stormwater treatment should continue to be monitored in accordance with FCC (2017).

_

¹ ANZECC & ARMCANZ (2000). *Australian and New Zealand Guidelines for Fresh and Marine Water Quality*. Australian and New Zealand Environment and Conservation Council and Agriculture and Resource Council of Australia and New Zealand.



Contents

| Executiv | ve Summary | ii |
|-----------|--|-----|
| List of A | Acronyms | vii |
| 1.0 | Introduction and Objectives | 1 |
| 1.1 | Background | 1 |
| 1.2 | Objectives | 1 |
| 1.3 | Scope of Work | 2 |
| 1.4 | Deviations from WMP | 2 |
| 2.0 | Monitoring Rational and Methodology | 3 |
| 2.1 | Monitoring Locations | 3 |
| 2.2 | Sampling Methodology | 3 |
| 2.3 | Water Quality Assessment Criteria | 5 |
| 2.3.1 | Surface Water Trigger Levels and Action Responses | 5 |
| 2.3.2 | Surface Water Quality Assessment Criteria | 6 |
| 2.3.3 | Groundwater Trigger Levels and Action Responses | 6 |
| 2.3.4 | Groundwater Assessment Criteria | 7 |
| 3.0 | Results | 9 |
| 3.1 | Site Inspections | 9 |
| 3.2 | Rainfall Prior to Sampling Events | 9 |
| 3.3 | Surface Water | 9 |
| 3.3.1 | Observations and Geochemical Parameters | 9 |
| 3.3.2 | Analytical Results | 10 |
| 3.3.3 | Statistical Analysis | 12 |
| 3.4 | Groundwater | 13 |
| 3.4.1 | Observations and Geochemical Parameters | 13 |
| 3.4.2 | Analytical Results | 13 |
| 3.4.3 | Groundwater Baseline Conditions and Statistical Analysis | 16 |
| 3.4.4 | Contaminant Concentration Trends | 18 |
| 3.5 | Data Quality Review | 19 |
| 4.0 | Conclusions and Recommendations | 20 |
| 4.1 | Conclusions | 20 |
| 4.2 | Recommendations | 22 |
| 5.0 | Principles and Limitations of Investigation | 23 |
| 6.0 | References | 24 |



Tables in Text

| Table 2-1: Inspection, Surface Water and Groundwater Monitoring Methodology | 3 |
|---|----|
| Table 2-2: Stormwater Quality Triggers and Action Responses | 5 |
| Table 2-3 Groundwater Management Plan Trigger Level and Action Responses | 6 |
| Table 2-4: Groundwater Assessment Criteria | 7 |
| Table 3-1: Rainfall prior to surface water monitoring events | g |
| Table 3-2 Surface water observations and geochemical parameters | S |
| Table 3-3 Surface Water Analytical Summary | 11 |
| Table 3-4: Statistics for Surface Water Samples in 2023 and 2024 | 12 |
| Table 3-5: Geochemical Parameters of Groundwater | 13 |
| Table 3-6 Groundwater Analytical Summary | 15 |
| Table 3-7: Groundwater Statistical Analysis of Analytical Data | 17 |
| Table 3-8: Groundwater Concentration Trend Analysis Summary 2023 – 2024 | 18 |

Appendices

Figures

Tables

Appendix A: OEMP Inspections

Appendix B: Field Sheets

Appendix C: Calibration Certificates

Appendix D: Quality Assessment / Quality Control

Appendix F: Laboratory Reports

Appendix G: Mann Kendall Groundwater Trend Analysis



List of Acronyms

| Acronym | Definition |
|---------|--|
| ADWG | Australian Drinking Water Guidelines |
| ALS | Australian Laboratory Services |
| ANZECC | Australian and New Zealand Environment and Conservation Council |
| ANZG | Australian and New Zealand Guidelines |
| ARMCANZ | Agriculture and Resource Council of Australia and New Zealand |
| ASC | Assessment of Site Contamination |
| вом | Bureau of Meteorology |
| ВТЕХ | Benzene, toluene, ethylbenzene, xylenes |
| COA | Conditions of approval |
| CoPC | Contaminant of potential concern |
| DGV | Default guideline value |
| DO | Dissolved oxygen |
| DQI | Data Quality Indicator |
| DQO | Data Quality Objective |
| EC | Electrical conductivity |
| EPA | Environment Protection Authority (NSW) |
| EPL | Environmental Protection Licence |
| FCC | Fairfield City Council |
| FGO | Food and garden organics |
| FLD | Food and liquid depackaging |
| HDPE | High-density polyethylene |
| HSL | Health screening level |
| km | Kilometre |
| LOR | Limit of reporting |
| m | Metres |
| m AHD | Metres Australian Height Datum |
| m bgl | Metres below ground level |
| m btoc | Metres below top of casing |
| mg/L | Milligrams per litre |

| Acronym | Definition |
|----------|--|
| MW | Monitoring well |
| NAPL | Non-aqueous phase liquid |
| NATA | National Association of Testing Authorities |
| NEPC | National Environment Protection Council |
| NEPM | National Environment Protection Measure |
| NHMRC | National Health and Medical Research Council |
| NSW | New South Wales |
| OEMP | Operational Environmental Management Plan |
| PAH | Polycyclic aromatic hydrocarbons |
| PCR | Primary contact recreation |
| POEO Act | Protection of the Environment Operations Act 1997 |
| QA | Quality assurance |
| QC | Quality control |
| RPD | Relative percentage difference |
| SSD | State Significant Development |
| SWL | Standing water level |
| TDS | Total dissolved solids |
| TSS | Total suspended solids |
| TKN | Total kjeldahl nitrogen |
| TN | Total nitrogen |
| TON | Total organic nitrogen |
| ТР | Total phosphorus |
| TRH | Total recoverable hydrocarbons |
| μg/L | Micrograms per litre |
| WME | Water Monitoring Event |
| WMP | Water Management Plan |



1.0 Introduction and Objectives

Senversa Pty Ltd (Senversa) was engaged by reDirect Recycling Pty Ltd (reDirect) to conduct four biannual (six-monthly) groundwater and surface water monitoring events (WME), along with associated reporting, over a two-year period at the reDirect Resource Recovery Facility located at 24 Davis Road, Wetherill Park, New South Wales (NSW) (the site). The site location and layout are presented on **Figure 1**.

Senversa (2023)² documented the surface water monitoring conducted in February and August 2023. This report documents the surface water monitoring conducted in February and July 2024, as well as the baseline groundwater conditions established over the two-year period from February 2023 to July 2024.

1.1 Background

A Water Management Plan (WMP) has previously been prepared for the site³ and is currently being implemented in accordance with the site's operational environmental protection licence (EPL) 21092, issued by the NSW Environment Protection Authority (EPA). The facility commenced operation under EPL 21092 in August 2022.

The WMP outlines the initial requirement for six-monthly (bi-annual) monitoring of surface water and groundwater across and under the site for a two-year period following commencement of operations. The requirements for subsequent on-going monitoring will be determined based on the results from the initial two-year period.

The purpose of the two-year period of surface water monitoring was to assess the quality of stormwater collected on-site prior to treatment, as well as being discharged to the Council stormwater system, to assess whether the stormwater controls across the site are meeting their performance targets. The purpose of the groundwater monitoring was to establish baseline groundwater conditions, which may be used to assess whether there are site-related operational impacts to groundwater quality in the future.

The existing monitoring network comprises five groundwater monitoring wells and two surface water sampling locations, as defined in the WMP, and shown on **Figure 2**.

The WMP requires a factual surface water and baseline groundwater condition report after the initial two-year monitoring period, which is this report.

1.2 Objectives

The objectives of surface water and groundwater monitoring were to:

- Comply with the requirements of the WMP.
- Verify whether surface water/stormwater controls are adequately maintained and performing to meet the performance targets set out in the State Significant Development (SSD) Conditions of Approval (COA) and Fairfield City Council (FCC) (2017) Stormwater Management Policy.
- Assess surface water/stormwater quality with respect to Condition L1.1 of EPL 21092.
- Assess geochemical parameters and target analytes in groundwater to form a baseline characterisation of the groundwater on site.

² Senversa (2023). Surface Water Monitoring Report – Annual 2023, ReDirect Resource Recovery Facility, 24 Davis Road, Wetherill Park, NSW. 21 September 2023.

³ Senversa (2022). Water Management Plan, reDirect Resource Recovery Facility – 24 Davis Road, Wetherill Park, NSW. 5 April 2022



1.3 Scope of Work

The scope of work was in accordance with the WMP and included the following:

- Ongoing inspections (since September 2023) by reDirect of the site areas outside of the covered
 and controlled processing areas (e.g. driveway, car park area, ramp) and all surface water sampling
 points and subsurface drains.
- Six-monthly (following a rainfall event) sampling and analysis of two surface water sampling points on-site (SW1 in the sand filter and SW2 in the Ecoceptor outflow sampling point) (since February 2024).
- Six-monthly gauging, sampling and analysis of five on-site groundwater monitoring wells (MW1, MW2, MW3, MW4 and MW6) (since February 2023).
- Preparation of this report.

1.4 Deviations from WMP

There were no material deviations from the surface water management plan requirements in the WMP.

The deviations from the groundwater management plan requirements in the WMP include the following:

- Groundwater monitoring well MW5 was lost prior to the February 2023 WME (the first of the four biannual WMEs). Gravel fill had been imported into the area as part of construction works post installation of the monitoring well, was compacted and covered MW5. ReDirect and Senversa tried several times to locate the monitoring well, including using a metal detector, but efforts were unsuccessful. Senversa concluded that this monitoring well was not critical to establishment of baseline groundwater conditions, as it was located in the centre of the site. Should future groundwater monitoring indicate that concentrations of contaminants of potential concern (CoPC) migrating off-site (to the south) have increased, then reDirect should consider installing a replacement monitoring well in the vicinity of MW5 to assist in identification and delineation of potential impacts.
- Groundwater geochemical parameters were not collected from groundwater monitoring well MW6 in the July 2024 WME. There was not enough volume of the groundwater sample (collected via Hydrasleeve®) for laboratory analysis and field measurement of geochemical parameters, therefore the laboratory analysis was prioritised. This is not considered to impact on the establishment of the baseline groundwater quality and conditions at this location or across the site, since three measurements of geochemical parameters were collected at this location during the two year period and geochemical parameters were measured at the other wells in the groundwater monitoring well network during the July 2024 WME.



2.0 Monitoring Rational and Methodology

2.1 Monitoring Locations

Monitoring locations included the following, as shown on Figure 2:

- General site areas outside of covered and controlled processing areas (e.g. driveway, car park area, ramp) (inspection only by reDirect).
- Two surface water locations:
 - SW1 Sand filter bed inflow sampling point (to assess quality of surface water across the site prior to treatment).
 - SW2 Ecoceptor outflow sampling point (to assess quality of surface water across the site following treatment and prior to discharge from site).
- Five groundwater monitoring wells:
 - MW1 Hydraulically down-gradient of the stormwater treatment sand filter box, representative
 of groundwater likely migrating off-site to the south.
 - MW2 Hydraulically down-gradient of the Ecoceptor, also representative of groundwater likely migrating off-site to the south.
 - MW3 Western site boundary, hydraulically down-gradient of neighbouring property.
 - MW4 Hydraulically down-gradient of the drill mud processing facility on eastern boundary.
 - MW6 Hydraulically up-gradient monitoring well that captures the quality of background groundwater migrating onto the site from the north.

2.2 Sampling Methodology

The inspection, surface water and groundwater assessment methodology are summarised below.

Table 2-1: Inspection, Surface Water and Groundwater Monitoring Methodology

Activity Details Each week, reDirect was responsible for conducting a site inspection in which they observed the Inspections (since Senversa, 2023) general site areas outside of covered and controlled processing areas (e.g. driveway, car park area, ramp). These records are presented in Appendix A A quarterly inspection of all surface water sampling points and subsurface drain pits was conducted in March 2024 and June 2024. This included the following methodologies: Removal of the grate and inspection of the internal walls and base. Removal of any collected sediment, debris, litter and vegetation Inspection and ensuring the grate was clear following any removal of objects. Ensuring there was a flush placement of the grate upon refitment. Drainage structures were inspected noting any dilapidation, with repairs been carried out if necessary. Rainwater tanks were checked for evidence of litter and pests and the structural integrity of the tank was assessed. The sediment chamber for the Ecoceptor was checked and cleaned, with any damages These records are also presented in Appendix A.



Activity

Details

Surface Water Sampling (since Senversa, 2023)

Surface water sampling commenced after a period of rainfall, to ensure there was enough water to sample from the sampling locations. Rainfall data was monitored prior to each surface water sampling event. The rainfall data was collected from the Australian Bureau of Meteorology (BOM), measured from Prospect Reservoir (station 067019) 1 kilometre (km) north of the site.

- Surface water sampling was completed on the following dates:
- 7 February 2024.
- 9 July 2024.

Laboratory prepared and supplied bottles/vials were filled directly from the sampling location using an extendable sampling pole. A sub-sample was filtered using a $0.45 \, \mu m$ filter in the field prior to placing into sample container for dissolved metals analysis. Vials and bottles were filled to minimise headspace and placed into an insulated cooler containing bagged ice.

A separate aliquot of water was collected for field measurement of general water quality parameters⁴.

A new pair of nitrile gloves were worn for each sample collection event.

Sampling field records are presented in Error! Reference source not found.. Calibration certificates for the equipment used during the field program are presented in **Appendix C**.

Monitoring Well Gauging

The groundwater standing water level (SWL) was measured in the monitoring wells (MW1, MW2, MW3, MW4 and MW6) using a calibrated electronic water/oil interface probe.

Groundwater gauging records are presented in **Table 1**, with field notes included in **Appendix B** and calibration certificates presented in **Appendix C**.

Groundwater Sampling

Groundwater samples were collected from monitoring wells utilising no-purge high-density polyethylene (HDPE) HydraSleeves \circledR .

The HydraSleeves® were placed a minimum of 1 metre (m) below the measured SWL and left *insitu* for a minimum of 24-hrs prior to sample collection. During sampling, collected groundwater was transferred from the HydraSleeve® directly into laboratory supplied sample containers. Geochemical parameters were measured *ex-situ* in the field using a calibrated water quality meter.

The geochemical water quality data are presented in **Table 2**, with field notes included in **Appendix B** and calibration certificates in **Appendix C**.

Sample Analytical Schedule

Samples were analysed at laboratories by methods endorsed by the National Association of Testing Authorities (NATA), including:

- ALS Environmental (ALS): analysis of primary surface water samples.
- Envirolab: Analysis of February 2023 inter-laboratory duplicate sample.
- Eurofins: Analysis of August 2023, February 2024 and August 2024 interlaboratory duplicate sample.

Surface Water

Surface water samples were analysed for constituents required by the WMP: total dissolved solids (TDS), total suspended solids (TSS), total nitrogen (TN), total phosphorus (TP), dissolved metals (arsenic, cadmium, chromium, copper, lead, mercury, nickel, zinc, iron and manganese), total recoverable hydrocarbons (TRH), polycyclic aromatic hydrocarbons (PAH) and phenols.

Groundwater

Groundwater samples were analysed for constituents required by the WMP: Ammonia (as N), nitrate, TN, TP, dissolved metals (same as for surface water), TRH, BTEX⁵ and PAH.

⁴ General water quality parameters including: pH, electrical conductivity (EC), dissolved oxygen (DO) and redox potential.

⁵ Benzene, toluene, ethylbenzene and xylenes.



Activity

Details

Quality assurance and quality control

Data quality assurance (QA) and quality control (QC) procedures consistent with the guidance in the WMP were implemented (refer **Appendix D**) including:

- Field QA procedures: Inspections were conducted by suitably experienced persons familiar
 with the site operations; water sampling was conducted by suitable trained and experienced
 persons; dedicated sampling equipment was used; field and equipment calibration records
 were retained.
- Field QC samples: The following QC samples were analysed:
 - One intra-laboratory duplicate sample per WME.
 - One inter-laboratory duplicate sample per WME.
 - One rinsate sample per WME.
 - One trip-blank was analysed in the February 2023 and August 2023 WMEs and two were analysed in each of the February 2024 and July 2024 WMEs.
 - One trip-spike was analysed in February 2023 and August 2023 WMEs and two were analysed in each of the February 2024 and July 2024 WMEs.
- Laboratory QA/QC procedures and controls were implemented refer **Appendix D**.

The data validation process involved checking both the analytical procedure compliance, as well as the accuracy and precision of the sampling methods used throughout the sampling program (refer **Appendix D**).

2.3 Water Quality Assessment Criteria

Condition L1 of the EPL states that the licensee must comply with section 120 of the *Protection of the Environment Operations Act 1997* (POEO Act), which prohibits the pollution of waters. The below subsections outline the adopted assessment criteria for surface water and groundwater to assess whether pollution of waters may have occurred.

2.3.1 Surface Water Trigger Levels and Action Responses

Stormwater trigger levels and action responses are provided in Section 4.3.5 of the WMP. Most of the monitoring tasks and maintenance actions relate to the stormwater network and devices, however the monitoring item related to stormwater quality, which informs the objectives of the stormwater monitoring program, is reproduced in **Table 2-2** below.

Table 2-2: Stormwater Quality Triggers and Action Responses

| Aspect | Trigger | Purpose of Monitoring | Action |
|---|---|---|--|
| Exceedance of water quality objectives | Condition L1 of the EPL states that the licensee must comply with section 120 of the POEO Act, which prohibits the pollution of waters. Stormwater quality should also meet FCC stormwater quality, discharge requirements or approval conditions. In the absence of any EPL or FCC criteria, site-specific risk-based screening criteria should be adopted from NSW EPA made or approved guidance appropriate for the commercial/industrial land use and heavily disturbed receiving environment. These include: ANZG (2018) Australian and New Zealand Guidelines for Fresh and Marine Water Quality for heavily disturbed environments; and, primary contact recreation (PCR) guidelines adopted from National Health and Medical Research Council (NHMRC) (2011), Australian Drinking Water Guidelines and NHMRC (2008) Guidelines for Managing Risks in Recreational Water. | Verify soil and erosion, and stormwater, management controls in SSD-7401 are performing as designed. | Review the above triggers and actions. |



2.3.2 Surface Water Quality Assessment Criteria

In the absence of any EPL or FCC criteria, the WMP adopted site-specific risk-based screening criteria from NSW EPA made or approved guidance appropriate for the commercial/industrial land use and heavily disturbed receiving environment. These include (refer **Table 3** for criteria values):

- Health risk screening: Direct contact exposure based on PCR guidelines adopted from the
 Australian Drinking Water Guidelines (ADWG, 2022)⁶ and NHMRC (2008) Guidelines for Managing
 Risks in Recreational Water for recreational exposure. This is also conservative for incidental
 exposure to workers.
- Ecological risk screening: ANZG⁷ (2018) Australian and New Zealand Guidelines for Fresh and Marine Water Quality for heavily disturbed environments. Surface water that discharges the site may migrate and discharge into Prospect Creek (approximately 1.5 km to the east of the site), which is the nearest surface water body downstream of the site, though the ultimate receiving environment is the George's River and Botany Bay (marine). The WMP indicates previous studies found that the local receiving waterways are heavily disturbed. The relevant ecological guidelines for toxicants, are therefore, the freshwater default guidelines values (DGVs) for heavily disturbed environments from ANZG (2018).
 - Given that ANZG (2018) does not specify DGVs for nutrients, the physical stressor trigger values from ANZECC & ARMCANZ (2000) have conservatively been adopted, however these do not take into consideration the surrounding land use. Given the setting in an urban environment, consideration is given to concentrations in stormwater runoff in urban or commercial/ industrial areas in east coast Australia reported by Drapper et al (2022) and Fletcher et al (2004), which are more applicable to the site setting.
- Aesthetic impacts: e.g. no gross aesthetic impacts such as non-aqueous phase liquids (NAPL).

2.3.3 Groundwater Trigger Levels and Action Responses

Trigger levels and action responses applied to the groundwater monitoring program are provided in Section 5.5 of the WMP and repeated in **Table 2-3** below.

Table 2-3 Groundwater Management Plan Trigger Level and Action Responses

| Aspect | Trigger | Actions |
|-------------|---|---|
| Groundwater | Concentrations of key indicator analytes outlined in the WMP exceed performance | Consider re-sampling or increased sampling frequency to confirm results. |
| | criteria and do not show a stable or decreasing trend. | Assess possible sources of contamination – i.e. change in site operations, change in neighbouring site operations or chemical spills. |
| | | Assess the significance of associated environmental risk – where a potentially unacceptable risk is identified, a suitably qualified and experienced professional should assess whether the monitoring program is adequate to assess potential contamination risks, and recommend program changes (if necessary) (e.g., additional sampling locations, more frequent monitoring or different CoPC). |
| | | Implement the amended monitoring program. |
| | | Develop and implement management/remedial actions if necessary. |

_

⁶ NHMRC (2011). Australian Drinking Water Guidelines, updated January 2022. National Health and Medical Research Council.

⁷ Australian and New Zealand Governments and Australian state and territory governments.



7

| Aspect | Trigger | Actions | | | |
|-----------------|--|---|--|--|--|
| | Concentrations of key indicator analytes in the WMP that are less than the performance criteria and show statistically significant stable or decreasing trend over a minimum of three events. | Assessment to determine the residual environmental risk and review the monitoring program by a suitable qualified and experienced professional. If monitoring results are consistently decreasing to levels below the performance guidelines outlined in section 5.4 of the WMP and the residual environmental risk from ongoing primary sources is considered low by a suitably qualified and experienced professional, the groundwater monitoring program may end. | | | |
| | Damaged or lost wells | Assess whether ongoing monitoring at the location is necessary. If required, repair or re-install the well. | | | |
| Site Activities | Incident (e.g. spill or release of a material or liquid) that could result in impact to surface or groundwater. | Assess whether monitoring program is adequate to assess potential impact associated with the incident. This assessment should be undertaken by a suitable qualified and experienced professional and desumented in a report with aleast conclusions. | | | |
| Site Activities | Change in nature or management of imported materials that has the potential to result in a significantly increased risk of impact from leachate. Including commencement of Stage 2 operations. | professional and documented in a report with clear conclusions and recommendations for amendments (if necessary). Implement program changes – these may include increased monitoring frequency, inclusion of additional monitoring locations installation and monitoring of additional wells, broader analytical suite to assess the chemicals of concern. | | | |

2.3.4 Groundwater Assessment Criteria

Assessment of groundwater quality in the future will principally be via comparison against baseline and site background conditions, which have been established in this report. **Table 2-4** below summarises the groundwater quality criteria (or performance criteria) that are adopted in the WMP.

Table 2-4: Groundwater Assessment Criteria

| Receptor | Adopted Assessment Criteria |
|--|---|
| Change to baseline / background conditions | No statistically significant increasing trend or 20% increase over baseline / background concentrations or field parameters. |
| Human Health | Relevant criteria for the commercial/industrial land use setting have been adopted as a screening levels. This includes: |
| | Direct contact criteria (same as for surface water, based on guidelines adopted from ADWG (2022) and NHMRC (2008)) have been considered due to the relatively shallow depth to groundwater in some locations, which may be encountered during intrusive (sub-surface) construction/maintenance works. The presence of concrete and asphalt hardstand however indicates that groundwater will be predominantly inaccessible to humans. |
| | Health Screening Level for commercial/industrial land use (HSL-D) in the ASC NEPM⁸ for assessment of vapour intrusion risks. Concentrations above the HSLs may pose a risk to users of enclosed buildings. |
| | No gross aesthetic impacts such as NAPL. |
| | Drinking water guidelines will not be considered, given the site geology, land use and provision of a reticulated drinking water supply. |
| Ecological | Relevant criteria include: |
| | ANZG (2018) freshwater DGVs for heavily disturbed environments, on the basis that groundwater may migrate and discharge into Prospect Creek. |

S20102_006_RPT_rev1 | Annual Surface Water and Baseline Groundwater Condition Report – 2024

⁸ NEPC (2013). *National Environment Protection (Assessment of Site Contamination) Amendment Measure (No.1)*. National Environment Protection Council.



ANZG (2018) notes that exceedance of a DGV does not necessarily imply that there is an inherent risk, rather that further assessment and monitoring may be required prior to implementing appropriate management actions. These adopted assessment criteria should be used for screening purposes to trigger further assessment, rather than to directly assess the level of risk to any identified receptors.



3.0 Results

3.1 Site Inspections

The following key observations were made by reDirect during quarterly inspections of the surface water sampling points and subsurface drain pits in March 2024 and June 2024:

- The grates were cleaned when observations indicated that sediment was present and debris was removed when necessary.
- The sediment chamber of the Ecoceptor was checked during each quarterly observation, with no further action required on each occasion.
- No repairs were required for the surface water and stormwater drain structures.
- The rainwater tank was clear of pests and debris on each occasion, with no repairs required.

3.2 Rainfall Prior to Sampling Events

The surface water sampling events were targeted to follow a rainfall event to maximise the potential for a sufficient volume of water (for sampling and analysis) to have been treated and discharged from the Ecoceptor. The following rainfall data was collected from the BOM Prospect Reservoir (station 067019). **Table 3-1** outlines the rainfall that occurred in the 3-day period prior to each monitoring event.

Table 3-1: Rainfall prior to surface water monitoring events

| Date | 24-hour Rainfall (including day of sampling) | 3-Day Rainfall (including day of sampling) |
|-----------------|---|---|
| 7 February 2024 | 22 millimetres (mm) | 22 mm |
| 9 July 2024 | 2 mm | 6 mm |

3.3 Surface Water

3.3.1 Observations and Geochemical Parameters

The field-measured surface water geochemical parameters for the sampling events are presented in the table below.

Table 3-2 Surface water observations and geochemical parameters

| Location | Event | Dissolved Oxygen (mg/L) | Electrical Conductivity (μs/cm) | рН | Redox (Eh) (mV) | Temperature (°C) | Observations |
|-----------------------------|------------------|-------------------------------|---------------------------------------|------|-----------------------|---------------------|--|
| SW1 (untreated water) | February 2024 | 3.07 | 574 | 7.51 | 307 | 23.9 | Colourless, no odour, no sheen, suspended sediments |
| | July 2024 | 1.06 | 334 | 7.08 | 243 | 13.6 | Light grey brown, no odour, no sheen, moderately turbid |



| Location | Event | Dissolved Oxygen (mg/L) | Electrical Conductivity (μs/cm) | рН | Redox (Eh) (mV) | Temperature (°C) | Observations |
|---------------------------|------------------|-------------------------------|---------------------------------------|------|-----------------------|---------------------|--|
| SW2 (treated water) | February 2024 | 4.19 | 656 | 7.70 | 286 | 25.0 | Colourless, no odour, no sheen, suspended sediments |
| | July 2024 | 6.09 | 370 | 8.48 | 187 | 13.7 | Light grey, no odour, no sheen, slightly turbid |

Based on the above information, the following key observations were made:

- The DO of the treated water (SW2) was higher than the untreated water (SW1), likely due to the flow and agitation of the water through the stormwater treatment process. The stormwater being released from the site at the time of sampling (based on the results from SW2) was aerobic.
- The EC measurements at both SW1 and SW2 were indicative of freshwater. This indicates that the EC of the rain falling across the site had not been significantly altered by conditions at the site by the time the rain was discharged from the site (as stormwater).
- The measured pH was indicative of neutral to moderately alkaline conditions, which is within the adopted acceptable range.
- The measured Eh of SW1 and SW2 was indicative of slightly to moderately oxidising conditions.
- The range of measured temperatures is reflective of seasonal changes.
- The sampled untreated (SW1) and treated (SW2) stormwater did not have any visual / olfactory indicators of contamination.

3.3.2 Analytical Results

The surface water analytical results and screening against adopted assessment criteria are provided in **Table 3**. The laboratory analysis reports (**Appendix E**) contain the laboratory analytical results. The surface water exceedances of site criteria are also presented on **Figure 4** (attached).

A summary of exceedances of water quality objectives is provided in the table below.



Table 3-3 Surface Water Analytical Summary

| Analyte / Value | Screening Criter | ia Exceedances | | Comment | | | |
|--------------------------------|------------------|--|-----------------|--|--|--|--|
| | Health-Risk | Ecological Risk | Aesthetics | | | | |
| Heavy metals and metalloids | None identified | Heavy metal concentrations were reported at low levels, less than relevant screening criteria for highly disturbed environments, with the exceptions of: Copper (at SW1 and SW2 in February 2024 and July 2024). Zinc (at SW2 in July 2024). | - | Metal concentrations were generally less than or similar to relevant screening criteria for disturbed ecosystems consistent with the WMP. Reported copper concentrations were slightly elevated above ecological screening criteria at both SW1 (untreated) and SW2 (treated), indicating that the treatment train has not reduced copper concentrations in the stormwater. The reported zinc concentrations at SW2 in February 2024 and July 2024 were higher than reported in SW1. This scenario was also noted in August 2023. The cause for the increase in zinc concentrations is unclear, but may be related to the treatment train and should be reviewed. The average concentration of zinc in SW2 over the four sampling events in 2023 and 2024 (0.02 mg/L) was also above the ecological criterion. | | | |
| Nutrients | None identified | No exceedances for toxicants. Exceedances of conservative physical stressor values for total oxidised nitrogen (as N), TN and TP at SW1 and SW2 in both February 2024 and July 2024. | - | Concentrations are similar to the median values for TN (1.7 mg/L) and TP (0.31 mg/L) in stormwater runoff in urban or commercial/ industrial areas in east coast Australia reported by Drapper et al (2022) and Fletcher et al (2004). TN concentrations are generally less than DGV (90% spp) for nitrate (as N) (5.6 mg/L) The concentrations of TN and TP were lower in SW2 (treated) than in SW1 (untreated). This is further discussed below. | | | |
| Organic CoPC | None identified | Exceedance of TRH >C10-C16 fraction minus naphthalene (F2) at SW1 in February 2024. | None identified | BTEX, PAHs and phenols were not detected in water samples. TRH >C10-C16 fraction minus naphthalene (F2) exceeded the ecological criteria at SW1 in February 2024, but was reported below the laboratory limit of reporting (LOR) in SW2, indicating that the treatment train was effective in removing the petroleum contamination. The concentrations of TRH in both SW1 and SW2 were below the LOR in July 2024, potentially indicating that there may have been an unreported or undetected spill or leak of petroleum products during February 2024. | | | |
| Physico-chemical Parameters | None identified | None identified | None identified | The TSS was almost two orders of magnitude greater in July 2024 compared to February 2024 at SW1, which may be related to the rainfall preceding both events. The concentrations reported in SW2 were lower (by at least one order of magnitude), indicating that the treatment train is effective in reducing the TSS concentration under a range of turbidity conditions. | | | |

3.3.3 Statistical Analysis

The minimum concentration, maximum concentration and mean concentrations of the main CoPCs is presented in the table below, with exceedances of the site assessment criteria outlined.

Table 3-4: Statistics for Surface Water Samples in 2023 and 2024.

| Analyte | | Criteria | | Concentra | Concentration Reported in 2023 and 2024 Monitoring Events | | | | | | |
|---|--------|--------------|-----------------------|--|---|---------|---|---|---------------------|--|--|
| (units) | Health | Ecological | Physical Stressors | | SW1 | | | SW2 | | | |
| | | | | Min | Max | Average | Min | Max | Average | | |
| TSS (mg/L) | - | = | - | 86 | 7,260 | 1,919 | 8 | 90 | 52 | | |
| Copper (mg/L) | 20 | 0.0018 | - | 0.003 | 0.006 | 0.004 | <u>0.001</u> | 0.004 | 0.003 | | |
| Zinc (mg/L) | 60 | <u>0.015</u> | - | <0.005 | 0.005 | <0.005 | <0.005 | 0.038 | 0.020 | | |
| Total Oxidised Nitrogen (as N) (mg/L) | | - | 0.04 | 0.32 | 0.68 | 0.44 | 0.28 | 0.62 | 0.48 | | |
| Total Nitrogen (as N) (mg/L) | - | - | 0.35 | 0.7 | 22.9 | 7.0 | 1.2 | 4.3 | 2.1 | | |
| Phosphorus (as P) (mg/L) | - | - | 0.025 | 0.06 | 6.68 | 1.80 | 0.03 | 0.63 | 0.24 | | |
| >C10-C16 Fraction minus naphthalene (F2) (µg/L) | 900 | <u>440</u> | - | <lor< th=""><th><u>630</u></th><th>232</th><th><lor< th=""><th><lor< th=""><th><lor< th=""></lor<></th></lor<></th></lor<></th></lor<> | <u>630</u> | 232 | <lor< th=""><th><lor< th=""><th><lor< th=""></lor<></th></lor<></th></lor<> | <lor< th=""><th><lor< th=""></lor<></th></lor<> | <lor< th=""></lor<> | | |

Based on the statistics provided in **Table 3-4**, there are indications that concentrations of key parameters (TSS, TN and TP) are lower at SW2 (downstream of treatment train) than SW1 (upstream of system), which was also noted during the 2023 sampling events. The water, sediment and erosion controls in the WMP should continue to be followed to minimise migration of sediments and fines into the stormwater system.

3.4 Groundwater

3.4.1 Observations and Geochemical Parameters

The field-measured groundwater geochemical parameters for the sampling events are discussed in the table below.

Table 3-5: Geochemical Parameters of Groundwater

| Parameter | Minimum | Maximum | Average | Comment |
|--|---------|---------|---------|--|
| Depth to Groundwater (metres below top of casing – m btoc) | 1.9 | 4.7 | 3.0 | The depth to groundwater is generally shallower under the southern portion of the site compared to under the northern portion of the site. The top of well casing elevations have not been surveyed and therefore the standing water level (SWL) in m AHD ⁹ has not been able to be calculated. It is anticipated that groundwater flows in a south-easterly direction based on previous reports and local hydrogeological features – this is shown on Figure 2 . |
| Dissolved Oxygen (mg/L) | 0.0 | 6.3 | 2.0 | The groundwater conditions have ranged from anoxic to moderately aerobic both spatially and temporally. |
| Electrical Conductivity (μS/cm) | 1,362 | 34,645 | 19,099 | The groundwater conditions ranged from fresh under the northern portion of the site (MW6) to saline under the southern portion of the site. A previous report prepared for the site ¹⁰ indicated that the high salinity of the groundwater is associated with the Bringelly Shales underlying the site. |
| Redox Potential | 146 | 359 | 246 | The groundwater conditions have ranged from mildly to moderate oxidising conditions both spatially and temporally. |
| рН | 5.78 | 7.41 | 6.52 | The pH of the groundwater was slightly acidic to neutral and within the adopted acceptable range. |
| Temperature | 15.9 | 29.6 | 20.5 | A regular seasonal pattern was observed with warmer temperatures in summer (February 2024) and cooler temperatures in winter (July 2024). |

The variability noted above was also noted in Douglas Partners (2016).

3.4.2 Analytical Results

The groundwater sample analytical results and screening against adopted assessment criteria are provided in **Table 4**. The laboratory analysis reports (**Appendix E**) contain all analysis results. The groundwater exceedances of site criteria are also presented in **Figure 4** (attached).

⁹ Metres Australian Height Datum

¹⁰ Douglas Partners (2016). Report on Groundwater Assessment, Proposed Resource Recovery & Recycling Centre, 24 Davis Road, Wetherill Park. September 2016.

A piper plot has been generated (**Exhibit 3-1** below) to show the major ionic water constituents. The plot indicates that the hydrochemistry of the groundwater in MW6 is distinctly different from groundwater at the remaining monitoring wells, which are dominated by sodium chloride (which is also demonstrated through high EC). This may indicate that MW6 intersects a different aquifer to the remaining monitoring wells, which is supported by the following observations:

- The recharge of groundwater following purging (during well development) was noted to be higher in MW6 than in the other monitoring wells.
- The monitoring well screen at MW6 intersects a lens of sand above the natural clay, which was not observed in the other locations.

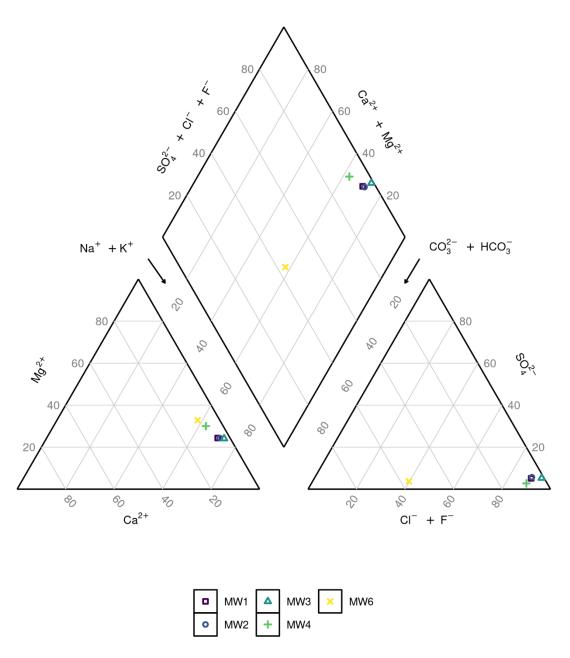


Exhibit 3-1: Piper Plot showing Hydrochemistry of Site Groundwater



A summary of exceedances of water quality objectives is provided in the table below.

Table 3-6 Groundwater Analytical Summary

| Analyte / Value | Health-Risk | Ecological Risk | Comment | | | |
|-----------------------------|---|---|---|--|--|--|
| Heavy metals and metalloids | Manganese concentrations exceeded the recreation health assessment criteria in MW3 and MW4. The nickel concentration at MW3 also exceeded the recreation health assessment criteria in one sampling event (August 2023). | *************************************** | close to or marginally exceeded the recreation health assessment criteria. It is likely that the concentration of these metals are elevated naturally and indicative of regional groundwater conditions. | | | |
| Nutrients | None identified | No exceedances for toxicants. | - | | | |
| Organic CoPC | None identified | None identified | PAHs and phenols were not detected above the LOR in the groundwater samples. One sample (MW4 in February 2024) reported a very low concentration of toluene (below assessment criteria). All other BTEX concentrations were reported below the LOR in other locations and in other sampling events. Low concentrations of semi-volatile TRH (below assessment criteria) were reported in MW1 and MW3. The detections of toluene and TRH may be associated with the former use of the site as an asphalt batching plant. | | | |

3.4.3 Groundwater Baseline Conditions and Statistical Analysis

The analytical results for the groundwater samples are presented in **Table 4**, with exceedances of the adopted site assessment criteria displayed on **Figure 3**. **Table 3-7** below outlines the minimum, maximum and mean concentration of key CoPCs in each groundwater monitoring well during the four biannual sampling events, thus establishing the baseline groundwater conditions. Additional statistics are presented in **Appendix F**.



Table 3-7: Groundwater Statistical Analysis of Analytical Data

| | | Arsenic | Chromium | Copper | Manganese | Nickel | Zinc | Ammonia (as N) | Nitrate (as N) | ВТЕХ | TRH (C6-C10) | TRH (>C10-C40) | PAH / Phenol |
|----------|-----------|---------|----------|--------|-------------|--------------|--------------|-------------------|-------------------|-------------------------|-------------------------|-------------------|---------------------|
| Criteria | Eco. | 0.042 | 0.0033 | 0.0018 | <u>2.5</u> | 0.013 | 0.015 | <u>1.43</u> | 3.8 | <u>0.11^a</u> | <u>0.44^b</u> | <u>0.64°</u> | <u>_d</u> |
| (mg/L) | Health | 0.1 | 0.5 | 20 | 5 | 0.2 | 60 | - | 110 | 0.01ª | 0.9 ^b | 0.9° | <u>_d</u> |
| Well ID | Statistic | | | | | | | | | | | | |
| MW1 | Min | 0.007 | <0.001 | <0.001 | 0.59 | <u>0.015</u> | 0.012 | 0.19 | <0.01 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Max | 0.012 | <0.001 | 0.015 | 4.84 | <u>0.156</u> | <u>0.174</u> | 0.71 | 0.02 | <0.001 | <0.02 | 0.52 | <lor< td=""></lor<> |
| | Mean | 0.010 | <0.001 | 0.005 | 2.15 | 0.058 | 0.062 | 0.47 | 0.01 | <0.001 | <0.02 | 0.2 | <lor< td=""></lor<> |
| MW2 | Min | 0.004 | <0.001 | <0.001 | 0.96 | 0.005 | <0.005 | 0.26 | <0.01 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Max | 0.008 | <0.001 | 0.011 | 3.28 | 0.006 | 0.009 | 0.52 | 0.03 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Mean | 0.005 | <0.001 | 0.003 | 1.75 | 0.006 | 0.007 | 0.44 | 0.02 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| MW3 | Min | 0.002 | <0.001 | <0.010 | <u>6.15</u> | <u>0.191</u> | 0.122 | 0.22 | <0.01 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Max | 0.011 | 0.005 | 0.006 | 7.4 | 0.207 | 0.253 | 0.29 | 0.10 | <0.001 | <0.02 | 0.91 | <lor< td=""></lor<> |
| | Mean | 0.005 | 0.004 | 0.003 | <u>6.8</u> | 0.199 | 0.214 | 0.27 | 0.03 | <0.001 | <0.02 | 0.31 | <lor< td=""></lor<> |
| MW4 | Min | 0.005 | <0.001 | <0.001 | 4.00 | 0.011 | <0.005 | 0.28 | <0.01 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Max | 0.008 | <0.001 | 0.005 | 6.04 | 0.021 | 0.006 | 0.34 | 0.01 | 0.002 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Mean | 0.007 | <0.001 | 0.002 | <u>5.13</u> | 0.017 | 0.002 | 0.31 | <0.01 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| MW6 | Min | <0.001 | <0.001 | <0.001 | <0.01 | <0.001 | <0.005 | <0.01 | 0.18 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Max | 0.002 | <0.001 | 0.003 | 0.225 | 0.002 | 0.006 | 0.09 | 1.93 | <0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |
| | Mean | 0.002 | <0.001 | 0.001 | 0.81 | 0.001 | 0.002 | 0.03 | 1.19 | < 0.001 | <0.02 | <0.1 | <lor< td=""></lor<> |

^a Most conservative assessment criteria for BTEX displayed. Table 4 presents the applicable criteria for each BTEX compound.

^b Assessment criteria for TRH C6-C10 fraction minus BTEX (F1) displayed.

^c Adopted assessment criteria for TRH >C34-C40 displayed. Table 4 presents the applicable criteria for each TRH fraction.

^d Table 4 presents the adopted assessment criteria for each PAH/phenol compound, where available.



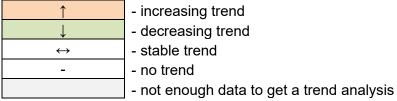
3.4.4 Contaminant Concentration Trends

Statistical evaluation of concentration trends and plots for key indicator analytes for available data from the baseline investigation (February 2023 to August 2024) are presented in **Appendix F** and summarised in **Table 3-8** below.

Table 3-8: Groundwater Concentration Trend Analysis Summary 2023 – 2024

| Chemical Name / Well ID | Arsenic | Chromium | Copper | Manganese | Nickel | Zinc | Ammonia (as N) | Nitrate (as N) | втех | TRH (C6-C10) | TRH >C10-C40 | Sum of PAH |
|----------------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| MW1 | \leftrightarrow | \leftrightarrow | - | \leftrightarrow | - | - | ↓ | - | \leftrightarrow | \leftrightarrow | - | \leftrightarrow |
| MW2 | \leftrightarrow | \leftrightarrow | - | ↑ | \leftrightarrow |
| MW3 | \leftrightarrow | \leftrightarrow | \leftrightarrow | 1 | \leftrightarrow | \leftrightarrow | \leftrightarrow | - | \leftrightarrow | \leftrightarrow | - | \leftrightarrow |
| MW4 | \leftrightarrow | \leftrightarrow | - | \leftrightarrow | ↓ | \leftrightarrow | \ | | \leftrightarrow | \leftrightarrow | \leftrightarrow | \leftrightarrow |
| MW6 | \leftrightarrow | \leftrightarrow | \leftrightarrow | - | \leftrightarrow | \leftrightarrow | - | \leftrightarrow | \leftrightarrow | \leftrightarrow | \leftrightarrow | \leftrightarrow |

Symbol



Note: If all results were <LOR, the Mann-Kendall trend was listed as "Stable"

3.5 Data Quality Review

A review of the sampling and laboratory QA/QC data completed by Senversa is presented in **Appendix D**. The QA/QC review indicated that results are generally within the relevant data quality indicator acceptance criteria for the analyses conducted and that any identified non-conformances are unlikely to affect the suitability of the data set for the purposes of this investigation. The quality of the analytical data is considered reliable for the purpose of this investigation.

4.0 Conclusions and Recommendations

4.1 Conclusions

Based on the available data and with respect to the objectives, the following conclusions are made:

Compliance with WMP:

Surface water monitoring was conducted consistent with requirements in the WMP during 2024.

Groundwater monitoring was conducted consistent with requirements in the WMP during 2023 and 2024, with the following exceptions:

- Groundwater monitoring well MW5 could not be found prior to the February 2023 WME, nor
 afterwards despite several attempts to locate the well. This monitoring well was therefore not
 sampled in any of the four biannual sampling events.
- Groundwater geochemical parameters were not collected from groundwater monitoring well MW6 in the July 2024 WME due to an insufficient volume of groundwater sample obtained.

Ensure surface water/stormwater controls are adequately maintained and performing to meet the performance targets set out in the SSD COA and FCC (2017) Stormwater Management Policy:

No repairs were identified to be required. The quarterly inspections reported that there were no outstanding factors that needed addressing during the monitoring period.

Although TRH was reported in surface water sample SW1 in February 2024, reDirect had not reported any spills that had been left unattended during that period. The treatment train appeared to be effective in removing the TRH contamination, as the laboratory did not report any TRH above the LOR in the stormwater sample collected from the outflow of the Ecoceptor (SW2), which is indicative of the quality of stormwater being discharged from the site.

Through comparison of analytical results at SW1 (untreated stormwater) vs SW2 (treated stormwater), the stormwater treatment train was effective at reducing the following contaminants / pollutants, thus meeting the stormwater pollutant reduction targets outlined in FCC (2017):

- TSS concentrations were reduced by 91 99% (FCC target of 80%).
- TP concentrations were reduced by 70 91% (FCC target of 55%).
- TN concentrations were reduced by 43 81% (FCC target of 40%).

Assess surface water/stormwater quality with respect to Condition L1.1 of EPL 21092:

During the 2024 stormwater monitoring events:

- Concentrations of all analytes tested in surface water were reported below the adopted healthbased assessment criteria.
- Concentrations of copper, zinc and nutrients were reported above the conservative ecological
 screening criteria at the Ecoceptor discharge point (SW2), indicative of the quality of stormwater
 being discharged from site. Given the intermittent nature of stormwater flows (rainfall dependent),
 mixing with downstream stormwater discharges and distance to the nearest ecological receptor
 (1.5 km), this is not considered to pose an unacceptable risk to the receptor or constitute a
 pollution event. In addition, the following is noted in regards to the concentrations reported:
 - Copper concentrations (0.004 mg/L) marginally exceeded the ANZG (2018) DGV (0.0018 mg/L). The concentrations are below the range provided in Fletcher et al (2004) for expected copper concentrations in stormwater from industrial land uses (0.02 0.3 mg/L during wet weather events).

- Zinc concentrations in stormwater increased following treatment (reported up to 0.023 mg/L) and were slightly above the adopted ecological assessment criteria at the end of the treatment process (0.015 mg/L). The concentrations are below the range provided in Fletcher et al (2004) for expected zinc concentrations in stormwater from industrial land uses (0.1 1 mg/L during wet weather events).
- Nutrients (total oxidised nitrogen, TN and TP) were an order of magnitude above the conservative ANZECC & ARMCANZ (2000) physical stressor trigger levels. These trigger levels are overly conservative for urban environments and are not cited in the more recent ANZG (2018) guidelines, so have been used here purely for screening purposes. The TN and TP concentrations were reduced following treatment, thus meeting the requirements of FCC (2017), and reported concentrations were also within the range provided in Fletcher et al (2004) for expected concentrations in stormwater from industrial land uses during wet weather events:
 - TN concentrations at SW2 (1.2 4.3 mg/L) were within the range of 0.7 6 mg/L in the literature.
 - TP concentrations at SW2 (0.03 0.63 mg/L) were within the range of 0.08 0.8 mg/L in the literature.

Assess geochemical parameters and target analytes in groundwater to form a baseline characterisation of the groundwater on site.

During the 2023 – 2024 monitoring events:

- Baseline groundwater conditions were established, which can be used for assessing potential impacts to groundwater from operations at the site in the future.
- The laboratory reported either low concentrations, or concentrations below the LOR, for BTEX, TRH, PAH and phenols, which are the primary contaminants associated with the historical use of the site as an asphalt batching plant.
- Concentrations of all analytes tested in groundwater were below the adopted health-based assessment criteria, with the following exceptions, which are considered indicative of regional groundwater conditions:
 - Manganese in MW3 and MW4; and
 - Nickel in MW3.
- Concentrations of chromium, copper, manganese, nickel and zinc were reported above the
 conservative ecological screening criteria. Metal concentrations were generally highest at MW3,
 which may be indicative of the quality of groundwater migrating onto the site from the
 neighbouring property to the west and/or regional groundwater quality. The potential risk to
 ecological receptors is considered low given the distance to the nearest receptor and the low
 hydraulic conductivity of the groundwater.

In relation to the groundwater trigger levels:

- The concentrations of the analytes that exceeded the assessment criteria in groundwater (chromium, copper, manganese, nickel and zinc) showed either no trend, a stable trend or a decreasing trend, with the exception of manganese at MW2 and MW3. Given that manganese concentrations are considered to be naturally elevated in this area, the increasing trend may be attributed to increased rainfall or standing water levels (SWLs) in the local area facilitating leaching of manganese from the natural soils, or another external factor not associated with site operations. No management or remedial actions are considered necessary.
- Concentrations of BTEX, TRH (C6-C10), PAHs and phenols were below the assessment criteria
 and showed a statistically significant stable trend, and therefore can be removed from the
 analytical program going forward.

- Although groundwater monitoring well MW5 was lost, Senversa concluded that this monitoring
 well was not critical to establishment of baseline groundwater conditions, as it was located in the
 centre of the site. Should future groundwater monitoring indicate that concentrations of
 contaminants of concern migrating off-site (to the south) have increased, then reDirect should
 consider installing a replacement monitoring well in the vicinity of MW5 to assist in identification
 and delineation of potential impacts.
- No incidents were reported that may have resulted in an impact to groundwater.

4.2 Recommendations

On the basis of the results from this investigation, Senversa recommends the following:

- reDirect undertake scheduled maintenance on the site's stormwater treatment train, including the sand filter detention pit and Ecoceptor, to remove possible sediment build-up that may be causing zinc concentrations to increase through the treatment process.
- Per the WMP, the following monitoring should continue:
 - Ongoing weekly, quarterly and biannual environmental inspections and maintenance of the stormwater system by reDirect in accordance with the site's operational environmental management plan (OEMP).
 - Annual surface water sampling (per Table 4.2 of the WMP) from both SW1 and SW2 for the following analytes:
 - pH.
 - TSS.
 - TP.
 - TN.
 - Copper and Zinc.
- Although the WMP outlined that annual groundwater monitoring be conducted for three years post completion of baseline monitoring, Senversa recommends that this is no longer required. Baseline groundwater monitoring commenced at the same time as commencement of operations at the site and no detrimental statistical trends, considered to be associated with site operations, have been noted in groundwater quality during this time. It is considered unlikely that additional changes in groundwater quality would be noted after a further one year of groundwater monitoring given the sealed nature of the operational portion of the site, the low hydraulic conductivity of the underlying aquifer and adherence to the Applicant's Management and Mitigation Measures that form Appendix B of the Development Consent. Further triggers, in accordance with Table 5.2 of the WMP, for additional groundwater monitoring should include if additional processes commence at the site (e.g. food and garden organics [FGO], food and liquid depackaging [FLD], or other trackable liquid waste), if a potentially contaminating substance is to be stored or used/processed on the site, or a major incident occurs at the site (e.g. spill or leak of liquid substance/leachate, fire, etc). A review of the requirement for groundwater monitoring should be conducted every three years and when additional processes commence at the site.
- The potential risk to intrusive maintenance workers from elevated concentrations of manganese
 and nickel in the groundwater is considered to be low, given that exposure is unlikely due to no
 identified extraction/use and the average depth to water exceeds 2 metres below ground level
 (m bgl). However, it may be prudent to reduce potential risks during any deep intrusive works via
 minimising contact with groundwater and implementing good hygiene practices. These potential
 risks and management controls should be documented in safe work method statements (SWMS).
- Nutrient concentrations should no longer be compared against the overly conservative ANZECC & ARMCANZ (2000) physical stressor trigger levels. Reductions in nutrient levels during stormwater treatment should continue to be monitored in accordance with FCC (2017).

5.0 Principles and Limitations of Investigation

The following principles are an integral part of site contamination assessment practices and are intended to be referred to in resolving any ambiguity or exercising such discretion as is accorded the user or site assessor.

| Area | Uncertainties and Limitations |
|------------------------------------|--|
| Elimination of Uncertainty | Some uncertainty is inherent in all site investigations. Furthermore, any sample, either surface or subsurface, taken for chemical testing may or may not be representative of a larger population or area. Professional judgment and interpretation are inherent in the process, and even when exercised in accordance with objective scientific principles, uncertainty is inevitable. Additional assessment beyond that which was reasonably undertaken may reduce the uncertainty. |
| Failure to Detect | Even when site investigation work is executed competently and in accordance with the appropriate Australian guidance, such as the National Environment Protection (Assessment of Site Contamination) Amendment Measure ('the NEPM'), it must be recognised that certain conditions present especially difficult target analyte detection problems. Such conditions may include, but are not limited to, complex geological settings, unusual or generally poorly understood behaviour and fate characteristics of certain substances, complex, discontinuous, random, or heterogeneous distributions of existing target analytes, physical impediments to investigation imposed by the location of services, structures and other man-made objects, and the inherent limitations of assessment technologies. |
| Limitations of Information | The effectiveness of any site investigation may be compromised by limitations or defects in the information used to define the objectives and scope of the investigation, including inability to obtain information concerning historic site uses or prior site assessment activities despite the efforts of the user and assessor to obtain such information. |
| Chemical Analysis Error | Chemical testing methods have inherent uncertainties and limitations. Senversa routinely seeks to require the laboratory to report any potential or actual problems experienced, or non-routine events which may have occurred during the testing, so that such problems can be considered in evaluating the data. |
| Level of Assessment | The investigation herein should not be considered to be an exhaustive assessment of environmental conditions on a property. There is a point at which the effort of information obtained and the time required to obtain it outweigh the benefit of the information gained and, in the context of private transactions and contractual responsibilities, may become a material detriment to the orderly conduct of business. If the presence of target analytes is confirmed on a property, the extent of further assessment is a function of the degree of confidence required and the degree of uncertainty acceptable in relation to the objectives of the assessment. |
| Comparison with Subsequent Inquiry | The justification and adequacy of the investigation findings in light of the findings of a subsequent inquiry should be evaluated based on the reasonableness of judgments made at the time and under the circumstances in which they were made. |
| Data Useability | Investigation data generally only represent the site conditions at the time the data were generated. Therefore, the usability of data collected as part of this investigation may have a finite lifetime depending on the application and use being made of the data. In all respects, a future reader of this report should evaluate whether previously generated data are appropriate for any subsequent use beyond the original purpose for which they were collected or are otherwise subject to lifetime limits imposed by other laws, regulations or regulatory policies. |
| Nature of Advice | The investigation works herein are intended to develop and present sound, scientifically valid data concerning actual site conditions. Senversa does not seek or purport to provide legal or business advice. |

6.0 References

ANZECC & ARMCANZ (2000). Australian and New Zealand Guidelines for Fresh and Marine Water Quality. Australian and New Zealand Environment and Conservation Council and Agriculture and Resource Council of Australia and New Zealand.

ANZG (2018) Australian and New Zealand Guidelines for Fresh and Marine Water Quality. Australian and New Zealand Governments and Australian state and territory governments, Canberra ACT, Australia. Available at www.waterquality.gov.au/anz-quidelines.

Drapper D, Olive K, McAlister T, Coleman R, Lampard J-L (2022) A Review of Pollutant Concentrations in Urban Stormwater Across Eastern Australia, After 20 Years. Front. Environ. Chem. 3:853764.

Fletcher T., Duncan H., Poelsma P., Lloyd S. (2004). *Stormwater Flow and Quality, and the Effectiveness of Non-Proprietary Stormwater Treatment Measures: A Review and Gap Analysis*. Cooperative Research Centre for Catchment Hydrology, Technical Report 04/8, December 2004.

NEPC (1999). National Environment Protection Measure, Assessment of Site Contamination, as amended 2013 (ASC NEPM 2013). National Environment Protection Council.

NSW EPA (2020). *Contaminated Land Guidelines: Consultants Reporting on Contaminated Land.* 5 May 2020. NSW Environment Protection Authority.

NHMRC (2008) *Guidelines for Managing Risks in Recreational Water*. National Health and Medical Research Council.

NHMRC & NRMMC (2011). Australian Drinking Water Guidelines Paper 6 National Water Quality Management Strategy. Version 3.7 Updated January 2022. National Health and Medical Research Council, National Resource Management Ministerial Council, Commonwealth of Australia, Canberra. (ADWG, 2022).

Senversa (2022). Water Management Plan reDirect Resource Recovery Facility – 24 Davis Road, Wetherill Park NSW. 5 April 2022.

Senversa (2023). Surface Water Monitoring Report – Annual 2023, ReDirect Resource Recovery Facility, 24 Davis Road, Wetherill Park, NSW. 21 September 2023.



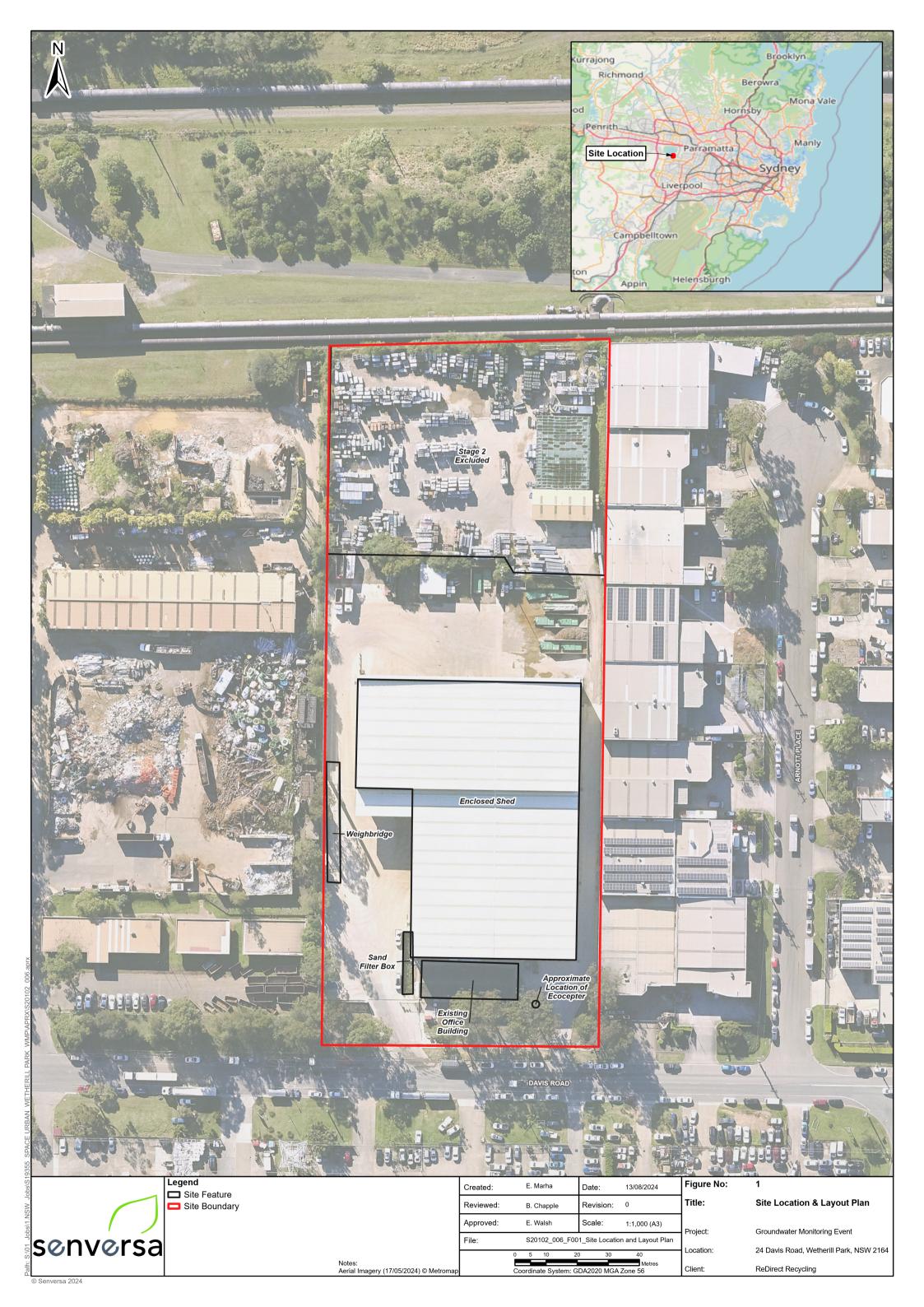
Figures

Figure 1: Site Location & Layout Plan

Figure 2: Surface Water and Groundwater Sampling Locations

Figure 3: Surface Water Exceedances

Figure 4: Groundwater Exceedances











Tables

Table 1: Groundwater Gauging Measurements

Table 2: Groundwater Geochemical Parameters

Table 3: Surface Water Analytical Results

Table 4: Groundwater Analytical Results

Table 1: Groundwater Gauging Measurements

Client: ReDirect Recycling
Site Address: ReDirect Resource Recovery Facility, 24 Davis Road, Wetherill Park, NSW



| Monito | oring Data | | Survey Data | | Groundwater Data | | | | | | | | | |
|------------------|-------------|----------|-------------|---------------|------------------|------------------|------------------|----------------------|--------------------------|-----------------------------|--|--|--|--|
| Location Code | Date Gauged | Easting | Northing | Top of Casing | Depth to Water | Total Well Depth | Depth to Product | Product Thickness | Groundwater Elevation | Gauging Comments | | | | |
| Code | | | | (m AHD) | (m bTOC) | (m bTOC) | (m bTOC) | (m) | (m AHD) | | | | | |
| MW1 | 6/02/2023 | 305715.1 | 6253955.6 | | 2.145 | 6.61 | - | - | | | | | | |
| MW1 | 8/02/2023 | 305715.1 | 6253955.6 | | 2.111 | 6.61 | - | - | | | | | | |
| MW1 | 2/08/2023 | 305715.1 | 6253955.6 | | 2.634 | 6.61 | - | - | | | | | | |
| MW1 | 7/02/2024 | 305715.1 | 6253955.6 | | 2.25 | 6.61 | - | - | | | | | | |
| MW1 | 9/02/2024 | 305715.1 | 6253955.6 | | 2.235 | 6.61 | - | - | | Before hydrasleeve sampling | | | | |
| MW1 | 9/07/2024 | 305715.1 | 6253955.6 | | 1.93 | 6.61 | - | - | | | | | | |
| MW1 | 11/07/2024 | 305715.1 | 6253955.6 | | 1.925 | 6.61 | - | - | | | | | | |
| MW2 | 6/02/2023 | 305714.5 | 6253975.4 | | 2.96 | 7.59 | - | - | | | | | | |
| MW2 | 8/02/2023 | 305714.5 | 6253975.4 | | 2.957 | 7.59 | - | - | | | | | | |
| MW2 | 2/08/2023 | 305714.5 | 6253975.4 | | 3.19 | 7.57 | - | - | | | | | | |
| MW2 | 7/02/2024 | 305714.5 | 6253975.4 | | 3.01 | 7.57 | - | - | | | | | | |
| MW2 | 9/02/2024 | 305714.5 | 6253975.4 | | 2.97 | 7.57 | - | - | | Before hydrasleeve sampling | | | | |
| MW2 | 9/07/2024 | 305714.5 | 6253975.4 | | 2.577 | 7.58 | - | - | | | | | | |
| MW2 | 11/07/2024 | 305714.5 | 6253975.4 | | 2.555 | 7.58 | - | - | | | | | | |
| MW3 | 6/02/2023 | 305677.6 | 6254060.5 | | 2.971 | 8.08 | - | - | | | | | | |
| MW3 | 8/02/2023 | 305677.6 | 6254060.5 | | 2.945 | 8.08 | - | - | | | | | | |
| MW3 | 2/08/2023 | 305677.6 | 6254060.5 | | 3.599 | 8.07 | - | - | | | | | | |
| MW3 | 7/02/2024 | 305677.6 | 6254060.5 | | 3.18 | 8.05 | - | - | | | | | | |
| MW3 | 9/02/2024 | 305677.6 | 6254060.5 | | 3.175 | 8.05 | - | - | | Before hydrasleeve sampling | | | | |
| MW3 | 9/07/2024 | 305677.6 | 6254060.5 | | 2.88 | 8.05 | - | - | | | | | | |
| MW3 | 11/07/2024 | 305677.6 | 6254060.5 | | 2.875 | 8.05 | - | - | | | | | | |
| MW4 | 6/02/2023 | 305722.8 | 6254066.3 | | 2.205 | 6.99 | - | - | | | | | | |
| MW4 | 8/02/2023 | 305722.8 | 6254066.3 | | 2.224 | 6.99 | - | - | | | | | | |
| MW4 | 2/08/2023 | 305722.8 | 6254066.3 | | 2.565 | 6.98 | - | - | | | | | | |
| MW4 | 7/02/2024 | 305722.8 | 6254066.3 | | 2.65 | 6.98 | - | - | | | | | | |
| MW4 | 9/02/2024 | 305722.8 | 6254066.3 | | 2.53 | 6.98 | - | - | | Before hydrasleeve sampling | | | | |
| MW4 | 14/02/2024 | 305722.8 | 6254066.3 | | 2.77 | 6.98 | - | - | | Post hydrasleeve sampling | | | | |
| MW4 | 9/07/2024 | 305722.8 | 6254066.3 | | 2.325 | 6.98 | - | - | | | | | | |
| MW4 | 11/07/2024 | 305722.8 | 6254066.3 | | 2.31 | 6.98 | - | - | | | | | | |
| MW6 | 6/02/2023 | 305682.0 | 6254069.3 | | 4.444 | 7.19 | - | - | | | | | | |
| MW6 | 8/02/2023 | 305682.0 | 6254069.3 | | 4.444 | 7.19 | - | - | | | | | | |
| MW6 | 2/08/2023 | 305682.0 | 6254069.3 | | 4.748 | 7.17 | - | - | | | | | | |
| MW6 | 7/02/2024 | 305682.0 | 6254069.3 | | 4.369 | 7.16 | - | - | | | | | | |
| MW6 | 9/02/2024 | 305682.0 | 6254069.3 | | 4.357 | 7.16 | - | - | | Before hydrasleeve sampling | | | | |
| MW6 | 9/07/2024 | 305682.0 | 6254069.3 | | 3.565 | 7.16 | - | - | | Brown silt on IP | | | | |
| MW6 | 11/07/2024 | 305682.0 | 6254069.3 | | 3.557 | 7.16 | - | - | | | | | | |

Project: Baseline Groundwater Condition Report - 2024

Table 2: Groundwater Geochemical Parameters

Client: ReDirect Recycling

Site Address: ReDirect Resource Recovery Facility, 24 Davis Road, Wetherill Park, NSW



| Monitoring We | ell Information | | | | | | Water Quality | Stabilised Res | sults | | | | |
|---------------|-----------------|-----------|------------|--------|-------|----------|---------------|----------------|----------------------|---|-------------|---------------------|--|
| Location Code | Sample Date | DO (mg/L) | EC (µS/cm) | TDS | pН | ORP (Er) | Redox (mV) | Temp (°C) | | Field O | oservations | | |
| Location Code | Sample Date | ±10% | ±3% | | ±0.05 | (mV) | ±10mV | ±10% | Colour | Sheen | Odour | Turbidity | |
| MW1 | 8/02/2023 | 2.02 | 22,382 | 14,548 | 6.47 | 28.3 | 233.3 | 22.4 | orange | no sheen | no odour | Suspended sediments | |
| MW1 | 14/08/2023 | 6.33 | 19,738 | 12,830 | 6.34 | 116.4 | 321.4 | 17.0 | colourless to orange | no sheen | no odour | Non-turbid | |
| MW1 | 9/02/2024 | 3.07 | 25,870 | 16,816 | 6.66 | 38.1 | 243.1 | 22.3 | colourless | no sheen | no odour | Suspended sediments | |
| MW1 | 11/07/2024 | 1.06 | 22,394 | 14,556 | 6.27 | -58.0 | 147.0 | 17.5 | light brown | no sheen | no odour | Slightly turbid | |
| MW2 | 8/02/2023 | 1.35 | 21,545 | 14,004 | 6.33 | 154.4 | 359.4 | 20.8 | colourless | no sheen | no odour | Slightly turbid | |
| MW2 | 14/08/2023 | 1.20 | 17,006 | 11,054 | 6.44 | 129.0 | 334.0 | 17.0 | colourless | no sheen | no odour | Slightly turbid | |
| MW2 | 9/02/2024 | 3.05 | 27,224 | 17,696 | 6.59 | 92.7 | 297.7 | 23.2 | colourless | no sheen | no odour | Suspended sediments | |
| MW2 | 11/07/2024 | 1.42 | 23,588 | 15,332 | 6.45 | -17.6 | 187.4 | 18.2 | colourless | no sheen | no odour | Suspended sediments | |
| MW3 | 8/02/2023 | 0.79 | 29,765 | 19,347 | 5.78 | 65.7 | 270.7 | 23.6 | colourless | no sheen | no odour | Slightly turbid | |
| MW3 | 14/08/2023 | 0.92 | 24,992 | 16,245 | 5.91 | 62.6 | 267.6 | 15.9 | colourless | no sheen | no odour | Slightly turbid | |
| MW3 | 9/02/2024 | 2.11 | 34,645 | 22,519 | 6.50 | 1.5 | 206.5 | 23.5 | light brown | no sheen | no odour | Slightly turbid | |
| MW3 | 11/07/2024 | 2.20 | 28,201 | 18,331 | 5.86 | 15.0 | 220.0 | 18.3 | light brown | no sheen | no odour | Slightly turbid | |
| MW4 | 8/02/2023 | 0.00 | 17,881 | 11,623 | 6.54 | 69.0 | 274.0 | 22.7 | colourless | no sheen | no odour | Slightly turbid | |
| MW4 | 14/08/2023 | 1.70 | 7,133 | 4,636 | 6.55 | 18.7 | 223.7 | 17.1 | colourless | no sheen | no odour | Non-turbid | |
| MW4 | 14/02/2024 | 3.80 | 19,817 | 12,881 | 6.66 | -11.2 | 193.8 | 29.6 | colourless | no sheen | no odour | Slightly turbid | |
| MW4 | 11/07/2024 | 1.29 | 14,807 | 9,625 | 6.64 | -58.7 | 146.3 | 18.4 | colourless | no sheen | no odour | Non-turbid | |
| MW6 | 8/02/2023 | 0.10 | 2,323 | 1,510 | 7.19 | 89.8 | 294.8 | 22.7 | light brown | no sheen | no odour | Moderately turbid | |
| MW6 | 14/08/2023 | 0.81 | 1,362 | 885 | 7.22 | -6.4 | 198.6 | 16.8 | yellow | no sheen | sulphurous | Slightly turbid | |
| MW6 | 9/02/2024 | 5.30 | 2,204 | 1,433 | 7.41 | 56.6 | 261.6 | 22.5 | colourless | no sheen | no odour | Slightly turbid | |
| MW6 | 11/07/2024 | - | - | - | - | - | - | - | parameter | parameters not collected due to limited water available | | | |

Comments

Values presented are those after stabilisation. In accordance with EPA Publication 669, the parameters were considered stable when three consecutive readings (obtained several minutes apart) were within the specified parameters.

DO = Dissolved Oxygen

EC = Electrical Conductivity.

TDS = Total Dissolved Solids

ORP = Oxidation Reduction Potential as millivolts (mV). Field values (Er values, mV) taken with redox probe with a platinum electrode and silver/silver chloride reference electrode. For interpretation of the Er results can be converted to Eh values using the following conversion: Eh (mV) = Er (mV) + 205.

^{*} TDS calculated by EC multiplied by 0.65



| | | | | | Location Code | SW1 | SW1 | SW1 | SW1 | SW2 | SW2 | SW2 | SW2 |
|--|--------------|---------|---|--|--|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| | | | | | Field ID | SW1 | SW1 | SW 1 | SW1 | SW2 | SW2 | SW 2 | SW2 |
| | | | | | Date | 10/02/2023 | 14/08/2023 | 07/02/2024 | 09/07/2024 | 10/02/2023 | 14/08/2023 | 07/02/2024 | 09/07/2024 |
| | | | | | Sample Type | Normal |
| | 1 | 1 | ANIZO (0040) | ANIZEGO (2022) | Lab Report No. | ES2304342 | ES2327328 | ES2403942 | ES2422553 | ES2304342 | ES2327328 | ES2403942 | ES2422553 |
| | Unit | EQL | ANZG (2018) Aquatic ecosystems DGV - highly disturbed (90%) - | ANZECC (2000) - physical stressors - South-east Australia Lowland River | NHMRC (2008) Primary Contact Recreation - Health | | | | | | | | |
| Physical Parameters | | | | | | | | | | | | | |
| Total Dissolved Solids Total Suspended Solids | mg/L mg/L | 10 5 | | | | 240 86 | 316 238 | 374 91 | 7,260 | 352 69 | 105 39 | 394 8 | 282 90 |
| pH (Lab) | pH Units | 0.01 | | | 6.5-8.5 ^{#10} | - | 8.03 | - | 7,260 | - 69 | 7.75 | - | 90 |
| Metals | prionits | 0.01 | | | 0.5-0.5 | - | 8.03 | - | | | 7.75 | - | - |
| Arsenic (filtered) | mg/L | 0.001 | 0.042#1 | | 0.1 ^{#11} | 0.001 | 0.001 | 0.001 | < 0.001 | < 0.001 | < 0.001 | 0.003 | < 0.001 |
| Cadmium (filtered) | mg/L | 0.0001 | 0.0004#2 | | 0.02 ^{#11} | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 |
| Chromium (filtered) | mg/L | 0.001 | 0.0033 ^{#3} | | 0.5 ^{#12} | 0.002 | 0.001 | < 0.001 | 0.001 | 0.002 | < 0.001 | < 0.001 | 0.001 |
| Copper (filtered) | mg/L | 0.001 | 0.0018 ^{#4} | | 20 ^{#11} | 0.006 | 0.004 | 0.004 | 0.003 | 0.003 | 0.001 | 0.004 | 0.004 |
| Iron (filtered) | mg/L | 0.05 | 40 | | 140 ^{#13} | 0.06 | < 0.05 | < 0.05 | < 0.05 | 0.06 | < 0.05 | < 0.05 | < 0.05 |
| Lead (filtered) | mg/L | 0.001 | 0.0056#2 | | 0.1 ^{#11} | <0.001 | < 0.001 | <0.001 | <0.001 | <0.001 | < 0.001 | <0.001 | <0.001 |
| Manganese (filtered) | mg/L | 0.001 | 2.5 ^{#4} | | 5 ^{#11} | 0.01 | 0.016 | 0.02 | 0.045 | <0.01 | 0.007 | <0.01 | 0.015 |
| Mercury (filtered) | mg/L | 0.0001 | 0.0006#5 | | 0.01 ^{#11} | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| Nickel (filtered) | mg/L | 0.001 | 0.013 ^{#2} 0.015 ^{#2} | | 0.2 ^{#11} 60 ^{#13} | 0.001 | <0.001 | <0.001 | <0.001 | 0.001 | <0.001 | <0.001 | <0.001 |
| Zinc (filtered) Inorganics | mg/L | 0.005 | 0.015 | | 60 | <0.005 | 0.005 | <0.005 | <0.005 | <0.005 | 0.038 | 0.014 | 0.023 |
| Total Oxidised Nitrogen (as N) | mg/L | 0.01 | | 0.04 | | 0.36 | 0.68 | 0.41 | 0.32 | 0.50 | 0.62 | 0.28 | 0.51 |
| Total Kjeldahl Nitrogen | mg/L | 0.1 | | | | 0.3 | 1.7 | 1.7 | 22.6 | 1.0 | 0.7 | 0.9 | 3.8 |
| Total Nitrogen (as N) | mg/L | 0.1 | | 0.35 | | 0.7 | 2.4 | 2.1 | 22.9 | 1.5 | 1.3 | 1.2 | 4.3 |
| Phosphorus (as P) BTEX | mg/L | 0.01 | | 0.025 | | 0.06 | 0.35 | 0.10 | 6.68 | 0.19 | 0.09 | 0.03 | 0.63 |
| Benzene | μg/L | 1 | 1,300#4 | | 10 ^{#11} | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Toluene | μg/L | 2 | 230 ^{#4} | | 8,000 ^{#11} | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Ethylbenzene | μg/L | 2 | 110 ^{#4} | | 3,000**11 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Xylene (m & p) | μg/L | 2 | | | ., | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Xylene (o) | μg/L | 2 | 470 ^{#4} | | | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total Xylene | μg/L | 2 | | | 6,000 ^{#11} | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total BTEX | μg/L | 1 | | | | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Total Petroleum Hydrocarbons C6-C9 Fraction | μg/L | 20 | | | | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C10-C14 Fraction | μg/L | 50 | | | | <50 | <50 | 490 | <50 | <50 | <50 | <50 | <50 |
| C15-C28 Fraction | μg/L | 100 | | | | <100 | <100 | 560 | 290 | <100 | <100 | <100 | <100 |
| C29-C36 Fraction | μg/L | 50 | | | | <50 | <50 | <50 | 130 | <50 | <50 | <50 | <50 |
| C10-C36 Fraction (Sum) | μg/L | 50 | | | | <50 | <50 | 1,050 | 420 | <50 | <50 | <50 | <50 |
| Total Recoverable Hydrocarbons C6-C10 Fraction | μg/L | 20 | | | | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C6-C10 Fraction minus BTEX (F1) | µg/L | 20 | 440#6 | | 900#14 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| >C10-C16 Fraction | μg/L | 100 | | | | <100 | <100 | 630 | <100 | <100 | <100 | <100 | <100 |
| >C10-C16 Fraction minus naphthalene (F2) | μg/L | 100 | 440 ^{#6} | | 900#14 | <100 | <100 | 630 | <100 | <100 | <100 | <100 | <100 |
| >C16-C34 Fraction | μg/L | 100 | 640 ^{#7} | | 900#15 | <100 | <100 | 460 | 380 | <100 | <100 | <100 | <100 |
| >C34-C40 Fraction | μg/L | 100 | 640 ^{#8} | | 900#15 | <100 | <100 | <100 | <100 | <100 | <100 | <100 | <100 |
| >C10-C40 Fraction (Sum) | μg/L | 100 | | | | <100 | <100 | 1,090 | 380 | <100 | <100 | <100 | <100 |
| PAHs Acceptable po | ug/l | 4 | | | 5,300 ^{#13} | -1.0 | -4.0 | -4.0 | -4.0 | -4.0 | -4.0 | -4.0 | -1.0 |
| Acenaphthene Acenaphthylene | μg/L μg/L | 1 | | | 5,300 | <1.0 <1.0 |
| Anthracene | μg/L | 1 | 0.4 ^{#5} | | 18,000#13 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | μg/L | 1 | | | 12,000 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | μg/L | 0.5 | 0.2#5 | | 0.1 ^{#11} | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Benzo(b+j)fluoranthene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(g,h,i)perylene Benzo(k)fluoranthene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Chrysene | μg/L μg/L | 1 | | | | <1.0 <1.0 |
| Dibenz(a,h)anthracene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluoranthene | μg/L | 1 | 1.4 ^{#5} | | 8,000 ^{#13} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluorene | μg/L | 1 | | | 2,900#13 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Indeno(1,2,3-c,d)pyrene | μg/L | 1 | - #4 | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Naphthalene (VOC) | μg/L | 1 | 37#4 | | 700#16 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Naphthalene (VOC) Phenanthrene | μg/L μg/L | 5 1 | 2 ^{#5} | | | <1.0 | <1.0 | <1.0 | <5 <1.0 | <1.0 | <1.0 | <1.0 | <5 <1.0 |
| Pyrene | μg/L μg/L | 1 | 2 | | 1,200#13 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene TEQ (Zero) | μg/L | 0.5 | | | 0.1 ^{#17} | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L | 0.5 | | | | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Phenois | | | | | | | | | | | | | |
| 2-Methylphenol | μg/L | 1 | | | 9,300 ^{#13} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2-Nitrophenol | μg/L | 1 | 0#9 | | 0.000#13 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2,4-Dimethylphenol 3-&4-Methylphenol (m&p-cresol) | μg/L μg/L | 2 | 2 ^{#9} | | 3,600#13 | <1.0 <2.0 |
| 4-Chloro-3-methylphenol | μg/L μg/L | 1 | | | 14,000#13 | <1.0 | <1.0 | <2.0 | <1.0 | <1.0 | <2.0 | <1.0 | <2.0 |
| Phenol | μg/L μg/L | 1 | 600#4 | | 58,000 ^{#13} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Halogenated Phenois | P9/∟ | | 000 | | 20,000 | /1.0 | 71.0 | V1.0 | V1.0 | V1.0 | V1.0 | V1.0 | <1.0 |
| 2,4,5-Trichlorophenol | μg/L | 1 | | | 12,000 ^{#13} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2,4,6-Trichlorophenol | μg/L | 1 | 20 ^{#5} | | 200#11 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2,4-Dichlorophenol | μg/L | 1 | 160#5 | 4 | 2,000#11 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2,6-Dichlorophenol | μg/L | 1 | 34 ^{#9} | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| 2-Chlorophenol | μg/L | 1 | 490#5 | | 3,000 ^{#11} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Pentachlorophenol | µg/L | 2 | 10 ^{#5} | | 100#11 | <2.0 | <2.0 | <2.0 | <2.0 | <2.0 | <2.0 | <2.0 | <2.0 |

- #1 ANZG (2018). The more conservative value (Arsenic AsV) out of the available arsenic species was adopted for initial screening purposes.
 #2 ANZG (2018). Adjust DGVs for site-specific hardness using the hardness-dependent algorithm in Warne et al. (2018)
 #3 ANZG (2018). The more conservative value (Chromium CrIII) out of the available chromium species was adopted for initial screening purposes.

- #4 ANZG (2018)
 #5 ANZG (2018). Higher species protection level adopted as recommended
 #6 CRWB (2019). Lowest of values for gasoline (C4-C12) and diesel (C8-C21) range hydrocarbons.
 #7 CRWB (2019). Value for diesel (C8-C21) mixture.
- #8 CRWB (2019). Value for diesel (C8-C21) mixture. No value derived for TPH >C21 as not considered soluble; diesel value used for screening. #9 ANZG (2018). Unknown species protection level
- #11 NHMRC (2011) Health. Multiplied by a factor of x10
 #12 NHMRC (2011) Health. Guideline for Cr (VI) conservatively adopted for comparision to total chromium. Speciated analysis should be undertaken where guideline is exceeded. Multiplied by a factor of x10
 #13 USEPA Tap Water RSL (TR=1E-06; THQ=0.1) May 2024. Multiplied by a factor of x10
- #14 WHO (2008). Lowest derived value for aliphatic and aromatic fractions in this range. Multiplied by a factor of x10
- #15 Lowest derived value for aliphatic and aromatic fractions in this range (90 ug/L). Multiplied by a factor of x10 #16 NHMRC (2011) Health. Derived as per NHMRC (2011) based on TDI used for NEPM HSL derivation. Multiplied by a factor of x10 #17 NHMRC (2011) Health. Value is for BaP but applies to TEQ. Multiplied by a factor of x10

MW2 MW2

MW2

MW2

MW3

MW3

MW3 MW3

MW1 MW1 MW1



| | | | | | Field ID | MW1 | MW1 | MW1 | MW1 | MW2 | MW2 | MW2 | MW2 | MW3 | MW3 | MW3 | MW3 |
|--|--------------|-------------|---|---|--|--------------|---------------|--------------|--|--------------|--|----------------|--------------|-------------------------|--------------------------|--------------------------|--------------------------|
| | | | | | Date | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 |
| | | | | | Sample Type | Normal | Normal | Normal | Normal | Normal | Normal | Normal | Normal | Highest Conc. | Highest Conc. | Highest Conc. | Highest Conc. |
| | | | | | Lab Report No. | ES2304011 | ES2327328 | ES2404239 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 | ES2304011 and 316159 | ES2327328 and 1020195 | ES2404239 and 1067666 | ES2423038 and 1117968 |
| | Unit | EQL | ANZG (2018) Aquatic ecosystems DGV- highly disturbed (90%) freshwater | NEPM (2013) Table 1A(4) Comm/Ind HSL D Vapour Intrusion, Sand (2m-4m) | NHMRC (2008) Primary Contact Recreation - Health | | | | | | | | | | | | |
| Physical Parameters | | | | | | | | | | | | | | | | | |
| Electrical Conductivity | μS/cm | 1 | | | | 25,800 | - | - | - | 25,700 | - | - | - | 34,200 | - | - | - |
| Total Dissolved Solids | mg/L | 1 | | | 6.5-8.5 ^{#12} | 16,800 | - | - | - | 16,700 | - | - | - | 22,200 | - | - | - |
| pH (Lab) | pH Units | 0.01 | | | 6.5-8.5 | 7.74 | - | - | - | 7.70 | - | - | - | 7.09 | - | - | - |
| Metals Arsenic (filtered) | | 0.004 | 0.042#2 | | 0.1#13 | 0.044 | 0.000 | 0.040 | 0.007 | 0.004 | 0.004 | 0.004 | 0.000 | 0.004 | 0.000 | 0.044 | 0.000 |
| Cadmium (filtered) | mg/L | 0.001 | 0.042 | | 0.02 ^{#13} | 0.011 | 0.008 | 0.012 | 0.007 | 0.004 | 0.004 | 0.004 | 0.008 | 0.004 | 0.002 | 0.011 | 0.003 |
| | mg/L | 0.0001 | | | 0.02 0.5 ^{#14} | <0.0001 | <0.0001 | <0.0001 | 0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | 0.0001 | <0.0010 | <0.0010 | <0.0001 |
| Chromium (filtered) | mg/L | 0.001 | 0.0033 ^{#4} | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | 0.002 | <0.010 | 0.005 | <0.001 |
| Copper (filtered) | mg/L | 0.001 | 0.0018 ^{#5} | | 20 ^{#13} | 0.015 | <0.001 | <0.001 | 0.003 | 0.011 | <0.001 | <0.001 | <0.001 | <0.010 | 0.002 | 0.006 | 0.002 |
| Iron (filtered) | mg/L | 0.01 | #2 | | 140 ^{#15} | 4.97 | 2.01 | 3.96 | 87.2 | 0.40 | 0.58 | 0.54 | 2.30 | 5.05 | 6.04 | 8.08 | 7.4 |
| Lead (filtered) | mg/L | 0.001 | 0.0056 ^{#3} | | 0.1 ^{#13} | < 0.001 | < 0.001 | < 0.001 | < 0.001 | < 0.001 | < 0.001 | < 0.001 | <0.001 | < 0.010 | 0.002 | 0.005 | < 0.001 |
| Manganese (filtered) | mg/L | 0.001 | 2.5 ^{#5} | | 5 ^{#13} | 0.92 | 2.26 | 0.59 | 4.84 | 0.96 | 1.00 | 1.76 | 3.28 | 6.15 | 6.57 | 7.08 | 7.4 |
| Mercury (filtered) | mg/L | 0.00005 | 0.0006 ^{#6} | | 0.01 ^{#13} | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 |
| Nickel (filtered) | mg/L | 0.001 | 0.013 ^{#3} | | 0.2 ^{#13} | 0.023 | 0.036 | 0.015 | 0.156 | 0.006 | 0.005 | 0.006 | 0.006 | 0.191 | 0.207 | 0.197 | 0.200 |
| Zinc (filtered) | mg/L | 0.001 | 0.015 ^{#3} | | 60 ^{#15} | 0.012 | 0.045 | 0.016 | 0.174 | 0.008 | 0.009 | 0.009 | < 0.005 | 0.23 | 0.122 | 0.253 | 0.25 |
| Inorganics | | | #5 | | | | | <u> </u> | | <u> </u> | | <u> </u> | | | | | |
| Ammonia (as N) | mg/L | 0.01 | 1.43 ^{#5} | | #40 | 0.71 | 0.49 | 0.48 | 0.19 | 0.52 | 0.52 | 0.44 | 0.26 | 0.22 | 0.29 | 0.29 | 0.28 |
| Nitrate (as N) | mg/L | 0.01 | 3.8 ^{#7} | | 110 ^{#16} | <0.10 | 0.02 | < 0.01 | < 0.01 | 0.03 | < 0.01 | <0.01 | 0.02 | < 0.01 | < 0.01 | 0.10 | 0.01 |
| Nitrite (as N) | mg/L | 0.01 | | | 9 ^{#17} | < 0.10 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 |
| Total Oxidised Nitrogen (as N) | mg/L | 0.01 | | | | <0.10 | 0.02 | < 0.01 | < 0.01 | 0.03 | < 0.01 | < 0.01 | 0.02 | 0.02 | < 0.01 | 0.11 | 0.02 |
| Total Kjeldahl Nitrogen | mg/L | 0.1 | | | | 0.9 | 0.6 | 0.6 | 1.1 | 1.0 | 0.6 | 0.8 | 0.6 | 1.3 | 0.5 | 1.3 | 1.0 |
| Total Nitrogen (as N) | mg/L | 0.1 | | | | 0.9 | 0.6 | 0.6 | 1.1 | 1.0 | 0.6 | 0.8 | 0.6 | 1.3 | 0.5 | 1.4 | 1.0 |
| Phosphorus (as P) | mg/L | 0.01 | | | | < 0.05 | 0.02 | <0.01 | 0.04 | 0.06 | 0.04 | 0.05 | 0.05 | 0.8 | 0.02 | 0.08 | 0.04 |
| Phosphate (as P) Ortho-phosphate (as P) | mg/L | 0.01 | | | | -0.04 | - | - | - | - 0.00 | - | - | - | -0.01 | 0.03 | 0.05 | 0.03 |
| Fluoride | mg/L | 0.01 | | | 15 ^{#13} | <0.01 | - | - | - | 0.02 | - | - | - | <0.01 | - | - | - |
| Sodium Absorption Ratio (filtered) | mg/L | 0.1 0.01 | | | 13 | 0.8 30.4 | - | - | | 0.7 31.6 | - | - | - | 1.2 37.8 | - | - | |
| Major lons | + - | 0.01 | | | | 30.4 | | | | 31.0 | _ | - - | - | 37.0 | - | - | |
| Calcium (filtered) | mg/L | 1 | | | | 273 | - | - | - | 232 | - | - | - | 181 | - | - | - |
| Chloride | mg/L | 1 | | | | 8,840 | - | - | - | 8,800 | - | - | - | 11,900 | - | - | - |
| Magnesium (filtered) | mg/L | 1 | | | | 810 | - | - | - | 826 | - | - | - | 1,040 | - | - | - |
| Potassium (filtered) | mg/L | 1 | | | | 25 | - | - | - | 21 | - | - | - | 14 | - | - | - |
| Sulfate (as SO4) (filtered) | mg/L | 1 | | | | 691 | - 1 | - | - 1 | 756 | - | - | - | 907 | - | - | - |
| Sodium (filtered) | mg/L | 1 | | | | 4,430 | - | - | - | 4,590 | - | - | - | 5,980 | - | - | - |
| Anions Total | meq/L | 0.01 | | | | 282 | - | - | - | 280 | - | - | - | 359 | - | - | - |
| Cations Total | meq/L | 0.01 | | | | 274 | - | - | - | 280 | - | - | - | 355 | - | - | - |
| Ionic Balance | % | 0.01 | | | | 1.52 | - | - | - | 0.09 | - | - | - | 0.55 | - | - | - |
| Alkalinity | | <u> </u> | | | | | igwdown | | | | | ' | | | | | |
| Bicarbonate Alkalinity (as CaCO3) Carbonate Alkalinity (as CaCO3) | mg/L | 1 | | | | 916 | - | - | - | 815 | - | - | - | 222 | - | - | - |
| Hydroxide Alkalinity (as CaCO3) | mg/L mg/L | 1 | | | | <1 <1 | - | - | - | <1 <1 | - | - | - | <1 <1 | - | - | - |
| Total Alkalinity (as CaCO3) | mg/L | 1 | | | | 916 | - | - | - | 815 | - | - | - | 222 | - | - | - |
| Hardness (as CaCO3) (filtered) | mg/L | 1 | | | | 4,020 | - | - | - | 3,980 | - | - | - | 4,730 | - | - | - |
| ВТЕХ | T | | | | | ,, | | | | | | | | , | | | |
| Benzene | μg/L | 1 | 1,300#5 | 5,000 ^{#1} | 10 ^{#13} | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Toluene | μg/L | 1 | 230#5 | NL #1 | 8,000#13 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Ethylbenzene | μg/L | 1 | 110 ^{#5} | NL ^{#1} | 3,000#13 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Xylene (m & p) | µg/L | 2 | | | , | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Xylene (o) | μg/L | 1 | 470 ^{#5} | | | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total Xylene | μg/L | 2 | | NL ^{#1} | 6,000#13 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total BTEX | µg/L | 1 | | | ., | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Total Petroleum Hydrocarbons | T | | | | | | | | | | 1 | | 1 | | | | 1 |
| C6-C9 Fraction | μg/L | 10 | | | | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C10-C14 Fraction | μg/L | 50 | | | | <50 | <50 | <50 | 250 | <50 | <50 | <50 | <50 | <50 | <50 | <50 | 90 |
| C15-C28 Fraction | μg/L | 100 | | | | <100 | <100 | <100 | 170 | <100 | <100 | <100 | <100 | 140 | <100 | <100 | 810 |
| C29-C36 Fraction | μg/L | 50 | | | | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 | 300 |
| C10-C36 Fraction (Sum) | μg/L | 50 | | | | <50 | <50 | <50 | 420 | <50 | <50 | <50 | <50 | 140 | <50 | <50 | 810 |
| Total Recoverable Hydrocarbons | | 4.5 | | | | ~~ | | | | | | | 0.5 | 00 | | 000 | 22 |
| C6-C10 Fraction | μg/L | 10 | 4.40#R | 0.000#1 | 200#18 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C6-C10 Fraction minus BTEX (F1) | μg/L | 10 | 440 ^{#8} | 6,000#1 | 900#18 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| >C10-C16 Fraction | μg/L | 50 | #9 | #1 | #18 | <100 | <100 | <100 | 260 | <100 | <100 | <100 | <100 | 130 | <100 | <100 | 110 |
| >C10-C16 Fraction minus naphthalene (F2) | μg/L | 50 | 440#8 | NL ^{#1} | 900 ^{#18} | <100 | <100 | <100 | 260 | <100 | <100 | <100 | <100 | 130 | <100 | <100 | 110 |
| | μg/L | 100 | 640 ^{#9} | I | 900#19 | <100 | <100 | <100 | 260 | <100 | <100 | <100 | <100 | <100 | <100 | <100 | 600 |
| >C16-C34 Fraction | | | #40 | | #10 | | | - | | | + | | 1 | 1 | 1 | | |
| >C16-C34 Fraction >C34-C40 Fraction >C10-C40 Fraction (Sum) | μg/L μg/L | 100 | 640 ^{#10} | | 900 ^{#19} | <100 <100 | <100 <100 | <100 <100 | <100 520 | <100 <100 | <100 <100 | <100 <100 | <100 <100 | <100 130 | <100 <100 | <100 <100 | 200 910 |

Location Code MW1



| | | | | i | | | | | | | | | , |
|--|--------------|-----------|--|--------------------------------------|---|-------------------|---------------------|-------------------|-------------------|-------------------|----------------------|----------------------|-------------------|
| | | | | | Location Code | MW4 | MW4 | MW4 | MW4 | MW6 | MW6 | MW6 | MW6 |
| | | | | | Field ID Date | MW4 08/02/2023 | MW4 14/08/2023 | MW4 14/02/2024 | MW4 11/07/2024 | MW6 08/02/2023 | MW6 14/08/2023 | MW6 09/02/2024 | MW6 11/07/2024 |
| | | | | | Sample Type | Normal | Normal | Normal | Normal | Normal | Normal | Normal | Normal |
| | | | | | | ES2304011 | ES2327328 | ES2404752 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 |
| | 1 | T | | | Lab Report No. | E32304011 | E32321326 | E32404732 | E32423036 | E32304011 | E32321326 | E32404239 | E32423036 |
| | | | ANZG (2018) | NEPM (2013) Table | | | | | | | | | |
| | Unit | EQL | Aquatic ecosystems DGV- | 1A(4) Comm/Ind HSL | NHMRC (2008) Primary Contact | | | | | | | | |
| | Offic | | highly disturbed (90%) | _ | Recreation - Health | | | | | | | | |
| | | | freshwater | Sand (2m-4m) | | | | | | | | | |
| Physical Parameters | | | | | | | | | | | | | |
| Electrical Conductivity | μS/cm | 1 | | | | 19,900 | - | - | - | 2,310 | - | - | - |
| Total Dissolved Solids | mg/L | 1 | | | #12 | 12,900 | - | - | - | 1,500 | - | - | - |
| pH (Lab) | pH Units | 0.01 | | | 6.5-8.5 ^{#12} | 7.72 | - | - | - | 8.06 | - | - | - |
| Metals Arsenic (filtered) | mg/L | 0.001 | 0.042#2 | | 0.1#13 | 0.005 | 0.007 | 0.008 | 0.007 | < 0.001 | 0.002 | 0.002 | <0.001 |
| Cadmium (filtered) | mg/L | 0.0001 | 0.0004#3 | | 0.02#13 | <0.0001 | <0.007 | <0.0001 | <0.007 | <0.001 | <0.002 | <0.002 | <0.0001 |
| Chromium (filtered) | mg/L | 0.001 | 0.0033#4 | | 0.5#14 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Copper (filtered) | mg/L | 0.001 | 0.0018 ^{#5} | | 20#13 | 0.005 | <0.001 | 0.001 | <0.001 | 0.003 | <0.001 | <0.001 | <0.001 |
| Iron (filtered) | mg/L | 0.01 | | | 140#15 | 1.22 | 2.91 | 2.04 | 1.90 | < 0.05 | 0.20 | <0.05 | <0.05 |
| Lead (filtered) | mg/L | 0.001 | 0.0056#3 | | 0.1#13 | < 0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Manganese (filtered) | mg/L | 0.001 | 2.5 ^{#5} | | 5 ^{#13} | 5.45 | 6.04 | 5.03 | 4.00 | 0.04 | 0.225 | 0.06 | <0.01 |
| Mercury (filtered) | mg/L | 0.00005 | 0.0006 ^{#6} | | 0.01 ^{#13} | <0.0001 | < 0.0001 | <0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | < 0.0001 | <0.0001 |
| Nickel (filtered) | mg/L | 0.001 | 0.013 ^{#3} | | 0.2 ^{#13} | 0.021 | 0.020 | 0.017 | 0.011 | < 0.001 | 0.002 | 0.001 | <0.001 |
| Zinc (filtered) | mg/L | 0.001 | 0.015 ^{#3} | | 60 ^{#15} | < 0.005 | < 0.005 | 0.006 | < 0.005 | < 0.005 | 0.006 | < 0.005 | < 0.005 |
| Inorganics | | | #F | | | | | | | | | | |
| Ammonia (as N) | mg/L | 0.01 | 1.43 ^{#5} | | #16 | 0.34 | 0.32 | 0.30 | 0.28 | 0.02 | 0.09 | <0.01 | <0.01 |
| Nitrate (as N) | mg/L | 0.01 | 3.8 ^{#7} | | 110 ^{#16} 9 ^{#17} | 0.01 | <0.01 | <0.01 | <0.01 | 1.00 | 0.18 | 1.93 | 1.64 |
| Nitrite (as N) Total Oxidised Nitrogen (as N) | mg/L | 0.01 | | | 9*'' | <0.01 | <0.01 | <0.01 <0.01 | <0.01 <0.01 | 0.25 1.25 | <0.01 0.18 | <0.01 1.93 | <0.01 1.64 |
| Total Kjeldahl Nitrogen | mg/L mg/L | 0.01 | | | | 1.1 | <0.01 0.5 | 1.2 | 0.4 | 0.4 | 0.18 | 1.93 | 0.3 |
| Total Nitrogen (as N) | mg/L | 0.1 | | | | 1.1 | 0.5 | 1.2 | 0.4 | 1.6 | 0.6 | 2.9 | 1.9 |
| Phosphorus (as P) | mg/L | 0.01 | | | | 0.09 | 0.01 | 0.07 | 0.04 | 0.09 | 0.14 | 0.03 | 0.05 |
| Phosphate (as P) | mg/L | 0.01 | | | | - | - | - | - | - | - | - | - |
| Ortho-phosphate (as P) | mg/L | 0.01 | | | #12 | < 0.01 | - | - | - | < 0.01 | - | - | - |
| Fluoride | mg/L | 0.1 | | | 15 ^{#13} | 1.6 | - | - | - | 1.8 | - | - | - |
| Sodium Absorption Ratio (filtered) Major Ions | - | 0.01 | | | | 21.4 | - | - | - | 6.70 | - | - | - |
| Calcium (filtered) | mg/L | 1 | | | | 299 | - | - | - | 50 | - | - | - |
| Chloride | mg/L | 1 | | | | 6,680 | - | - | - | 341 | - | - | - |
| Magnesium (filtered) | mg/L | 1 | | | | 786 | - | - | - | 112 | - | - | - |
| Potassium (filtered) Sulfate (as SO4) (filtered) | mg/L | 1 | | | | 35 | - | - | - | 6 | - | - | - |
| Sodium (filtered) | mg/L mg/L | 1 | | | | 280 3,100 | - | - | - | 44 373 | - | - | - |
| Anions Total | meg/L | 0.01 | | | | 216 | - | - | - | 27.2 | - | - | - |
| Cations Total | meq/L | 0.01 | | | | 215 | - | - | - | 28.1 | - | - | - |
| Ionic Balance | % | 0.01 | | | | 0.25 | - | - | - | 1.61 | - | - | - |
| Alkalinity | | | | | | | | | | | | | |
| Bicarbonate Alkalinity (as CaCO3) Carbonate Alkalinity (as CaCO3) | mg/L mg/L | 1 | | | | 1,110 <1 | - | - | - | 834 <1 | - | - | - |
| Hydroxide Alkalinity (as CaCO3) | mg/L | 1 | | | | <1 | - | - | - | <1 | - | - | - |
| Total Alkalinity (as CaCO3) | mg/L | 1 | | | | 1,110 | - | - | - | 834 | - | - | - |
| Hardness (as CaCO3) (filtered) | mg/L | 1 | | | | 3,980 | - | | - | 586 | - | - | - |
| BTEX | | | 4.000#5 | E 000#1 | 40#13 | | | | | | | | |
| Benzene | μg/L | 1 | 1,300 ^{#5} | 5,000 ^{#1} | 10 ^{#13} 8,000 ^{#13} | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Toluene Ethylbenzene | μg/L | 1 | 230" ⁵ 110 ^{#5} | NL ^{#1} NL ^{#1} | 8,000*** 3.000 ^{#13} | <2 | <2 | 2 | <2 | <2 | <2 | <2 | <2 |
| Xylene (m & p) | μg/L μg/L | 2 | 110 | NL | 3,000 | <2 <2 | <2 <2 | <2 <2 | <2 <2 | <2 <2 | <2 <2 | <2 <2 | <2 <2 |
| Xylene (o) | μg/L | 1 | 470 ^{#5} | | | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total Xylene | μg/L | 2 | | NL #1 | 6,000#13 | <2 | <2 | <2 | <2 | <2 | <2 | <2 | <2 |
| Total BTEX | μg/L | 1 | | | | <1 | <1 | 2 | <1 | <1 | <1 | <1 | <1 |
| Total Petroleum Hydrocarbons | | | | | | | | | | | | | |
| C6-C9 Fraction | μg/L | 10 | | | | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C10-C14 Fraction C15-C28 Fraction | μg/L μg/L | 50 100 | | | | <50 <100 | <50 <100 | <50 <100 | <50 <100 | <50 <100 | <50 <100 | <50 <100 | <50 <100 |
| C29-C36 Fraction | μg/L | 50 | | | | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 |
| C10-C36 Fraction (Sum) | μg/L | 50 | | | | <50 | <50 | <50 | <50 | <50 | <50 | <50 | <50 |
| Total Recoverable Hydrocarbons | | | | | | | | | | | _ | | |
| C6-C10 Fraction | μg/L | 10 | #R | 0.0041 | #18 | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| C6-C10 Fraction minus BTEX (F1) >C10-C16 Fraction | μg/L | 10 | 440 ^{#8} | 6,000#1 | 900 ^{#18} | <20 | <20 | <20 | <20 | <20 | <20 | <20 | <20 |
| >C10-C16 Fraction >C10-C16 Fraction minus naphthalene (F2) | μg/L μg/L | 50 50 | 440#8 | NL ^{#1} | 900#18 | <100 <100 | <100 <100 | <100 <100 | <100 <100 | <100 <100 | <100 <100 | <100 <100 | <100 <100 |
| >C10-C16 Fraction minus napritrialene (F2) >C16-C34 Fraction | μg/L μg/L | 100 | 640 ^{#9} | INC | 900 ^{#19} | <100 | <100 | <100 | <100 | <100 | <100 | <100 | <100 |
| >C34-C40 Fraction | μg/L | 100 | 640 ^{#10} | | 900#19 | <100 | <100 | <100 | <100 | <100 | <100 | <100 | <100 |
| | | 50 | | | | <100 | <100 | <100 | <100 | <100 | <100 | <100 | <100 |
| >C10-C40 Fraction (Sum) | μg/L | 30 | the state of the s | | | <100 | <100 | <100 | <100 | <100 | <100 | <100 | 4.00 |



| Location Code | MW1 | MW1 | MW1 | MW1 | MW2 | MW2 | MW2 | MW2 | MW3 | MW3 | MW3 | MW3 |
|---------------|------------|---------------------|------------|------------|------------|------------|------------|------------|---------------|---------------|---------------|---------------|
| Field ID | MW1 | MW1 | MW1 | MW1 | MW2 | MW2 | MW2 | MW2 | MW3 | MW3 | MW3 | MW3 |
| Date | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 |
| Sample Type | Normal | Normal | Normal | Normal | Normal | Normal | Normal | Normal | Highest Conc. | Highest Conc. | Highest Conc. | Highest Conc. |
| | ES2304011 | ES2327328 | ES2404239 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 |
| Lah Report No | | 2304011 202321320 | | | | | | | and 316150 | and 1020105 | and 1067666 | and 1117069 |

| | | | | Lab Report No. | ES2304011 | ES2327328 | ES2404239 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 | and 316159 | and 1020195 | and 1067666 | and 1117968 |
|------|--|--|--|---|------------|---|--|---|--|--|--|------------|--|---|--|--------------|
| Unit | EQL | DGV- | D | NHMRC (2008) Primary Contact Recreation - Health | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | |
| μg/L | 1 | | | 5,300 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 1 | 0.4 ^{#6} | | 18,000 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 0.5 | 0.2 ^{#6} | | 0.1 ^{#13} | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 |
| μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 2 | | | | - | - | - | - | - | - | - | - | - | - | - | - |
| | 1 | | | | | | | | | | | | | | | <1.0 |
| | | | | | | | | | | | | | | | | <1.0 |
| | 1 | uc | | #15 | | 1 | | | | | | l | 1 | | | <1.0 |
| | 1 | 1.4** | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 1 | | | 2,900#15 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 1 | "" | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 1 | 37#5 | NL #1 | 700#20 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| | 5 | | | | - | - | - | <5 | - | - | - | <5 | - | - | - | <5 |
| μg/L | 1 | 2 ^{#6} | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| μg/L | 0.5 | | | 0.1 ^{#21} | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 |
| μg/L | 0.5 | | | | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 |
| μg/L | 5 | | | 0.1 ^{#21} | - | - | - | - | - | - | - | - | - | - | - | - |
| μg/L | 1 | | | | - | - | - | - | - | - | - | - | 0 | - | - | - |
| | | | | | | | | | | | | | | | | |
| μg/L | 1 | | | 9,300 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| μg/L | 1 | | | | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| μg/L | 1 | 2 ^{#11} | | 3,600 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| μg/L | 2 | | | | <2.0 | - | - | - | <2.0 | - | - | - | <2.0 | - | - | - |
| μg/L | 1 | | | | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| μg/L | 1 | 600 ^{#5} | | 58,000 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| | | | | | | | | | | | | | | | | |
| μg/L | 1 | | | 12,000 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| μg/L | 1 | 20 ^{#6} | | 200#13 | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| | 1 | 160 ^{#6} | | 2,000#13 | <1.0 | - | - | - | <1.0 | - | - | - | <1.0 | - | - | - |
| | 1 | 34 ^{#11} | | , | | - | - | - | | - | - | - | | - | - | - |
| | 1 | | | 3.000#13 | | _ | _ | - | | _ | _ | _ | | _ | - | _ |
| | 2 | | | | | <u> </u> | | _ | | _ | _ | | 1 | <u> </u> | _ | |
| | Hg/L H | ру/L 1 | Unit EQL Aquatic ecosystems DGV- highly disturbed (90%) freshwater µg/L | Unit EQL Aquatic ecosystems DGV-highly disturbed (90%) freshwater µg/L | Unit EQL | Unit EQL ANZG (2018) Aquatic ecosystems DGV- highly disturbed (90%) freshwater 14,40 Comm/Ind HSL D Primary Contact Recreation - Health 14,00 Primary Contact Recreation - Health 15,300 Primary Contact Recreation - Health 14,00 Primary Contact Recreation - Health 15,300 Primary Contact Recreation - Health 16,300 Pr | Unit EQL ANZG (2018) Aquatic ecosystems DGV Nighly disturbed (90%) Freshwater Name (2m-4m) NHMRC (2008) Primary Contact Recreation - Health Primary Contact Primary | Unit EQL Aquatic ecosystems DGV DGV highly disturbed (90%) freshwater NHMRC (2008) Primary Contact Recreation - Health Primary Contact Primary Contact Recreation - Health Primary Contact Primary Contact Recreation - Health Primary Contact Primar | Unit ECL ANZG (2018) ANZG (2018) D ANZG (2018) D | Variable Color Color | Unit Eol. Aquatic ecosystems b D6V- phythy disturbed (90%) Phythy disturbed (90%) Primary Contact Recreation - Health Sand (2m-4m) руд. 1 | Unit ECL | Unit ECL August coopystems Neph (2013) Table August coopystems OCV OCV | Lau Psychia Lau Psychia | List Fig. ANZG (2018) New (2013) Table Ne | Unit EOL |

Comments

- #1 Value for shallow (2-4 m bgl) sand aquifer adopted for initial screening.
- #2 ANZG (2018). The more conservative value (Arsenic AsV) out of the available arsenic species was adopted for initial screening purposes.
- #3 ANZG (2018). Adjust DGVs for site-specific hardness using the hardness-dependent algorithm in Warne et al. (2018)
- #4 ANZG (2018). Chromium CrIII adopted for initial screening purposes.
- #5 ANZG (2018)
- #6 ANZG (2018). Higher species protection level adopted as recommended
- #7 Derived by NZ NIWA (2013) using ANZECC (2000) methodology. ANZECC (2000) value was withdrawn due to calculation errors.
- #8 CRWB (2019). Lowest of values for gasoline (C4-C12) and diesel (C8-C21) range hydrocarbons.
- #9 CRWB (2019). Value for diesel (C8-C21) mixture.
- #10 CRWB (2019). Value for diesel (C8-C21) mixture. No value derived for TPH >C21 as not considered soluble; diesel value used for screening.
- #11 ANZG (2018). Unknown species protection level
- #12 NHMRC (2008)
- #13 NHMRC (2011) Health. Multiplied by a factor of x10
- #14 NHMRC (2011) Health. Guideline for Cr (VI) conservatively adopted for comparision to total chromium. Speciated analysis should be undertaken where guideline is exceeded. Multiplied by a factor of x10
- #15 USEPA Tap Water RSL (TR=1E-06; THQ=0.1) May 2024. Multiplied by a factor of x10
- #16 NHMRC (2011) Health. Converted from guideline for nitrate (as nitrate). Multiplied by a factor of x10
- #17 NHMRC (2011) Health. Converted from guideline for nitrite (as nitrite). Multiplied by a factor of x10
- #18 WHO (2008). Lowest derived value for aliphatic and aromatic fractions in this range. Multiplied by a factor of x10
- #19 Lowest derived value for aliphatic and aromatic fractions in this range (90 ug/L). Multiplied by a factor of x10
- #20 NHMRC (2011) Health. Derived as per NHMRC (2011) based on TDI used for NEPM HSL derivation. Multiplied by a factor of x10
- #21 NHMRC (2011) Health. Value is for BaP but applies to TEQ. Multiplied by a factor of x10

MW4

MW4

MW6

MW6

MW6

MW6

MW6

MW6

MW6

MW6

MW4

MW4



| | | | | | Date | 08/02/2023 | 14/08/2023 | 14/02/2024 | 11/07/2024 | 08/02/2023 | 14/08/2023 | 09/02/2024 | 11/07/2024 |
|---|--------------|------------|---|---|--|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| | | | | | Sample Type | Normal |
| | | | | | Lab Report No. | ES2304011 | ES2327328 | ES2404752 | ES2423038 | ES2304011 | ES2327328 | ES2404239 | ES2423038 |
| | Unit | EQL | ANZG (2018) Aquatic ecosystems DGV- highly disturbed (90%) freshwater | NEPM (2013) Table 1A(4) Comm/Ind HSL D Vapour Intrusion, Sand (2m-4m) | NHMRC (2008) Primary Contact Recreation - Health | | | | | | | | |
| PAHs | | | | | | | | | | | | | |
| Acenaphthene | μg/L | 1 | | | 5,300 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Acenaphthylene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Anthracene | μg/L | 1 | 0.4 ^{#6} | | 18,000 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | μg/L | 0.5 | 0.2 ^{#6} | | 0.1 ^{#13} | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 | < 0.5 |
| Benzo(b+j)fluoranthene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(g,h,i)perylene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(b+j+k)fluoranthene | μg/L | 2 | | | | - | - | - | - | - | - | - | - |
| Benzo(k)fluoranthene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Chrysene Dibenz(a,h)anthracene | μg/L | 1 | | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| - | μg/L | 1 | 1.4 ^{#6} | | 8,000 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluoranthene | μg/L | 1 | 1.4 | | 2,900 ^{#15} | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluorene | μg/L | 1 | | | 2,900 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Indeno(1,2,3-c,d)pyrene | μg/L | 1 | 37 ^{#5} | NL ^{#1} | 700#20 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Naphthalene Naphthalene (VOC) | μg/L μg/L | 1 5 | 31 | NL | 700 | <1.0 | <1.0 | <1.0 | <1.0 <5 | <1.0 | <1.0 | <1.0 | <1.0 <5 |
| Phenanthrene | μg/L | 1 | 2 ^{#6} | | | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| - | | 1 | 2 | | 1,200 ^{#15} | | | | | <1.0 | | | |
| Pyrene Benzo(a)pyrene TEQ (Zero) | μg/L | - | | | 0.1 ^{#21} | <1.0 | <1.0 | <1.0 | <1.0 | | <1.0 | <1.0 | <1.0 |
| Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L μg/L | 0.5 0.5 | | | 0.1 | <0.5 <0.5 |
| Benzo(a)pyrene TEQ | μg/L | 5 | | | 0.1#21 | | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 | | |
| Total Positive PAHs | μg/L μg/L | 1 | | | 0.1 | - | - | - | - | - | - | - | - |
| Phenois | P9/ L | ' | | | | | _ | _ | _ | _ | _ | - | - |
| 2-Methylphenol | μg/L | 1 | | | 9,300#15 | <1.0 | - | _ | _ | <1.0 | - | - | - |
| 2-Nitrophenol | μg/L | 1 | | | 2,222 | <1.0 | - | - | - | <1.0 | - | - | - |
| 2,4-Dimethylphenol | μg/L | 1 | 2 ^{#11} | | 3,600#15 | <1.0 | _ | - | _ | <1.0 | - | - | _ |
| 3-&4-Methylphenol (m&p-cresol) | μg/L | 2 | | | ŕ | <2.0 | - | - | - | <2.0 | - | - | - |
| 4-Chloro-3-methylphenol | μg/L | 1 | | | 14,000 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - |
| Phenol | μg/L | 1 | 600 ^{#5} | | 58,000 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - |
| Halogenated Phenols | | | | | - | | | | | | | | |
| 2,4,5-Trichlorophenol | μg/L | 1 | | | 12,000 ^{#15} | <1.0 | - | - | - | <1.0 | - | - | - |
| 2,4,6-Trichlorophenol | μg/L | 1 | 20 ^{#6} | | 200#13 | <1.0 | - | - | - | <1.0 | - | - | - |
| 2,4-Dichlorophenol | μg/L | 1 | 160 ^{#6} | | 2,000#13 | <1.0 | - | - | - | <1.0 | - | - | - |
| 2,6-Dichlorophenol | μg/L | 1 | 34 ^{#11} | | | <1.0 | - | - | - | <1.0 | - | - | - |
| 2-Chlorophenol | μg/L | 1 | 490 ^{#6} | | 3,000#13 | <1.0 | - | - | - | <1.0 | - | - | - |
| Pentachlorophenol | μg/L | 2 | 10 ^{#6} | | 100#13 | <2.0 | - | - | - | <2.0 | - | - | - |

Location Code

Field ID

MW4

MW4

MW4

MW4

Comments

- #1 Value for shallow (2-4 m bgl) sand aquifer adopted for initial screening.
- #2 ANZG (2018). The more conservative value (Arsenic AsV) out of the available arsenic species was adopted for initial screening purpor
- #3 ANZG (2018). Adjust DGVs for site-specific hardness using the hardness-dependent algorithm in Warne et al. (2018)
- #4 ANZG (2018). Chromium CrIII adopted for initial screening purposes.
- #5 ANZG (2018)
- #6 ANZG (2018). Higher species protection level adopted as recommended
- #7 Derived by NZ NIWA (2013) using ANZECC (2000) methodology. ANZECC (2000) value was withdrawn due to calculation errors.
- #8 CRWB (2019). Lowest of values for gasoline (C4-C12) and diesel (C8-C21) range hydrocarbons.
- #9 CRWB (2019). Value for diesel (C8-C21) mixture.
- #10 CRWB (2019). Value for diesel (C8-C21) mixture. No value derived for TPH >C21 as not considered soluble; diesel value used for s
- #11 ANZG (2018). Unknown species protection level
- #12 NHMRC (2008)
- #13 NHMRC (2011) Health. Multiplied by a factor of x10
- #14 NHMRC (2011) Health. Guideline for Cr (VI) conservatively adopted for comparision to total chromium. Speciated analysis should I
- #15 USEPA Tap Water RSL (TR=1E-06; THQ=0.1) May 2024. Multiplied by a factor of x10
- #16 NHMRC (2011) Health. Converted from guideline for nitrate (as nitrate). Multiplied by a factor of x10
- #17 NHMRC (2011) Health. Converted from guideline for nitrite (as nitrite). Multiplied by a factor of x10
- #18 WHO (2008). Lowest derived value for aliphatic and aromatic fractions in this range. Multiplied by a factor of x10 #19 Lowest derived value for aliphatic and aromatic fractions in this range (90 ug/L). Multiplied by a factor of x10
- #20 NHMRC (2011) Health. Derived as per NHMRC (2011) based on TDI used for NEPM HSL derivation. Multiplied by a factor of x10
- #21 NHMRC (2011) Health. Value is for BaP but applies to TEQ. Multiplied by a factor of x10

Appendix A: OEMP Inspections





| • | RECYCLING PL | AN – WETHERILL PARK | INS | PECTION | I CHECK | (LIST not by chance |
|--------------|--|---|-----|-------------|---------|-------------------------|
| Locat | ion: | reDirect – Wetherill Park | Da | ate: | 01.08.2 | 3 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | M.P | Stewart |
| | | | | | | |
| 1. Ge | neral Management and m | nitigations 🗆 N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suital | bly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| | | | | | | |
| 2. Tra | ffic mitigations \square N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free by employees and visite | from obstruction and maintained for uprs? | ıse | Daily | Υ | |
| 2.3 | Vehicles are entering a | nd leaving the site in forward direction | n. | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust r | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and bo | t procedures are being implemented eing used? | | Daily | Y | |
| 3.2 | Good dust management the building): Sweeper working and bo | t procedures are implemented (outsid | e | Daily | Υ | |
| 3.3 | | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage syster | n? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | n stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra litter / debris? | ites have been inspected and clear of | | Monthly | N/A | |
| 5.5 | 1 | eaters, first flush devices and litter and are operating correctly. | | Monthly | N/A | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scourir | ng | Monthly | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | |

Monthly

Quarterly

(Mar, Jun,

Sep, Dec)

annually (Jun, Dec)

Bi-

N/A

N/A

N/A

the build up of litter material

flush placement of grate upon refitment.

Treatment areas and structures will be regularly checked for

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

5.7

5.8

5.9





| | TEAT WE ITERIEE I ARREITS | | | |
|---------|--|-------------------------------|-----------------|------------------------------------|
| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Vei | min and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Daily | Υ | |
| | including on mobile plant | | Ť | |
| 9. No | including on mobile plant ise and vibration mitigations | Frequency | Y/N/NA | General Comments |
| 9. No | | As required | | General Comments |
| 9.1 | se and vibration mitigations N/A | As | Y/N/NA | General Comments General Comments |
| 9.1 | se and vibration mitigations | As required | Y/N/NA | |
| 9.1 | Are defective plant parked up and not being used? aste management mitigations | As required Frequency | Y/N/NA Y Y/N/NA | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|---------------------------|-----------------|----------------------|----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be comp the end of each day. | leteddaily, stored in the | site file and u | ploaded to Data | station before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |



OPERATIONAL ENVIRONMENTAL MANGEMENT



| | RECYCLING PL | AN – WETHERILL PARK I | INS | SPECTION | I CHECK | KLIST Sale by Choice not by Chance |
|--------|---|---|-----|----------------|---------|------------------------------------|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 01.09.2 | 3 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | m.f | Stewart |
| | | | | | | |
| 1. Ge | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | oly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| | | | | | | |
| 2. Tra | iffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | Alll car spaces are free by employees and visito | from obstruction and maintained for u ors? | ise | Daily | Υ | |
| 2.3 | Vehicles are entering an | nd leaving the site in forward direction | ١. | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust r | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and be | procedures are being implemented eing used? | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outside eing used? | е | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage system | n? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra | ites have been inspected and clear of | | Monthly | N/A | |
| 5.5 | | aters, first flush devices and litter and are operating correctly. | | Monthly | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|--|------------|-----|------------|-----------------------|--------|--|--|
| Approved By: Date Issued: Version: Review Date: Author: Page Nul | | | | | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

Monthly

Monthly

Quarterly

(Mar, Jun,

Sep, Dec)

annually

(Jun, Dec)

Bi-

N/A

N/A

N/A

N/A

the build up of litter material

flush placement of grate upon refitment.

5.6

5.7

5.8

5.9

Site structires to be regularly checked for erosion and scouring

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

Treatment areas and structures will be regularly checked for





| | TEAT WETTERIEL FARR 11451 ECTION CITECREST SIMILOS | | | | | | | |
|---------|--|-------------------------------|-----------------|------------------------------------|--|--|--|--|
| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | | | | | |
| 6. Vei | min and pest management mitigations N/A | Frequency | Y/N/NA | General Comments | | | | |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | | | | | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | | | | | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | | | | | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | | | | | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Daily | Υ | | | | | |
| | including on mobile plant | | Ť | | | | | |
| 9. No | including on mobile plant ise and vibration mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 9. No | | As required | | General Comments | | | | |
| 9.1 | se and vibration mitigations N/A | As | Y/N/NA | General Comments General Comments | | | | |
| 9.1 | se and vibration mitigations | As required | Y/N/NA | | | | | |
| 9.1 | Are defective plant parked up and not being used? aste management mitigations | As required Frequency | Y/N/NA Y Y/N/NA | | | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|---------------------------|-----------------|----------------------|----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be comp the end of each day. | leteddaily, stored in the | site file and u | ploaded to Data | station before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| PLAN – WETHERILL PARK INSPECTION CHECKLIST On the chance | | | | | | | |
|--|--|---|---------|-------------|----------|-------------------------|--|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 03.10.23 | 3 | |
| Inspection Completed By: M.Stewart Sig | | | | gnature: | M.P | Stewart | |
| | | | | | | | |
| 1. Ge | neral Management and m | nitigations N/A | | Frequency | Y/N/NA | General Comments | |
| 1.2 | Employees and contract trained. | tors have been inducted and are s | uitably | As required | Υ | | |
| 1.3 | Plant and equipment be the start of the day? | eing used is in good working condi | tion at | Daily | Y | | |
| | | | | | | | |
| 2. Tra | iffic mitigations \Box N | N/A | | Frequency | Y/N/NA | General Comments | |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinate | or? | Daily | Υ | | |
| 2.2 | AllI car spaces are free from obstruction and maintained for use by employees and visitors? | | | Daily | Υ | | |
| 2.3 | Vehicles are entering a | nd leaving the site in forward dire | ction. | Daily | Υ | | |
| | | | | | | | |
| 3. Air | quality, odour and dust r | mitigations \square N/A | | Frequency | Y/N/NA | General Comments | |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | Daily | Υ | | |
| 3.2 | Good dust management procedures are implemented (outside the building): Sweeper working and being used? | | | Daily | Υ | | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check gener | al | Daily | Υ | | |
| | | | | | | | |
| 5. Sto | ormwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments | |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | | |
| 5.2 | Have storm water drains been inspected for any build up of sediment, debris, litter and vegetation within drainage system? | | | Monthly | N/A | | |
| 5.3 | If materials identified in stormwater drains, has it been removed? | | | Monthly | N/A | | |
| 5.4 | Inflow areas and pit gra | ates have been inspected and clea | r of | Monthly | N/A | | |
| 5.5 | | eaters, first flush devices and litter and are operating correctly. | • | Monthly | N/A | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | | |

Monthly

Monthly

Quarterly

(Mar, Jun,

Sep, Dec)

annually (Jun, Dec)

Bi-

N/A

N/A

N/A

N/A

the build up of litter material

flush placement of grate upon refitment.

Site structires to be regularly checked for erosion and scouring

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

Treatment areas and structures will be regularly checked for

5.6

5.7

5.8

5.9





| | TEAT WETTERIEL FARR 11451 ECTION CITECREST SIMILOS | | | | | | | |
|---------|--|-------------------------------|-----------------|------------------------------------|--|--|--|--|
| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | | | | | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | | | | | |
| 6. Vei | min and pest management mitigations N/A | Frequency | Y/N/NA | General Comments | | | | |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | | | | | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | | | | | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | | | | | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | | | | | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Daily | Υ | | | | | |
| | including on mobile plant | | Ť | | | | | |
| 9. No | including on mobile plant ise and vibration mitigations | Frequency | Y/N/NA | General Comments | | | | |
| 9. No | | As required | | General Comments | | | | |
| 9.1 | se and vibration mitigations N/A | As | Y/N/NA | General Comments General Comments | | | | |
| 9.1 | se and vibration mitigations | As required | Y/N/NA | | | | | |
| 9.1 | Are defective plant parked up and not being used? aste management mitigations | As required Frequency | Y/N/NA Y Y/N/NA | | | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|---------------------------|-----------------|----------------------|----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be comp the end of each day. | leteddaily, stored in the | site file and u | ploaded to Data | station before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| | RECYCLING PL | AN – WETHERILL PARK I | INS | SPECTION | I CHECK | KLIST Sale by Chance |
|--------|--|--|-----|----------------|----------|----------------------|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 01.11.2 | 3 |
| Inspe | ection Completed By: | M.Stewart | Si | ignature: | M.P | Stewart |
| 1. Ge | neral Management and m | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | oly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| 2. Tra | iffic mitigations | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free by employees and visito | from obstruction and maintained for u | se | Daily | Υ | |
| 2.3 | Vehicles are entering a | nd leaving the site in forward direction | | Daily | Υ | |
| 2 Air | anality adour and dust r | mitigations N/A | | Frequency | Y/N/NA | General Comments |
| 5. All | quality, odour and dust r | procedures are being implemented | | Daily | T/IV/IVA | General Comments |
| 3.1 | (inside building): Sweeper working and be | | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outside eing used? | е | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | ormwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | is been inspected for any build up of and vegetation within drainage system | ո? | Monthly | N/A | |
| 5.3 | If materials identified in stormwater drains, has it been removed? | | | Monthly | N/A | |
| 5.4 | Inflow areas and pit grates have been inspected and clear of litter / debris? | | | Monthly | N/A | |
| 5.5 | Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. | | | Monthly | N/A | |
| 5.6 | Site structires to be regularly checked for erosion and scouring | | | Monthly | N/A | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for aterial | | Monthly | N/A | |
| 5.8 | Remove grate and insp | ect internal walls and base. Remove ar | ıy | Quarterly | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

(Mar, Jun,

Sep, Dec)

annually

(Jun, Dec)

Bi-

N/A

N/A

5.9

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

flush placement of grate upon refitment.





| | TEAT WE ITERIEE I ARREITS | | | |
|---------|--|-------------------------------|-----------------|------------------------------------|
| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Vei | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Daily | Υ | |
| | including on mobile plant | | Ť | |
| 9. No | including on mobile plant ise and vibration mitigations | Frequency | Y/N/NA | General Comments |
| 9. No | | As required | | General Comments |
| 9.1 | se and vibration mitigations N/A | As | Y/N/NA | General Comments General Comments |
| 9.1 | se and vibration mitigations | As required | Y/N/NA | |
| 9.1 | Are defective plant parked up and not being used? aste management mitigations | As required Frequency | Y/N/NA Y Y/N/NA | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|---------------------------|-----------------|----------------------|----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be comp the end of each day. | leteddaily, stored in the | site file and u | ploaded to Data | station before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| • | RECYCLING PL | AN – WETHERILL PARK I | NS | SPECTION | I CHECK | (LIST oot by chance |
|--------|---|---|-------|--------------------------------------|----------|---------------------|
| Locat | ion: | reDirect – Wetherill Park | D | ate: | 01.12.23 | 3 |
| Inspe | ction Completed By: | M.Stewart | Si | gnature: | m.P | Stewart |
| 1 Ga | neral Management and m | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| | - | ors have been inducted and are suitab | ılv | As | | General Comments |
| 1.2 | trained. | ors have been madeled and are saltab | , ı y | required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| 2. Tra | ffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | Alll car spaces are free f by employees and visito | from obstruction and maintained for uors? | se | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust n | | | Frequency Daily | Y/N/NA | General Comments |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outside eing used? | 9 | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | s been inspected for any build up of and vegetation within drainage system | 1? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra litter / debris? | tes have been inspected and clear of | | Monthly | N/A | |
| 5.5 | | aters, first flush devices and litter and are operating correctly. | | Monthly | N/A | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scouring | g | Monthly | N/A | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for aterial | | Monthly | N/A | |
| 5.8 | collected sediment, deb | ect internal walls and base. Remove an oris, litter and vegetation. Inspect and lowing any removal of objects. Ensure | . | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|-------------------------|-----|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: Version: R | | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

Bi-

annually

(Jun, Dec)

N/A

5.9

flush placement of grate upon refitment.

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?





| | TEAT WE ITERIEE I ARREITS | | | |
|---------|--|-------------------------------|-----------------|------------------------------------|
| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Vei | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Daily | Υ | |
| | including on mobile plant | | Ť | |
| 9. No | including on mobile plant ise and vibration mitigations | Frequency | Y/N/NA | General Comments |
| 9. No | | As required | | General Comments |
| 9.1 | se and vibration mitigations N/A | As | Y/N/NA | General Comments General Comments |
| 9.1 | se and vibration mitigations | As required | Y/N/NA | |
| 9.1 | Are defective plant parked up and not being used? aste management mitigations | As required Frequency | Y/N/NA Y Y/N/NA | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | | |
|--|------------------------------|-----------------|----------------------|--------------|--|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| torage and Reference | Inspection Comple | ted By | | Date | | | | |
| o be reviewed at Site Meeting. | | | | | | | | |
| Vorkplace inspection checklists must be co | ompleteddaily, stored in the | site file and u | ploaded to Data | station hefo | | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| • | RECYCLING PL | AN – WETHERILL PARK I | NS | SPECTION | I CHECK | (LIST oot by chance |
|--------|--|--|----|--------------------------------------|-----------|---------------------|
| Locat | ion: | reDirect – Wetherill Park | D | ate: | 08.01.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | m.P | Stewart |
| | | | | | | |
| 1. Ge | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | ly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| 2. Tra | ffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free to | from obstruction and maintained for usors? | se | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | | Daily | Υ | |
| | | | | _ | v/10.10.0 | |
| 3. Air | quality, odour and dust r | - | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | Daily | Υ | |
| 3.2 | Good dust management procedures are implemented (outside the building): Sweeper working and being used? | | | | Υ | |
| 3.3 | Residual waste has beer waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage system | 1? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | n stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit grates have been inspected and clear of litter / debris? | | | Monthly | N/A | |
| 5.5 | Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. | | | Monthly | N/A | |
| 5.6 | 6 Site structires to be regularly checked for erosion and scouring | | | Monthly | N/A | |
| 5.7 | Treatment areas and structures will be regularly checked for the build up of litter material | | | Monthly | N/A | |
| 5.8 | collected sediment, del | ect internal walls and base. Remove an oris, litter and vegetation. Inspect and lowing any removal of objects. Ensure e upon refitment. | | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

Bi-

annually (Jun, Dec) N/A

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

5.9





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments | | | |
|---------------|--|---------------------------------|----------------------|------------------------------------|--|--|--|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | | | | |
| 6. Ve | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments | | | |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | | | | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | | | | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | | | | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments | | | |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | | | | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | | | | |
| | | | | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments | | | |
| 8. Fire | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Y/N/NA Y | General Comments | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | | | General Comments General Comments | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations N/A | Prequency As | Y Y/N/NA | | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations Are defective plant parked up and not being used? | Frequency As required | Y | General Comments | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y Y/N/NA Y Y/N/NA | General Comments | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|--|------------------------------|-----------------|----------------------|--------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| torage and Reference | Inspection Comple | ted By | | Date | | | |
| o be reviewed at Site Meeting. | | | | | | | |
| Vorkplace inspection checklists must be co | ompleteddaily, stored in the | site file and u | ploaded to Data | station hefo | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| • | RECYCLING PL | AN – WETHERILL PARK I | NS | SPECTION | I CHECK | (LIST not by chance |
|--|--|--|----------|--------------------------------------|---------|---------------------|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 01.02.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | m.f | Stewart |
| 1 Go | neral Management and m | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| | - | cors have been inducted and are suitab | lv | As | | General Comments |
| 1.2 | trained. | | required | Υ | | |
| 1.3 Plant and equipment being used is in good working condition at the start of the day? | | | | | Y | |
| 2. Tra | iffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 Alll car spaces are free from obstruction and maintained for use by employees and visitors? | | | | | Υ | |
| 2.3 | Vehicles are entering a | nd leaving the site in forward direction | | Daily | Υ | |
| 3. Air | quality, odour and dust r | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | Daily | Υ | | |
| 3.2 | Good dust management procedures are implemented (outside the building): Sweeper working and being used? | | | | Υ | |
| 3.3 | Residual waste has beer waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | is been inspected for any build up of and vegetation within drainage system | າ? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | n stormwater drains, has it been | | Monthly | N/A | |
| Inflow areas and pit grates have been inspected and clear of litter / debris? | | | Monthly | N/A | | |
| 5.5 | 5.5 Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. | | | Monthly | N/A | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scouring | g | Monthly | N/A | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for aterial | | Monthly | N/A | |
| 5.8 | collected sediment, del | ect internal walls and base. Remove ar oris, litter and vegetation. Inspect and lowing any removal of objects. Ensure e upon refitment. | | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

Bi-

annually

(Jun, Dec)

N/A

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

5.9





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments | | | |
|---------------|--|---------------------------------|----------------------|------------------------------------|--|--|--|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | | | | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | | | | |
| 6. Ve | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments | | | |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | | | | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | | | | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | | | | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments | | | |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | | | | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | | | | |
| | | | | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments | | | |
| 8. Fire | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Y/N/NA Y | General Comments | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | | | General Comments General Comments | | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations N/A | Prequency As | Y Y/N/NA | | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations Are defective plant parked up and not being used? | Frequency As required | Y | General Comments | | | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y Y/N/NA Y Y/N/NA | General Comments | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | |





| Action Pla | n - to be transferred as a 'Ha | zard Report' | | |
|--|--------------------------------|-----------------|----------------------|--------------|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| torage and Reference | Inspection Comple | ted By | | Date |
| o be reviewed at Site Meeting. | | | | |
| Vorkplace inspection checklists must be co | ompleteddaily, stored in the | site file and u | ploaded to Data | station hefo |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | |





| • | RECYCLING PL | AN – WETHERILL PARK I | NS | SPECTION | I CHECK | (LIST not by chance |
|--------|--|--|----|--------------------------------------|---------|---------------------|
| Locat | ion: | reDirect – Wetherill Park | D | ate: | 20.02.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | m.P | Stewart |
| 1. Gei | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | ly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| 2. Tra | ffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | nitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free f | rom obstruction and maintained for uors? | se | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust r | | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and be | procedures are being implemented eing used? | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outside eing used? | 9 | Daily | Υ | |
| 3.3 | | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | s been inspected for any build up of and vegetation within drainage system | ո? | Monthly | N/A | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra litter / debris? | tes have been inspected and clear of | | Monthly | N/A | |
| 5.5 | 1 | aters, first flush devices and litter and are operating correctly. | | Monthly | N/A | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scouring | g | Monthly | N/A | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for iterial | | Monthly | N/A | |
| 5.8 | collected sediment, det | ect internal walls and base. Remove an oris, litter and vegetation. Inspect and lowing any removal of objects. Ensure e upon refitment. | | Quarterly (Mar, Jun, Sep, Dec) | N/A | |
| 5.9 | Have all drainage struct | ures been inspected noting any | | Bi- | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | |

N/A

annually (Jun, Dec)

dilapidation, if so have repairs been carried out?





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|---------------|--|---------------------------------|----------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ve | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8. Fire | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Y/N/NA Y | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | | | General Comments General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations N/A | Prequency As | Y Y/N/NA | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations Are defective plant parked up and not being used? | Frequency As required | Y | General Comments |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y Y/N/NA Y Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | |





| Action Pla | n - to be transferred as a 'Ha | zard Report' | | |
|--|--------------------------------|-----------------|----------------------|--------------|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| torage and Reference | Inspection Comple | ted By | | Date |
| o be reviewed at Site Meeting. | | | | |
| Vorkplace inspection checklists must be co | ompleteddaily, stored in the | site file and u | ploaded to Data | station hefo |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |



OPERATIONAL ENVIRONMENTAL MANGEMENT



| • | RECYCLING PL | AN – WETHERILL PARK | INS | SPECTION | I CHEC | (LIST oot by chance |
|---------------|---|-------------------------------------|----------|-------------------------|----------|---------------------|
| Location: rel | | reDirect – Wetherill Park Da | | ate: | 01.03.24 | |
| Inspe | M.Stewart | Si | gnature: | M.P | Stewart | |
| | | | | | | |
| 1. Ge | neral Management and m | | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contractors have been inducted and are suitably trained. | | | As required | Υ | |
| 1.3 | Plant and equipment being used is in good working condition at the start of the day? | | | | Y | |
| 2. Tra | iffic mitigations \Box N | N/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free from obstruction and maintained for use by employees and visitors? | | | Daily | Υ | |
| 2.3 | Vehicles are entering and leaving the site in forward direction. | | | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust r | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | Daily | Υ | |
| 3.2 | Good dust management procedures are implemented (outside the building): Sweeper working and being used? | | | Daily | Υ | |
| 3.3 | Residual waste has been transported offsite (check general waste bin capacity)? | | | | Υ | |
| | | | | | | |
| 5. Sto | ormwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | Have storm water drains been inspected for any build up of sediment, debris, litter and vegetation within drainage system? | | | Monthly | N/A | |
| 5.3 | If materials identified in stormwater drains, has it been removed? | | | Monthly | N/A | |
| 5.4 | Inflow areas and pit grates have been inspected and clear of litter / debris? | | | Monthly | N/A | |
| 5.5 | Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. | | | Monthly | N/A | |
| 5.6 | Site structires to be regularly checked for erosion and scouring | | | Monthly | N/A | |
| 5.7 | Treatment areas and structures will be regularly checked for the build up of litter material | | | Monthly | N/A | |
| 5.8 | Remove grate and inspect internal walls and base. Remove any collected sediment, debris, litter and vegetation. Inspect and | | | Quarterly (Mar, Jun, | NI/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

N/A

N/A

Sep, Dec)

annually

(Jun, Dec)

Bi-

5.9

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

flush placement of grate upon refitment.





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|-------------------------------------|---|---------------------------------|----------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ve | rmin and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pollution management mitigations | | | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| a =: | | | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8. Fire | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Y/N/NA Y | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | | | General Comments General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations N/A | Prequency As | Y Y/N/NA | |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations N/A Are defective plant parked up and not being used? | Frequency As required | Y | General Comments |
| 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y Y/N/NA Y Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | N/A | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | N/A | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | |
|---|-----------------------|-----------------|--|-----------------|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | <u> </u> | | | |
| Storage and Reference | Inspection Comple | eted By | | Date | | |
| To be reviewed at Site Meeting. | | | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | | |





| | RECYCLING PL | AN – WETHERILL PARK | INS | SPECTION | I CHEC | KLIST ont by chance |
|--|--|---|----------------|-----------|---------|---------------------|
| Locat | ion: | reDirect – Wetherill Park | D | ate: | 29.03.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | M.F | Stewart |
| 1. Ge | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | As required | Υ | | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| | , | | | | | |
| 2. Tra | ffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | Alll car spaces are free to by employees and visitor | from obstruction and maintained for uprs? | ise | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | ۱. | Daily | Υ | |
| | | | | | | |
| 3. Air quality, odour and dust mitigations N/A | | | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | t procedures are implemented (outside eing used? | е | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage systen | n? | Monthly | Υ | |
| 5.3 | 3 If materials identified in stormwater drains, has it been removed? | | | Monthly | Y | |
| 5.4 | Inflow areas and pit gra litter / debris? | Monthly | Y | | | |
| 5.5 | | eaters, first flush devices and litter and are operating correctly. | | Monthly | Y | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scourin | g | Monthly | Y | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for aterial | | Monthly | Y | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | | |

Quarterly

(Mar, Jun,

Sep, Dec)

annually

(Jun, Dec)

Bi-

Y Dec

Y Dec

2022

2022

Lift grate, brush out lip for grate

and down walls remove debris

Inspected no action required

replace grate

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

flush placement of grate upon refitment.

5.8

5.9





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|------------------------|--|---------------------------------|-----------------|-------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | Y Dec 2022 | Check Basket – no litter |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | Y Dec 2022 | Empty tank inspect no sign of pests |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | Y Dec 2022 | No repairs required |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | Y Dec 2022 | Checked no action required |
| 6. Ver | rmin and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| - | | Daily | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | |
| | | Frequency | Y Y/N/NA | General Comments |
| 9. Noi | including on mobile plant | | | General Comments |
| 9. Noi | including on mobile plant ise and vibration mitigations | Frequency As | Y/N/NA | General Comments General Comments |
| 9. Noi 9.1 10. W | including on mobile plant ise and vibration mitigations N/A Are defective plant parked up and not being used? | Frequency As required | Y/N/NA | |
| 9.1 | including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y/N/NA Y Y/N/NA | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | | |





| 11. Flooding mitigations N/A | | | Y/N/NA | General Comments |
|--------------------------------|---|-------------------------------|---------------|-------------------------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | Y Dec 2022 | Fully stocked and in good condition |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | Y Dec 2022 | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|---------------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | Y Dec 2022 | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|--|-----------------------|-----------------|----------------------|-----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| Chause and Defen | | to d D | | Dete | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be completed the end of each day. | ddaily, stored in the | site file and u | oloaded to Dat | astation before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| • | RECYCLING PL | AN – WETHERILL PARK | INS | SPECTION | I CHECK | (LIST not by chance |
|--------|--|--|-----|-------------|----------|---------------------|
| Locat | ion: | reDirect – Wetherill Park | D | ate: | 30.04.24 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | M.P | Stewart |
| 1. Gei | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suita | bly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| 2. Tra | ffic mitigations | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | All car spaces are free from obstruction and maintained for use | | | Daily | Υ | |
| 2.3 | Vehicles are entering a | nd leaving the site in forward directio | n. | Daily | Υ | |
| | | | | | | |
| 3. Air | quality, odour and dust r | mitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and be | t procedures are being implemented eing used? | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and bo | t procedures are implemented (outsion eing used? | le | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills tha | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage syste | m? | Monthly | Υ | |
| 5.3 | If materials identified in removed? | n stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra litter / debris? | ites have been inspected and clear of | | Monthly | Y | |
| 5.5 | | eaters, first flush devices and litter and are operating correctly. | | Monthly | Y | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

Monthly

Monthly

Quarterly

(Mar, Jun,

Sep, Dec)

annually (Jun, Dec)

Bi-

Y

Y

Y

N/A

the build up of litter material

flush placement of grate upon refitment.

Site structires to be regularly checked for erosion and scouring

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

Treatment areas and structures will be regularly checked for

5.6

5.7

5.8

5.9





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|---------------|---|---------------------------------------|-------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ver | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As | Υ | |
| | | required | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8. Fire | e management mitigations N/A Fire extinguishers are positioned at readily accessible points, including on mobile plant | | Y/N/NA Y | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Frequency | | General Comments General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Υ | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations N/A | Frequency Daily Frequency As | Y Y/N/NA | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations Are defective plant parked up and not being used? | Frequency Daily Frequency As required | Y/N/NA N/A | General Comments |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations | Frequency As required Frequency | Y/N/NA N/A Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | | |

| 12. Bi | odiversity 🗆 N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|-----------------------|-----------------|--|-----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | <u> </u> | | | | |
| Storage and Reference | Inspection Comple | eted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| | RECYCLING PL | AN - WETHERILL PARK | IINS | PECHON | CHECK | (LIST |
|--------|--|--|------|--------------------------------------|---------|------------------|
| Locat | ion: | reDirect – Wetherill Park | Da | ate: | 30.11.2 | 2 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | M.P | Stewart |
| 1. Gei | neral Management and m | nitigations □ N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | - | ors have been inducted and are suitab | oly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| | | | | | | |
| 2. Tra | ffic mitigations \square N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | Alll car spaces are free f | rom obstruction and maintained for uors? | ise | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | ١. | Daily | Υ | |
| 3. Air | quality, odour and dust n | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and be | procedures are being implemented | | Daily | Υ | |
| 3.2 | Good dust management procedures are implemented (outside | | | Daily | Υ | |
| 3.3 | Residual waste has been transported offsite (check general | | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | have been left unattended? | | Daily | N | |
| 5.2 | | s been inspected for any build up of and vegetation within drainage systen | n? | Monthly | Υ | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra litter / debris? | tes have been inspected and clear of | | Monthly | Y | |
| 5.5 | 1 | aters, first flush devices and litter and are operating correctly. | | Monthly | Y | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scourin | g | Monthly | Y | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for Iterial | | Monthly | Y | |
| 5.8 | | | | Quarterly (Mar, Jun, Sep, Dec) | Y | |
| 5.9 | _ | ures been inspected noting any repairs been carried out? | | Bi- annually (Jun, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|-------------------------|-----|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: Version: F | | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|---------------|---|---------------------------------------|-------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ver | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As | Υ | |
| | | required | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8. Fire | e management mitigations N/A Fire extinguishers are positioned at readily accessible points, including on mobile plant | | Y/N/NA Y | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Frequency | | General Comments General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Υ | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations N/A | Frequency Daily Frequency As | Y Y/N/NA | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations Are defective plant parked up and not being used? | Frequency Daily Frequency As required | Y/N/NA N/A | General Comments |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations | Frequency As required Frequency | Y/N/NA N/A Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | | |

| 12. Bi | odiversity 🗆 N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | | |
|---|-----------------------|-----------------|--|-----------------|--|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | <u> </u> | | | | |
| Storage and Reference | Inspection Comple | eted By | | Date | | | |
| To be reviewed at Site Meeting. | | | | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| | RECYCLING PL | AN – WETHERILL PARK I | NS | PECTION | I CHEC | (LIST not by chance |
|--|---|--|----|-------------|---------|----------------------------|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 28.06.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | MI | Stewart |
| 1. Ge | neral Management and m | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | ly | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition a | at | Daily | Y | |
| 2. Tra | iffic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free by employees and visito | from obstruction and maintained for u ors? | se | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | | Daily | Υ | |
| | | | | | | |
| 3. Air quality, odour and dust mitigations \square N/A | | | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | | Υ | |
| 3.2 | Good dust management the building): Sweeper working and bo | t procedures are implemented (outside eing used? | 9 | Daily | Υ | |
| 3.3 | Residual waste has beer waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | ormwater mitigations |] N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | ns been inspected for any build up of and vegetation within drainage system | ո? | Monthly | Υ | |
| 5.3 | If materials identified in stormwater drains, has it been removed? | | | Monthly | N/A | |
| 5.4 | Inflow areas and pit gra | Monthly | Y | | | |
| 5.5 | | eaters, first flush devices and litter and are operating correctly. | | Monthly | Y | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scouring | g | Monthly | Y | |
| 5.7 | Treatment areas and st | ructures will be regularly checked for | | Monthly | V | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | | |

Quarterly

(Mar, Jun,

Sep, Dec)

annually

(Jun, Dec)

Bi-

Y

N/A

the build up of litter material

flush placement of grate upon refitment.

Remove grate and inspect internal walls and base. Remove any

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

5.8

5.9





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|---------------|---|---------------------------------------|-------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ver | min and pest management mitigations \[\subseteq N/A \] | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As | Υ | |
| | | required | | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8. Fire | e management mitigations N/A Fire extinguishers are positioned at readily accessible points, including on mobile plant | | Y/N/NA Y | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, | Frequency | | General Comments General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Frequency Daily | Υ | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations N/A | Frequency Daily Frequency As | Y Y/N/NA | |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations Are defective plant parked up and not being used? | Frequency Daily Frequency As required | Y/N/NA N/A | General Comments |
| 9. Noi 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant se and vibration mitigations | Frequency As required Frequency | Y/N/NA N/A Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | | |

| 12. Bi | odiversity 🗆 N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | | |
|---|-----------------------|-----------------|--|-----------------|--|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | <u> </u> | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | | |
| To be reviewed at Site Meeting. | | | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |





| | RECYCLING | AN – WETHERILL PARK | INS | SPECTION | CHECK | (LIST not by chance |
|--------|--|---|------|-------------|--------------|-------------------------|
| Loca | tion: | reDirect – Wetherill Park | D | ate: | 28.06.2 | 4 |
| Inspe | Inspection Completed By: M.Stewart | | Si | ignature: | M.P. Stewart | |
| 1. Ge | neral Management and m | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suita | ably | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | n at | Daily | Y | |
| 2. Tra | affic mitigations \Box N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | AllI car spaces are free to | from obstruction and maintained for ors? | use | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | n. | Daily | Υ | |
| 3. Air | quality, odour and dust r | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 3.1 | Good dust management (inside building): Sweeper working and be | procedures are being implemented eing used? | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outsi | de | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | ormwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | s been inspected for any build up of and vegetation within drainage syste | | Monthly | Υ | |

| 5. Sto | rmwater mitigations $\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \$ | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|---------------|---|
| 5.1 | Are there any spills that have been left unattended? | Daily | N | |
| 5.2 | Have storm water drains been inspected for any build up of sediment, debris, litter and vegetation within drainage system? | Monthly | Υ | |
| 5.3 | If materials identified in stormwater drains, has it been removed? | Monthly | Y | |
| 5.4 | Inflow areas and pit grates have been inspected and clear of litter / debris? | Monthly | Y | |
| 5.5 | Ensure downpipe leaf eaters, first flush devices and litter screens are unblocked and are operating correctly. | Monthly | Y | |
| 5.6 | Site structires to be regularly checked for erosion and scouring | Monthly | Y | |
| 5.7 | Treatment areas and structures will be regularly checked for the build up of litter material | Monthly | Y | |
| 5.8 | Remove grate and inspect internal walls and base. Remove any collected sediment, debris, litter and vegetation. Inspect and ensure grate is clear following any removal of objects. Ensure flush placement of grate upon refitment. | Quarterly (Mar, Jun, Sep, Dec) | Y Dec 2022 | Lift grate,brush out lip for grate and down walls remove debris replace grate |
| 5.9 | Have all drainage structures been inspected noting any dilapidation, if so have repairs been carried out? | Bi- annually (Jun, Dec) | Y Dec 2022 | Inspected no action required |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | |





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|------------------------|--|---------------------------------|-----------------|-------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | Y Dec 2022 | Check Basket – no litter |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | Y Dec 2022 | Empty tank inspect no sign of pests |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | Y Dec 2022 | No repairs required |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | Y Dec 2022 | Checked no action required |
| 6. Ver | rmin and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| - | | Daily | | |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Υ | |
| | | Frequency | Y Y/N/NA | General Comments |
| 9. Noi | including on mobile plant | | | General Comments |
| 9. Noi | including on mobile plant ise and vibration mitigations | Frequency As | Y/N/NA | General Comments General Comments |
| 9. Noi 9.1 10. W | including on mobile plant ise and vibration mitigations N/A Are defective plant parked up and not being used? | Frequency As required | Y/N/NA | |
| 9.1 | including on mobile plant ise and vibration mitigations | Frequency As required Frequency | Y/N/NA Y Y/N/NA | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|---------------|-------------------------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | Y Dec 2022 | Fully stocked and in good condition |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | Y Dec 2022 | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|---------------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | Y Dec 2022 | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | |
|---|-----------------------|-----------------|--|-----------------|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | <u> </u> | | |
| Storage and Reference | Inspection Comple | ted By | | Date | |
| To be reviewed at Site Meeting. | | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | | |





| | RECYCLING PL | AN – WETHERILL PARK | INS | SPECTION | I CHECK | (LIST not by chance |
|--|---|---|-----------|----------------|------------------|---------------------|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 30.07.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | M.P | Stewart |
| 1. Ge | neral Management and m | nitigations | | Frequency | Y/N/NA | General Comments |
| Employees and contractors have been inducted and are suitably As | | | | As required | Υ | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| | | | | _ | | |
| 2. Tra | iffic mitigations \square N | I/A | | Frequency | Y/N/NA | General Comments |
| 2.1 Traffic is continually monitored by Operations Coordinator? | | | | Daily | Υ | |
| 2.2 | Alll car spaces are free to by employees and visito | from obstruction and maintained for uprs? | ıse | Daily | Υ | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | ٦. | Daily | Υ | |
| | | _ | | | | |
| | | | Frequency | Y/N/NA | General Comments | |
| 3.1 | Good dust management procedures are being implemented (inside building): Sweeper working and being used? | | | Daily | Υ | |
| 3.2 | Good dust management the building): Sweeper working and be | procedures are implemented (outside ingused? | е | Daily | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | is been inspected for any build up of and vegetation within drainage syster | n? | Monthly | Υ | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | N/A | |
| 5.4 | Inflow areas and pit grates have been inspected and clear of litter / debris? | | | Monthly | Y | |
| 5.5 | screens are unblocked and are operating correctly. | | | Monthly | Y | |
| 5.6 | Site structires to be reg | ularly checked for erosion and scourir | ng | Monthly | Y | |
| 5.7 | Treatment areas and st the build up of litter ma | ructures will be regularly checked for aterial | | Monthly | Y | |
| 5.8 | Remove grate and inspe | ect internal walls and base. Remove a | ny | Quarterly | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | |

(Mar, Jun,

Sep, Dec)

annually

(Jun, Dec)

Bi-

Y

N/A

5.9

collected sediment, debris, litter and vegetation. Inspect and

ensure grate is clear following any removal of objects. Ensure

Have all drainage structures been inspected noting any

dilapidation, if so have repairs been carried out?

flush placement of grate upon refitment.





| 5. Sto | rmwater mitigations | Frequency | Y/N/NA | General Comments |
|--------------------------------|--|---|----------------------------|------------------------------------|
| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | N/A | |
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | N/A | |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | N/A | |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | N/A | |
| 6. Ve | rmin and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations □ N/A | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management | As | | |
| - · - | plan been provided in toolbox? | required | Υ | |
| | plan been provided in toolbox? e management mitigations N/A | | Y Y/N/NA | General Comments |
| | · · · | required | | General Comments |
| 8. Fire | e management mitigations | required Frequency | Y/N/NA | General Comments General Comments |
| 8. Fire | e management mitigations | Frequency Daily | Y/N/NA Y | |
| 8. Fire 8.1 9. No | e management mitigations | Frequency Daily Frequency As | Y/N/NA Y Y/N/NA | |
| 8. Fire 8.1 9. No | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency Daily Frequency As required | Y/N/NA Y Y/N/NA N/A | General Comments |
| 8. Fire 8.1 9. No 9.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant ise and vibration mitigations | Frequency Daily Frequency As required Frequency | Y/N/NA Y Y/N/NA N/A Y/N/NA | General Comments |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 11. Flo | ooding mitigations | Frequency | Y/N/NA | General Comments |
|---------|---|-------------------------------|--------|------------------|
| 11.1 | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | | |
| 11.2 | Yearly (at minimum) evacuation drills will be implemented as part of ongoing training onsite. | Yearly | | |

| 12. Bi | odiversity N/A | Frequency | Y/N/NA | General Comments |
|---------------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | N/A | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | | |





| Action Plan - to be transferred as a 'Hazard Report' | | | | | |
|--|------------------------------|-----------------|----------------------|---------------|--|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| Storage and Reference | Inspection Comple | ted By | | Date | |
| To be reviewed at Site Meeting. | | | | | |
| Workplace inspection checklists must be co | ompleteddaily, stored in the | site file and u | ploaded to Data | station befor | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | | |

Appendix B: Field Sheets



Monitoring Round: S20102_06 Feb 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW2

Arrival Date/Time 06/02/2023 09:54AM Departure Date/Time 08/02/2023 09:04AM

Executed By Bec Chapple

Weather Sunny

Comments

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:04 PM

Groundwater Data

Well MW2 Date/Time 06/02/2023 09:54AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 2.96
 Well Depth (mbTOC)
 7.59

Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW2 Date/Time 08/02/2023 08:31AM

Well Depth (mbTOC)

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.957

Product Depth

(mbTOC)

Comments & Product

Description
Equipment ID

Equipment ib

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 08/02/2023 08:43AM

Well MW2
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW2

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 3.122

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 08:42AM | 21545 | 6.33 | 154.4 | 1.35 | 20.8 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_06 Feb 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW4

Arrival Date/Time 06/02/2023 10:07AM Departure Date/Time 08/02/2023 10:44AM

Executed By Bec Chapple

Weather Sunny

Comments

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:05 PM

Groundwater Data

Well MW4 Date/Time 06/02/2023 10:08AM

Measurement MethodDipDryNoWater Depth (mbTOC)2.205Well Depth (mbTOC)6.99

Water Depth (mbTOC) 2.205 Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW4 Date/Time 08/02/2023 10:24AM

Well Depth (mbTOC)

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.224

Product Depth

(mbTOC)

Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 08/02/2023 10:38AM

Well MW4
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW4

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 2.554

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 10:37AM | 17881 | 6.54 | 69 | 0 | 22.7 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_06 Feb 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW6

Arrival Date/Time 06/02/2023 10:19AM Departure Date/Time 08/02/2023 11:09AM

Executed By Bec Chapple

Weather Sunny

Comments Stick up well

Well Information

Gatic Type Stick up

Key Type None

Well Condition Average

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:05 PM

Groundwater Data

Well MW6 Date/Time 06/02/2023 10:19AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 4.444
 Well Depth (mbTOC)
 7.19

Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW6 Date/Time 08/02/2023 10:52AM

Measurement Method Dip Dry No

Water Depth (mbTOC) 4.444 Well Depth (mbTOC)

Product Depth

(mbTOC)
Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 08/02/2023 10:53AM

Well MW6

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW6

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Moderately turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 4.446

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 10:53AM | 2323 | 7.19 | 89.8 | 0.1 | 22.7 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_06 Feb 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW3

Arrival Date/Time 06/02/2023 10:30AM Departure Date/Time 08/02/2023 10:10AM

Executed By Bec Chapple

Weather Sunny

Comments

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:06 PM

Groundwater Data

Well MW3 Date/Time 06/02/2023 10:31AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 2.971
 Well Depth (mbTOC)
 8.08

Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW3 Date/Time 08/02/2023 09:37AM

Well Depth (mbTOC)

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.945

Product Depth

(mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 08/02/2023 09:44AM

Well MW3

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW3

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description)colourlessSample Odour (Description)no odourSample Sheen (Description)no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1) QC101

QA Sample ID (2) QC201

QA Sample ID (3) QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 3.3

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 09:45AM | 29765 | 5.78 | 65.7 | 0.79 | 23.6 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_06 Feb 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW1

Arrival Date/Time 06/02/2023 10:37AM Departure Date/Time 08/02/2023 09:26AM

Executed By Bec Chapple

Weather Sunny

Comments No x-cap in place.

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:06 PM

Groundwater Data

Well MW1 Date/Time 06/02/2023 10:41AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 2.145
 Well Depth (mbTOC)
 6.61

Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW1 Date/Time 08/02/2023 09:05AM

Well Depth (mbTOC)

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.111

Product Depth

(mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 08/02/2023 09:06AM

Well MW1

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW1

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) orange
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 2.163

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 09:06AM | 22382 | 6.47 | 28.3 | 2.02 | 22.4 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_10 Feb 2023 _1

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW2

Arrival Date/Time 10/02/2023 08:16AM Departure Date/Time 10/02/2023 08:17AM

Executed By Bec Chapple

Weather Sunny

Comments

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:07 PM

Sample Data

Sampled Date/Time 10/02/2023 08:17AM

Field ID SW2

Sample Depth From

(m)

Sample Depth To

Sample Type Normal

Sample Comments

Matrix Type Water

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Moderately turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Field Chemistry

Temp 22.2 oC (-)

DO 3.85 mg/L (-)

EC 365.7 uS/cm (-)

pH 7.04 - (-)

Redox 215.3 mV (-)



Monitoring Round: S20102_10 Feb 2023 _1

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW1

Arrival Date/Time 10/02/2023 08:40AM Departure Date/Time 10/02/2023 08:42AM

Executed By Bec Chapple

Weather Sunny

Comments

Authorisation

Checked By Hayley Yellowlees

Date Checked 09 Aug 2024 03:07 PM

Sample Data

Sampled Date/Time 10/02/2023 08:42AM

Field ID SW1

Sample Depth From

(m)

Sample Depth To

Sample Type Normal

Sample Comments

Matrix Type Water

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Moderately turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Field Chemistry

Temp 21.8 oC (-)

DO 4.23 mg/L (-)

EC 196.2 uS/cm (-)

pH 7.54 - (-)

Redox 219.6 mV (-)



Monitoring Round: \$20102_02 Aug 2023

Departure Date/Time 14/08/2023 04:01PM

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW6

Arrival Date/Time 02/08/2023 10:33AM

Executed By Hayley Yellowlees

Weather Sunny

Comments

Well Information

Gatic Type monument

Key Type None
Well Condition Good

Authorisation

Checked By Bec Chapple

Date Checked 15 Aug 2023 02:15 PM

Groundwater Data

Well MW6 Date/Time 02/08/2023 10:34AM

Measurement MethodDipDryNoWater Depth (mbTOC)4.748Well Depth (mbTOC)7.17

Water Depth (mbTOC) 4.748 Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 14/08/2023 03:42PM

Well MW6

Matrix Type Water

Equipment ID

Sample Comments Pale yellow, slightly sulphurous, yellow/orange sediment in bottom of hydrasleeve.

Field ID (Primary) MW6

Purge Method

Sample Method Snap Sampler

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) yellow

Sample Odour (Description) sulphurous odour

Sample Sheen (Description) no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 4.825

Field Chemistry

| | Standing Water Level | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|-------------------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | m bTOC | uS/cm | pH Units | mV | mg/L | ° C |
| 03:41PM | 4.765 | 1362 | 7.22 | -6.4 | 0.81 | 16.8 |
| Stabilisation * | | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_02 Aug 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW3

Arrival Date/Time 02/08/2023 11:03AM Departure Date/Time 14/08/2023 04:46PM

Executed By Hayley Yellowlees

Weather Sunny

Comments

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 15 Aug 2023 02:21 PM

Groundwater Data

Well MW3 Date/Time 02/08/2023 11:04AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 3.599
 Well Depth (mbTOC)
 8.07

Product Depth

(mbTOC)

Comments & Product

Description

Hydrasleeves installed x2

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 14/08/2023 04:13PM

Well MW3

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW3

Purge Method

Sample Method

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 3.985

Field Chemistry

| | Standing Water Level | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|-------------------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | m bTOC | uS/cm | pH Units | mV | mg/L | ° C |
| 04:13PM | 3.425 | 24992 | 5.91 | 62.6 | 0.92 | 15.9 |
| Stabilisation * | | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_02 Aug 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW1

Arrival Date/Time 02/08/2023 11:26AM Departure Date/Time 14/08/2023 01:34PM

Executed By Hayley Yellowlees

Weather Overcast

Comments No x-cap. PVC too close to the gatic.

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 15 Aug 2023 02:23 PM

Groundwater Data

Well MW1 Date/Time 02/08/2023 11:28AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 2.634
 Well Depth (mbTOC)
 6.61

Product Depth (mbTOC)

Comments & Product

Description

Hydra sleeve installed

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 14/08/2023 01:17PM

Well MW1

Matrix Type Water

Equipment ID

Sample Comments Orange sediment at bottom of hydrasleeve

Field ID (Primary) MW01

Purge Method

Sample Method Snap Sampler

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless, orange

Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Non-turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 2.575

Field Chemistry

| | Standing Water Level | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|-------------------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | m bTOC | uS/cm | pH Units | mV | mg/L | ° C |
| 01:17PM | 2.575 | 19738 | 6.34 | 116.4 | 6.33 | 17 |
| Stabilisation * | | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_02 Aug 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW2

Arrival Date/Time 02/08/2023 11:43AM Departure Date/Time 14/08/2023 02:38PM

Executed By Hayley Yellowlees

Weather Overcast

Comments Ants nest in top of well

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 15 Aug 2023 02:36 PM

Groundwater Data

Well MW2 Date/Time 02/08/2023 11:43AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 3.19
 Well Depth (mbTOC)
 7.57

Water Depth (mbTOC) 3.19 Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 14/08/2023 02:22PM

Well MW2
Matrix Type Water

Equipment ID

Sample Comments Orange sediment in bottom of hydrasleeve

Field ID (Primary) MW2

Purge Method

Sample Method Snap Sampler

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 3.484

Field Chemistry

| | Standing Water Level | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|-------------------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | m bTOC | uS/cm | pH Units | mV | mg/L | ° C |
| 02:22PM | 3.155 | 17006 | 6.44 | 129 | 1.2 | 17 |
| Stabilisation * | | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_02 Aug 2023

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW4

Arrival Date/Time 02/08/2023 11:59AM Departure Date/Time 14/08/2023 03:36PM

Executed By Hayley Yellowlees

Weather Overcast

Comments

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 15 Aug 2023 02:36 PM

Groundwater Data

Well MW4 Date/Time 02/08/2023 12:01PM

Measurement MethodDipDryNoWater Depth (mbTOC)2.565Well Depth (mbTOC)6.98

Water Depth (mbTOC) 2.565 Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sampled Date/Time 14/08/2023 03:26PM

Well MW4
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW4

Purge Method

Sample Method Snap Sampler

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Non-turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m) 2.925

Field Chemistry

| | Standing Water Level | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|-------------------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | m bTOC | uS/cm | pH Units | mV | mg/L | °C |
| 03:25PM | 2.62 | 7133 | 6.55 | 18.7 | 1.7 | 17.1 |
| Stabilisation * | | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW6

Arrival Date/Time 07/02/2024 11:21AM Departure Date/Time 09/02/2024 11:26AM

Executed By Rowan Faint

Weather Sunny

Comments Stick up. Hydrasleeve installed

Well Information

Gatic Type Stick up

Key Type None

Well Condition Good

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:19 PM

Groundwater Data

Well MW6 Date/Time 07/02/2024 11:22AM

Measurement MethodDipDryNoWater Depth (mbTOC)4.369Well Depth (mbTOC)7.16

Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW6 Date/Time 07/02/2024 11:30AM

Measurement Method Dip Dry No

Water Depth (mbTOC) 4.36 Well Depth (mbTOC)

Product Depth

(mbTOC)

Comments & Product

Description

After hydrasleeve deployment

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW6 Date/Time 09/02/2024 11:08AM

Measurement Method Dip Dry Well Depth (mbTOC) No

Water Depth (mbTOC) 4.357

Product Depth (mbTOC)

Comments & Product

Description

Before hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Well MW6

09/02/2024 11:09AM Date/Time

Measurement Method Dip Dry

Well Depth (mbTOC)

Product Depth

(mbTOC)

Comments & Product Description

Water Depth (mbTOC)

After hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Sampled Date/Time 09/02/2024 11:25AM

Well MW6

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW6

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description)colourlessSample Odour (Description)no odourSample Sheen (Description)no sheenSample Turbidity (Description)Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 11:25AM | 2204 | 7.41 | 56.6 | 5.3 | 22.5 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW4

Arrival Date/Time 07/02/2024 11:55AM Departure Date/Time 14/02/2024 02:45PM

Executed By Rowan Faint

Weather Sunny

Comments Well head flooded on arrival. Hydrasleeve installed. First hydrasleeve got lost down the well. Install

new one on 9/2/24

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:21 PM

Groundwater Data

Well MW4 Date/Time 07/02/2024 11:57AM

Well Depth (mbTOC)

6.98

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.65 Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW4 Date/Time 07/02/2024 12:06PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.635 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description

After hydrasleeve deployment

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW4 Date/Time 09/02/2024 02:34PM

Measurement Method Dip

Dry No

Well Depth (mbTOC)

Water Depth (mbTOC) 2.49

Product Depth

(mbTOC)
Comments & Product

After installing replacement

Description hydrasleeve

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well

Well Head PID (PPM)

Product Confirmed by Bailer

MW4

No

Date/Time 09/02/2024 11:43AM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.53 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description

Before hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well

Well Head PID (PPM)

Product Confirmed by Bailer

MW4

Date/Time

14/02/2024 02:17PM

Measurement Method Dip Dry No

No

Water Depth (mbTOC) 2.77 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description

Post hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Sampled Date/Time 14/02/2024 02:34PM

Well MW4
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW4

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description)colourlessSample Odour (Description)no odourSample Sheen (Description)no sheenSample Turbidity (Description)Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 02:35PM | 19817 | 6.66 | -11.2 | 3.8 | 29.6 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW1

Arrival Date/Time 07/02/2024 12:28PM Departure Date/Time 09/02/2024 10:52AM

Executed By Rowan Faint

Weather Sunny

Comments No well cap. Hydrasleeve installed

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:21 PM

Groundwater Data

Well MW1 Date/Time 07/02/2024 12:32PM

Well Depth (mbTOC)

6.605

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.25 Product Depth

(mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW1 Date/Time 07/02/2024 12:38PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.25 Well Depth (mbTOC)

Product Depth

(mbTOC)

Comments & Product

Description

After hydrasleeve deployment

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW1 Date/Time 09/02/2024 10:30AM

Measurement Method Dip Dry

Well Depth (mbTOC)

No

Water Depth (mbTOC) 2.235

Product Depth

Comments & Product Description

Before hydrasleeve sampling

Equipment ID

(mbTOC)

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Well MW1 Date/Time

Dry

09/02/2024 10:40AM

Measurement Method Dip Water Depth (mbTOC) 2.39

Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

After hydrasleeve sampling

Description **Equipment ID**

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Sampled Date/Time 09/02/2024 10:50AM

Well MW1

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW1

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description) no sheen

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 10:50AM | 25870 | 6.66 | 38.1 | 3.07 | 22.3 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW3

Arrival Date/Time 07/02/2024 01:03PM Departure Date/Time 09/02/2024 10:17AM

Executed By Rowan Faint

Weather Sunny

Comments Two hydrasleeves installed

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:22 PM

Groundwater Data

Well MW3 Date/Time 07/02/2024 01:03PM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 3.18
 Well Depth (mbTOC)
 8.05

Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW3 Date/Time 07/02/2024 01:13PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 3.15 Well Depth (mbTOC)

Product Depth

(mbTOC)

Comments & Product

Description

After hydrasleeve deployment

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW3 Date/Time 09/02/2024 09:24AM

Measurement Method Dip Dry No

Well Depth (mbTOC)

Water Depth (mbTOC) 3.175

Product Depth (mbTOC)

Comments & Product

Description

Before hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

09/02/2024 09:35AM Well MW3 Date/Time

Measurement Method Dip Dry

Water Depth (mbTOC) 3.78 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

After hydrasleeve sampling Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Page 48 of 81

Sampled Date/Time 09/02/2024 09:37AM

Well MW3

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW3

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1) QC103

QA Sample ID (2) QC203

QA Sample ID (3) QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 09:36AM | 34645 | 6.5 | 1.5 | 2.11 | 23.5 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW1

Arrival Date/Time 07/02/2024 01:30PM Departure Date/Time 07/02/2024 01:43PM

Executed By Rowan Faint

Weather Sunny

Comments

Well Information

Gatic Type

Key Type

Well Condition

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:23 PM

Sampled Date/Time 07/02/2024 01:43PM

Well

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) SW1

Purge Method

Sample Method Grab

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 01:42PM | 574 | 7.51 | 101.6 | 4.67 | 23.9 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: \$20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW2

Arrival Date/Time 07/02/2024 02:28PM Departure Date/Time 07/02/2024 02:36PM

Executed By Rowan Faint

Weather Sunny

Comments

Well Information

Gatic Type

Key Type

Well Condition

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:24 PM

Sampled Date/Time 07/02/2024 02:29PM

Well

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) SW2

Purge Method

Sample Method Grab

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 02:29PM | 656 | 7.7 | 81 | 4.19 | 25 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_07 Feb 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW2

Arrival Date/Time 07/02/2024 02:37PM Departure Date/Time 09/02/2024 01:52PM

Executed By Rowan Faint

Weather Sunny

Comments

Well Information

Gatic Type Old

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:24 PM

Groundwater Data

Well MW2 Date/Time 07/02/2024 02:37PM

Measurement MethodDipDryNoWater Depth (mbTOC)3.01Well Depth (mbTOC)7.575

Product Depth

(mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW2 Date/Time 07/02/2024 02:43PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 3 Well Depth (mbTOC)

Product Depth

(mbTOC)

Comments & Product

Description

After hydrasleeve deployment

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW2 Date/Time 09/02/2024 01:31PM

Measurement Method Dip Dry

Well Depth (mbTOC)

No

09/02/2024 01:37PM

Water Depth (mbTOC)

Product Depth

(mbTOC) Comments & Product

Description

Before hydrasleeve sampling

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well

Well Head PID (PPM)

Product Confirmed by Bailer

No

MW2

2.97

Measurement Method Dip

Dry

Well Depth (mbTOC)

Date/Time

Water Depth (mbTOC) 3.27

Product Depth (mbTOC)

Comments & Product

After hydrasleeve sampling Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer

No

Page 58 of 81

Sampled Date/Time 09/02/2024 01:51PM

Well MW2
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW2

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 01:51PM | 27224 | 6.59 | 92.7 | 3.05 | 23.2 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW1

Arrival Date/Time 09/07/2024 08:40AM Departure Date/Time 09/07/2024 09:17AM

Executed By Rowan Faint
Weather Light rain

Comments

Well Information

Gatic Type Key Type

Well Condition

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:11 PM

Page 61 of 81

Sampled Date/Time 09/07/2024 09:16AM

Well

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) SW1

Purge Method

Sample Method Grab

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light brown, light grey

Sample Odour (Description) no odour

Sample Sheen (Description) no sheen

Sample Turbidity (Description) Moderately turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 09:15AM | 334 | 7.08 | 38 | 6.3 | 13.6 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code SW2

Arrival Date/Time 09/07/2024 09:54AM Departure Date/Time 09/07/2024 10:01AM

Executed By Rowan Faint

Weather Overcast

Comments

Well Information

Gatic Type

Key Type

Well Condition

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:12 PM

Page 64 of 81

Sampled Date/Time 09/07/2024 09:55AM

Well

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) SW2

Purge Method

Sample Method Grab

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light grey
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 09:54AM | 370.4 | 8.48 | -18 | 6.09 | 13.7 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW2

Arrival Date/Time 09/07/2024 10:09AM Departure Date/Time 11/07/2024 02:47PM

Executed By Rowan Faint

Weather Overcast

Comments

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Good

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:13 PM

Groundwater Data

Well MW2 Date/Time 09/07/2024 10:10AM

Measurement MethodDipDryNoWater Depth (mbTOC)2.577Well Depth (mbTOC)7.58

Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW2 Date/Time 11/07/2024 02:29PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.555 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 11/07/2024 02:36PM

Well MW2
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW2

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | °C |
| 02:36PM | 23588 | 6.45 | -17.6 | 1.42 | 18.2 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW4

Arrival Date/Time 09/07/2024 10:47AM Departure Date/Time 11/07/2024 03:15PM

Executed By Rowan Faint

Weather Overcast

Comments Gatic flooded

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:14 PM

Groundwater Data

Well MW4 Date/Time 09/07/2024 10:47AM

Well Depth (mbTOC)

6.795

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.325 Product Depth

(mbTOC)
Comments & Product
Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW4 Date/Time 11/07/2024 02:59PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.31 Well Depth (mbTOC)

Product Depth

(mbTOC)
Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 11/07/2024 03:09PM

Well MW4
Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW4

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen
Sample Turbidity (Description) Non-turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 03:08PM | 14807 | 6.64 | -58.7 | 1.29 | 18.4 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW6

Arrival Date/Time 09/07/2024 11:01AM Departure Date/Time 11/07/2024 03:35PM

Executed By Rowan Faint

Weather Overcast

Comments

Well Information

Gatic Type Stick up

Key Type None

Well Condition Good

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:15 PM

Groundwater Data

Well MW6 Date/Time 09/07/2024 11:02AM

Measurement MethodDipDryNoWater Depth (mbTOC)3.565Well Depth (mbTOC)7.16

Product Depth (mbTOC)

Comments & Product

Description

Brown silt on IP

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW6 Date/Time 11/07/2024 03:23PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 3.557 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 11/07/2024 03:34PM

Well MW6
Matrix Type Water

Equipment ID

Sample Comments Insufficient sample remaining for water quality parameters

Field ID (Primary) MW6

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) colourless
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Suspended sediments

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW3

Arrival Date/Time 09/07/2024 11:17AM Departure Date/Time 11/07/2024 04:22PM

Executed By Rowan Faint

Weather Overcast

Comments

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:16 PM

Groundwater Data

Well MW3 Date/Time 09/07/2024 11:33AM

Measurement MethodDipDryNoWater Depth (mbTOC)2.88Well Depth (mbTOC)8.05

Water Depth (mbTOC) 2.88 Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW3 Date/Time 11/07/2024 04:16PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 2.875 Well Depth (mbTOC)

Product Depth (mbTOC)

Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 11/07/2024 04:17PM

Well MW3

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW3

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1) QC104

QA Sample ID (2) QC204

QA Sample ID (3) QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 04:17PM | 28201 | 5.86 | 15 | 2.2 | 18.3 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.



Monitoring Round: S20102_09 Jul 2024

Location Visit

Site ID S20102 Monitoring Zone

Location Code MW1

Arrival Date/Time 09/07/2024 11:47AM Departure Date/Time 11/07/2024 04:54PM

Executed By Rowan Faint
Weather Overcast

Comments

Well Information

Gatic Type New

Key Type 8mm hex key

Well Condition Average

Authorisation

Checked By Bec Chapple

Date Checked 09 Aug 2024 01:16 PM

Groundwater Data

Well MW1 Date/Time 09/07/2024 11:55AM

 Measurement Method
 Dip
 Dry
 No

 Water Depth (mbTOC)
 1.93
 Well Depth (mbTOC)
 6.61

Product Depth

(mbTOC)
Comments & Product

Description
Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Well MW1 Date/Time 11/07/2024 04:40PM

Measurement Method Dip Dry No

Water Depth (mbTOC) 1.925 Well Depth (mbTOC)

Product Depth

(mbTOC)
Comments & Product

Description

Equipment ID

Sediment Thickness (m)

Depth to Water with Pump (m)

gauging

Well Head PID (PPM)

Product Confirmed by Bailer No

Sample Data

Sampled Date/Time 11/07/2024 04:41PM

Well MW1

Matrix Type Water

Equipment ID

Sample Comments

Field ID (Primary) MW1

Purge Method

Sample Method Hydrasleeve

Waste Disposal

Purge Observations (purge start)

Purge Colour (Description)

Purge Odour (Description)

Purge Sheen (Description)

Purge Turbidity (Description)

Sample Observations (purge end)

Sample Colour (Description) light brown
Sample Odour (Description) no odour
Sample Sheen (Description) no sheen

Sample Turbidity (Description) Slightly turbid

QA Samples

QA Sample ID (1)

QA Sample ID (2)

QA Sample ID (3)

QA Sample ID (4)

Purge/Sampling Comments

Air Bubbles in Vials No

Headspace PID Reading(s)

Reaction with Preservatives No

Recharge-ability

Water Depth at end of Sampling (m)

Field Chemistry

| | EC (Field) | pH (Field) | Redox (Field) | Dissolved Oxygen (Field) | Temp (Field) |
|-----------------|------------|----------------|---------------|-----------------------------|--------------|
| Time | uS/cm | pH Units | mV | mg/L | ° C |
| 04:40PM | 22394 | 6.27 | -58 | 1.06 | 17.5 |
| Stabilisation * | ±3% (3) | ±0.05pH (3) | ±10mV (3) | ±10% (3) | ±10% (3) |

Green indicates readings have stabilised according to the criteria shown, red indicates they haven't. The number in brackets indicates the number of readings that need to meet the criteria for the readings to be considered stable.

Appendix C: Calibration Certificates



Interface Meter Heron H.Oil

| Company Name | WAM Scientific |
|----------------|--|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | Heron H.Oil Interface Meter (30m) |
| Serial Number | 01-7967 |
| Client Name | Bec Chapple (Senversa) |
| Project Number | S20102 |

| | Instrument Check | | | | | |
|-----------------------|------------------------------|-------------|--|--|--|--|
| Item | Test | Test Passed | Comments | | | |
| 9V Battery | Klein Tools MM300 Multimeter | ✓ | Battery voltage reading above 7.9V | | | |
| Battery Box | Check | ✓ | No damage | | | |
| Face and Back Plates | Check | ✓ | No damage | | | |
| Thumb Screws | Check | ✓ | Rubber ends intact | | | |
| Tape Hangar/Protector | Check | ✓ | No damage | | | |
| On/Off Button | Operation | ✓ | Button is functional | | | |
| Buzzer | Operation | ✓ | Intermittent tone in H ₂ O, solid tone in product | | | |
| LED Signal Light | Operation | ✓ | LED light functional – green and red | | | |
| Probe | Operation/Check | ✓ | Decontaminated, cleaned and tested | | | |
| Tape | Condition/Check | ✓ | Decontaminated and cleaned, no damage | | | |
| Connection | Check | ✓ | Probe and link connected correctly and tightly | | | |
| РСВ | Operation | ✓ | Unit is fully functional | | | |
| Electronics Panel | Orientation | ✓ | Correctly aligned | | | |

| Instrument Readings | | | | | |
|---------------------|--------------|----------------|--|--|--|
| Product | Buzzer | LED Light | | | |
| H₂O | Intermittent | Blinking – Red | | | |
| Petroleum | Solid | Steady – Red | | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The interface meter was decontaminated, cleaned and tested with a mixture of tap water and petrol, shielded from ambient light.

| Checked By | William Pak |
|-------------------------|-------------|
| Calibration Date | 01/02/2023 |
| Calibration Due | 01/08/2023 |





Water Quality Meter YSI Professional Plus

| Company Name | WAM Scientific |
|----------------|--|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | YSI Professional Plus Water Quality Meter w/ 1m Quatro Cable |
| Serial Number | 21A102654 |
| Client Name | Bec Chapple (Senversa) |
| Project Number | S20102 |
| Comments | - |

| Instrument Check | | | | | |
|--|-----------------------|-------------|---|--|--|
| Item | Test | Test Passed | Comments | | |
| 2 x Alkaline C-size Batteries Klein Tools MM300 Multimeter | | ✓ | Both batteries reading above 2.9V | | |
| Battery Saver Function | Operation | ✓ | Automatically turns off after 60 minutes if idle | | |
| Unit Display | Operation | ✓ | Screen visible, no damage | | |
| Keypad | Operation | ✓ | Responsive, no damage | | |
| Connection Port and Cable | Condition/Check | ✓ | Clean, no damage | | |
| Monitor Housing | Condition/Check | ✓ | No damage | | |
| Firmware | Version | ✓ | 4.0.0 | | |
| pH Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| pH millivolts for pH 7.00 | Calibration | ✓ | pH 7.00 calibration range between 0 mV ± 50 mV | | |
| pH millivolts for pH 4.00 Calibration | | ✓ | pH 4 mV range +165 to +180 from 7 buffer mV value | | |
| pH slope Calibration | | ✓ | Range between 55 to 60 mV/pH (ideal value 59 mV) | | |
| Response time < 90 seconds Calibration | | ✓ | Responds to correct value within 90 seconds | | |
| ORP Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| ORP Reading | Calibration | ✓ | Within ± 80 mV of reference Zobell Reading | | |
| Response time < 90 seconds | Calibration | ✓ | Responds to correct value within 90 seconds | | |
| Conductivity/Temp Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| Conductivity Cell | Calibration | ✓ | Conductivity cell constant 5.0 ± 1.0 in GLP file | | |
| Clean Sensor Readings | Calibration | ✓ | Clean sensor reads less than 3 uS/cm in dry air | | |
| Dissolved Oxygen Probe Condition/Calibration | | ✓ | Calibrated and conforms to manufacturer's specs | | |
| DO Cap | Condition/Calibration | ✓ | 1.25 mil PE membrane (yellow membrane) | | |
| DO Sensor in Use | Condition | ✓ | Polarographic DO sensor | | |
| DO Sensor Value Calibration | | ✓ | (min 4.31 uA - max 8.00 uA) Avg 6.15 uA | | |

Instrument Readings

| mistrament headings | | | | | | |
|-------------------------------|---|---------------|-------------------|----------|--------|-------|
| Parameter | Standard Used | Reference No. | Calibration Value | Observed | Actual | Units |
| Temperature | Centre 370 Thermometer | Room Temp. | 26.6 | 26.5 | 26.6 | °C |
| pН | pH 4.00 | 386466 | 4.01 | 4.04 | 4.01 | рН |
| pН | pH 7.00 | 387329 | 7.00 | 6.96 | 7.00 | рН |
| Conductivity | 2760 μs/cm at 25°C | 388521 | 2760 | 2797 | 2760 | μs/cm |
| ORP (Ref. check only) | Zobell A & B | 380835/382785 | 229.9 | 223.8 | 229.9 | mV |
| Zero Dissolved O ₂ | NaSO ₃ in Distilled H ₂ O | 389912 | 0.0 | 0.3 | 0.0 | % |
| 100% Dissolved O ₂ | 100% Air Saturated H₂O | Fresh Air | 100.0 | 108.1 | 100.0 | % |

7Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

| Calibrated By | William Pak |
|-------------------------|-------------|
| Calibration Date | 01/02/2023 |
| Calibration Due | 01/08/2023 |





Interface Meter Heron H.Oil

| | - |
|----------------|--|
| Company Name | WAM Scientific |
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | Heron H.Oil Interface Meter (60m) |
| Serial Number | 01-8640 |
| Client Name | Bec Chapple (Senversa) |
| Project Number | S20102 |

| | Instrument Check | | | | |
|-----------------------|------------------------------|--|--|--|--|
| Item | Test | Test Passed | Comments | | |
| 9V Battery | Klein Tools MM300 Multimeter | ools MM300 Multimeter ✓ Battery voltage reading above 7.9V | | | |
| Battery Box | Check | ✓ | No damage | | |
| Face and Back Plates | Check | ✓ | No damage | | |
| Thumb Screws | Check | ✓ | Rubber ends intact | | |
| Tape Hangar/Protector | Check | ✓ | No damage | | |
| On/Off Button | Operation | ✓ | Button is functional | | |
| Buzzer | Operation | ✓ | Intermittent tone in H ₂ O, solid tone in product | | |
| LED Signal Light | Operation | ✓ | LED light functional – green and red | | |
| Probe | Operation/Check | ✓ | Decontaminated, cleaned and tested | | |
| Tape | Condition/Check | ✓ | Decontaminated and cleaned, no damage | | |
| Connection | Check | ✓ | Probe and link connected correctly and tightly | | |
| PCB | Operation | ✓ | Unit is fully functional | | |
| Electronics Panel | Orientation | ✓ | Correctly aligned | | |

| Instrument Readings | | | | |
|--------------------------|--------------|----------------|--|--|
| Product Buzzer LED Light | | | | |
| H₂O | Intermittent | Blinking – Red | | |
| Petroleum | Solid | Steady – Red | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The interface meter was decontaminated, cleaned and tested with a mixture of tap water and petrol, shielded from ambient light.

| Checked By | William Pak |
|------------------|-------------|
| Calibration Date | 30/07/2023 |
| Calibration Due | 30/01/2024 |



WAM Scientific: Sydney Office - Clemton Park 16 Lawn Avenue CLEMTON PARK NSW 2206 T: +61 405 241 484 E: rentals@wamscientific.com.au Website: www.wamscientific.com.au Alternate Email Addresses: admin@wamscientific.com.au accounts@wamscientific.com.au sales@wamscientific.com.au service@wamscientific.com.au



Water Quality Meter YSI Professional Plus

| Company Name | WAM Scientific |
|----------------|---|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | YSI Pro Plus Water Quality Meter w/ 1m Quatro Cable |
| Serial Number | 20B122031 |
| Client Name | Hayley Yellowlees/Chris Redford (Senversa) |
| Project Number | S20049 |
| Comments | - |

| Instrument Check | | | | | |
|-------------------------------|------------------------------|-------------|---|--|--|
| Item | Test | Test Passed | Comments | | |
| 2 x Alkaline C-size Batteries | Klein Tools MM300 Multimeter | ✓ | Both batteries reading above 2.9V | | |
| Battery Saver Function | Operation | ✓ | Automatically turns off after 60 minutes if idle | | |
| Unit Display | Operation | ✓ | Screen visible, no damage | | |
| Keypad | Operation | ✓ | Responsive, no damage | | |
| Connection Port and Cable | Condition/Check | ✓ | Clean, no damage | | |
| Monitor Housing | Condition/Check | ✓ | No damage | | |
| Firmware | Version | ✓ | 4.0.0 | | |
| pH Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| pH millivolts for pH 7.00 | Calibration | ✓ | pH 7.00 calibration range between 0 mV ± 50 mV | | |
| pH millivolts for pH 4.00 | Calibration | ✓ | pH 4 mV range +165 to +180 from 7 buffer mV value | | |
| pH slope | Calibration | ✓ | Range between 55 to 60 mV/pH (ideal value 59 mV) | | |
| Response time < 90 seconds | Calibration | ✓ | Responds to correct value within 90 seconds | | |
| ORP Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| ORP Reading | Calibration | ✓ | Within ± 80 mV of reference Zobell Reading | | |
| Response time < 90 seconds | Calibration | ✓ | Responds to correct value within 90 seconds | | |
| Conductivity/Temp Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| Conductivity Cell | Calibration | ✓ | Conductivity cell constant 5.0 ± 1.0 in GLP file | | |
| Clean Sensor Readings | Calibration | ✓ | Clean sensor reads less than 3 uS/cm in dry air | | |
| Dissolved Oxygen Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| DO Cap | Condition/Calibration | ✓ | 1.25 mil PE membrane (yellow membrane) | | |
| DO Sensor in Use | Condition | ✓ | Polarographic DO sensor | | |
| DO Sensor Value | Calibration | ✓ | (min 4.31 uA - max 8.00 uA) Avg 6.15 uA | | |

Instrument Readings

| | | | 0 | | | |
|-------------------------------|---|---------------|-------------------|----------|--------|-------|
| Parameter | Standard Used | Reference No. | Calibration Value | Observed | Actual | Units |
| Temperature | Centre 370 Thermometer | Room Temp. | 14.2 | 14.6 | 14.2 | °C |
| pН | pH 4.00 | 386466 | 4.01 | 4.05 | 4.01 | рН |
| pН | pH 7.00 | 387329 | 7.00 | 7.07 | 7.00 | рН |
| Conductivity | 2760 μs/cm at 25°C | 388521 | 2760 | 2629 | 2760 | μs/cm |
| ORP (Ref. check only) | Zobell A & B | 380835/382785 | 253.2 | 259.6 | 253.2 | mV |
| Zero Dissolved O ₂ | NaSO ₃ in Distilled H ₂ O | 389912 | 0.0 | 0.1 | 0.0 | % |
| 100% Dissolved O ₂ | 100% Air Saturated H₂O | Fresh Air | 100.0 | 100.4 | 100.0 | % |
| | | | | | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

| Calibrated By | William Pak |
|-------------------------|-------------|
| Calibration Date | 14/08/2023 |
| Calibration Due | 14/02/2024 |





Water Quality Meter YSI Professional Plus

| Company Name | WAM Scientific |
|----------------|---|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | YSI Pro Quatro Water Quality Meter w/ 1m Quatro Cable |
| Serial Number | 21A102654 |
| Client Name | Rowan Faint (Senversa) |
| Project Number | S20102 |
| Comments | - |

| Instrument Check | | | | | |
|-------------------------------|--|--|---|--|--|
| Item Test | | Test Passed | Comments | | |
| 2 x Alkaline C-size Batteries | Alkaline C-size Batteries Klein Tools MM300 Multimeter | | Both batteries reading above 2.9V | | |
| Battery Saver Function | Operation | ✓ | Automatically turns off after 60 minutes if idle | | |
| Unit Display | Operation | ✓ | Screen visible, no damage | | |
| Keypad | Operation | ✓ | Responsive, no damage | | |
| Connection Port and Cable | Condition/Check | ✓ | Clean, no damage | | |
| Monitor Housing | Condition/Check | ✓ | No damage | | |
| Firmware | Version | ✓ | 4.0.0 | | |
| pH Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| pH millivolts for pH 7.00 | Calibration | ✓ | pH 7.00 calibration range between 0 mV ± 50 mV | | |
| pH millivolts for pH 4.00 | Calibration | ✓ | pH 4 mV range +165 to +180 from 7 buffer mV value | | |
| pH slope | Calibration | ✓ Range between 55 to 60 mV/pH (ideal value 59 | | | |
| Response time < 90 seconds | Calibration | ✓ | ✓ Responds to correct value within 90 seconds | | |
| ORP Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| ORP Reading | Calibration | ✓ | Within ± 80 mV of reference Zobell Reading | | |
| Response time < 90 seconds | Calibration | ✓ | Responds to correct value within 90 seconds | | |
| Conductivity/Temp Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| Conductivity Cell | Calibration | ✓ | Conductivity cell constant 5.0 ± 1.0 in GLP file | | |
| Clean Sensor Readings | Calibration | ✓ | Clean sensor reads less than 3 uS/cm in dry air | | |
| Dissolved Oxygen Probe | Condition/Calibration | ✓ | Calibrated and conforms to manufacturer's specs | | |
| DO Cap | Condition/Calibration | ✓ | 1.25 mil PE membrane (yellow membrane) | | |
| DO Sensor in Use | Condition | ✓ | Polarographic DO sensor | | |
| DO Sensor Value | Calibration | ✓ | (min 4.31 uA - max 8.00 uA) Avg 6.15 uA | | |

Instrument Readings

| Parameter | Standard Used | Reference No. | Calibration Value | Observed | Actual | Units |
|-------------------------------|---|---------------|-------------------|----------|--------|-------|
| Temperature | Centre 370 Thermometer | Room Temp. | 27.7 | 28.1 | 27.7 | °C |
| рН | pH 4.00 | 386466 | 4.01 | 4.05 | 4.01 | рН |
| рН | pH 7.00 | 387329 | 7.00 | 7.02 | 7.00 | рН |
| Conductivity | 2760 μs/cm at 25°C | 388521 | 2760 | 2589 | 2760 | μs/cm |
| ORP (Ref. check only) | Zobell A & B | 380835/382785 | 225.3 | 230.1 | 225.3 | mV |
| Zero Dissolved O ₂ | NaSO ₃ in Distilled H ₂ O | 389912 | 0.0 | -0.1 | 0.0 | % |
| 100% Dissolved O ₂ | 100% Air Saturated H ₂ O | Fresh Air | 100.0 | 93.7 | 100.0 | % |
| | | | | | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

| Calibrated By | William Pak |
|-------------------------|-------------|
| Calibration Date | 31/01/2024 |
| Calibration Due | 31/07/2024 |





Interface Meter Heron H.Oil

| Company Name | WAM Scientific |
|----------------|--|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | Heron H.Oil Interface Meter (30m) |
| Serial Number | 01-8895 |
| Client Name | Rowan Faint (Senversa) |
| Project Number | S20102 |

| | Instrun | nent Check | |
|-----------------------|------------------------------|-------------|--|
| Item | Test | Test Passed | Comments |
| 9V Battery | Klein Tools MM300 Multimeter | ✓ | Battery voltage reading above 7.9V |
| Battery Box | Check | ✓ | No damage |
| Face and Back Plates | Check | ✓ | No damage |
| Thumb Screws | Check | ✓ | Rubber ends intact |
| Tape Hangar/Protector | Check | ✓ | No damage |
| On/Off Button | Operation | ✓ | Button is functional |
| Buzzer | Operation | ✓ | Intermittent tone in H ₂ O, solid tone in product |
| LED Signal Light | Operation | ✓ | LED light functional – green and red |
| Probe | Operation/Check | ✓ | Decontaminated, cleaned and tested |
| Tape | Condition/Check | ✓ | Decontaminated and cleaned, no damage |
| Connection | Check | ✓ | Probe and link connected correctly and tightly |
| PCB | Operation | ✓ | Unit is fully functional |
| Electronics Panel | Orientation | ✓ | Correctly aligned |

| Instrument Readings | | | | | |
|--------------------------|--------------|----------------|--|--|--|
| Product Buzzer LED Light | | | | | |
| H₂O | Intermittent | Blinking – Red | | | |
| Petroleum | Solid | Steady – Red | | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The interface meter was decontaminated, cleaned and tested with a mixture of tap water and petrol, shielded from ambient light.

| Checked By | William Pak |
|------------------|-------------|
| Calibration Date | 31/01/2024 |
| Calibration Due | 31/07/2024 |



WAM Scientific: Sydney Office - Clemton Park 16 Lawn Avenue CLEMTON PARK NSW 2206 T: +61 405 241 484 E: rentals@wamscientific.com.au Website: www.wamscientific.com.au Alternate Email Addresses: admin@wamscientific.com.au accounts@wamscientific.com.au sales@wamscientific.com.au service@wamscientific.com.au



Water Quality Meter YSI Professional Plus

| | · | WAM Scientific | | | | | | |
|-------------------------------|----------|-------------------------------|-------------------|---------------|---|---|---------------------------------------|-------|
| Company Name | | | | | | | | |
| Office Address | | _ | hipping Norton N | SW 2170 | | | | |
| Phone Number | | 5 241 484 | | | | | | |
| Contact Name | William | n Pak | | | | | | |
| Instrument | YSI Pro | Quatro Water Qu | uality Meter w/ 1 | m Quatro Cabl | e | | | |
| Serial Number | 21A102 | 2654 | | | | | | |
| Client Name | Rowan | Faint (Senversa) | | | | | | |
| Project Number | S20102 | S20102 | | | | | | |
| Comments | - | - | | | | | | |
| | | | Instrum | ent Check | | | | |
| Item | | Te | st | Test Passed | | Co | mments | |
| 2 x Alkaline C-size Ba | itteries | Klein Tools MM3 | 300 Multimeter | ✓ | Both | batteries reading a | bove 2.9V | |
| Battery Saver Fund | ction | Opera | ation | ✓ | | | after 60 minutes if i | dle |
| Unit Display | | Opera | | ✓ | Scree | en visible, no dama | ge | |
| Keypad | | Opera | ation | ✓ | Resp | onsive, no damage | | |
| Connection Port and | l Cable | Condition | n/Check | ✓ | Clear | n, no damage | | |
| Monitor Housin | ng | Condition | n/Check | ✓ | No d | amage | | |
| Firmware | | Vers | | ✓ | 4.0.0 | 1 | | |
| pH Probe | | Condition/0 | Calibration | ✓ | | librated and conforms to manufacturer's specs | | |
| pH millivolts for pH | | Calibr | ation | ✓ | | | ge between 0 mV ± 5 | |
| pH millivolts for pH | 4.00 | Calibr | | ✓ | | | +180 from 7 buffer | |
| pH slope | | Calibr | | ✓ | | |) mV/pH (ideal value | |
| Response time < 90 s | econds | Calibr | | √ | | onds to correct value within 90 seconds | | |
| ORP Probe | | Condition/0 | | ✓ | | brated and conforms to manufacturer's spe | | |
| ORP Reading | | Calibr | | √ | 1 | Within ± 80 mV of reference Zobell Reading | | |
| Response time < 90 se | | Calibr | | √ | Responds to correct value within 90 seconds | | | |
| Conductivity/Temp | | Condition/0 | | <u>√</u> | | | s to manufacturer's | • |
| Conductivity Ce | | Calibr | | <u>√</u> | 1 | • | nt 5.0 ± 1.0 in GLP fi | |
| Clean Sensor Read | | Calibr | | <u>√</u> | | | than 3 uS/cm in dry | |
| Dissolved Oxygen F DO Cap | rope | Condition/C | | <u>√</u> | | | s to manufacturer's (yellow membrane) | specs |
| DO Cap DO Sensor in Us | 20 | Condition/C | | ✓ | | rographic DO senso | | |
| DO Sensor Valu | | Calibra | | <u> </u> | | 4.31 uA - max 8.00 | | |
| DO SEIISOI VAIU | | Calibra | | nt Readings | [(111111 | JI UA IIIAX 0.00 | ary Avg 0.13 uA | |
| Parameter | Sta | ndard Used | Reference No. | Calibration \ | /alue | Pre-Cal Value | Post-Cal Value | Units |
| Temperature | | re 370 Therm. | Room Temp. | 15.0 | raiue | 14.7 | 15.0 | °C |
| pH | Cent | pH 4.00 | 417183 | 4.01 | | 4.05 | 4.01 | pH |
| pH | | pH 7.00 | 419528 | 7.00 | | 6.93 | 7.00 | рН |
| Conductivity | 2760 | μs/cm at 25°C | 399819 | 2760 | | 2750 | 2760 | μs/cm |
| ORP | | obell A & B | 420448/418958 | 251.0 | | 260.3 | 251.0 | mV |
| Zero Dissolved O ₂ | | in Distilled H ₂ O | 426184 | 0.0 | | 0.5 | 0.0 | % |
| 100% Dissolved O ₂ | | ir Saturated H ₂ O | Fresh Air | 100.0 | | 93.9 | 100.0 | % |
| _ | | | | ration | | | | |
| | | | | | | | | |

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied

William Pak

27/06/2024

27/12/2024

was obtained in accordance with manufacturer's specifications using solutions of known values.



Calibrated By

Calibration Date

Calibration Due



Interface Meter Heron H.Oil

| Company Name | WAM Scientific |
|----------------|--|
| Office Address | 26 Bungarra Crescent, Chipping Norton NSW 2170 |
| Phone Number | +61 405 241 484 |
| Contact Name | William Pak |
| Instrument | Heron H.Oil Interface Meter (30m) |
| Serial Number | 01-09486 |
| Client Name | Rowan Faint (Senversa) |
| Project Number | S20102 |

| | Instrun | nent Check | |
|-----------------------|------------------------------|-------------|--|
| Item | Test | Test Passed | Comments |
| 9V Battery | Klein Tools MM300 Multimeter | ✓ | Battery voltage reading above 7.9V |
| Battery Box | Check | ✓ | No damage |
| Face and Back Plates | Check | ✓ | No damage |
| Thumb Screws | Check | ✓ | Rubber ends intact |
| Tape Hangar/Protector | Check | ✓ | No damage |
| On/Off Button | Operation | ✓ | Button is functional |
| Buzzer | Operation | ✓ | Intermittent tone in H ₂ O, solid tone in product |
| LED Signal Light | Operation | ✓ | LED light functional – green and red |
| Probe | Operation/Check | ✓ | Decontaminated, cleaned and tested |
| Tape | Condition/Check | ✓ | Decontaminated and cleaned, no damage |
| Connection | Check | ✓ | Probe and link connected correctly and tightly |
| PCB | Operation | ✓ | Unit is fully functional |
| Electronics Panel | Orientation | ✓ | Correctly aligned |

| Instrument Readings | | | | | |
|--------------------------|--------------|----------------|--|--|--|
| Product Buzzer LED Light | | | | | |
| H₂O | Intermittent | Blinking – Red | | | |
| Petroleum | Solid | Steady – Red | | | |

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The interface meter was decontaminated, cleaned and tested with a mixture of tap water and petrol, shielded from ambient light.

| Checked By | William Pak |
|------------------|-------------|
| Calibration Date | 27/06/2024 |
| Calibration Due | 27/12/2024 |



Appendix D: Quality Assessment / Quality Control



| Job Number: | S20102 |
|----------------------------------|--|
| Report Title: | Surface Water and Groundwater Monitoring |
| Client: | ReDirect Recycling |
| | |
| | |
| Completed By: | Bec Chapple |
| | Bec Chapple 9-Aug-24 |
| Completed By: Date: Verified By: | |

| SAMPLE DELIVERY GROUP (SDG): | ES2304342 | SAMPLE DELIVERY GROUP (SDG): | ES2304011 |
|------------------------------------|-----------|------------------------------------|-----------|
| Laboratory: | ALS | Laboratory: | ALS |
| Sample Dates: | 10-Feb-23 | Sample Dates: | 8-Feb-23 |
| Sample Media: | Water | Sample Media: | Water |

| Quality Assurance Process Standard Procedures | | | | | | | |
|---|---|--|---|-----------------------------|--|----------------------------|---|
| | Objectives & Measure | Acceptance Criteria | Source of Information | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| | Standard field sampling procedures and forms used | No deviation from standard procedure and | Borelogs, field sheets, COCs, data | Criteria Met? Yes | | Criteria Met? Yes | |
| | otalidad liod saliipiing procedures and roms acco | forms used. | tables | 100 | | 100 | |
| Equipment Calibration | All equipment calibrated in accordance with manufacturers | All equipment calibrated in accordance with | Calibration Certificates / Records | Yes | | Yes | |
| Testing Method | specifications NATA accredited methods used for all analyses determined | manufacturers specifications. Primary and secondary laboratories to use | Laboratory Report | Yes | | Yes | |
| Accreditation | | NATA accredited methods for all analytes | | | | | |
| Quality Control Sampling | Field QC sampling frequency in accordance with AS4482.1- | determined. Field (Intra-laboratory) Duplicates - ≥ 1 in 20 | QA/QC register (within field book) | N/A | Relevant intra-laboratory QC samples for this WME reported | Yes | QC101 |
| | 2005 | primary samples. | , | | in batch ES2304011. | | |
| | | (note that PFAS NEMP recommends 1 in 10 for PFAS investigations) | | | | | |
| | | Secondary (inter-laboratory) duplicates - ≥ 1 in | QA/QC register (within field book) | N/A | Relevant inter-laboratory QC samples for this WME reported | N/A | Relevant inter-laboratory QC samples for this WME reported |
| | | 20 primary samples. | | | in batch 316159. | | in batch 316159. |
| | | (note that PFAS NEMP recommends 1 in 10 for PFAS investigations) | | | | | |
| | | Rinsate Blanks - ≥ 1 per day, per matrix per | QA/QC register (within field book) | N/A | Relevant intra-laboratory QC samples for this WME reported | Yes | QC301 |
| | | equipment. | | | in batch ES2304011. | | |
| | | Trip Blanks - ≥ 1 per esky containing samples | QA/QC register (within field book) | N/A | Relevant intra-laboratory QC samples for this WME reported | Yes | QC401 |
| | Laboratory CO and win for suppose in accordance with NEDO | for volatiles. | Laborator Donasto | No | in batch ES2304011. | No | A laboratory dualizate for DALI/Dhonele and TDLI |
| | Laboratory QC analysis frequency in accordance with NEPC 2013 | Laboratory Duplicates - at least 1 in 10 analyses or 1 per process batch. | Laboratory Reports | No | A laboratory duplicate for PAH/Phenols and TRH- Semivolatile fraction was not analysed, resulting in a non- | No | A laboratory duplicate for PAH/Phenols and TRH- Semivolatile fraction was not analysed, resulting in a non- |
| | | | | | conformance for frequency for this analysis. Not considered to | | conformance for frequency for this analysis. Not considered t |
| | | | | | impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory | | impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory |
| | | | | | field duplicates were analysed for PAH/ phenols and TRH | | field duplicates were analysed for PAH/ phenols and TRH |
| | | | | | semivolatile fraction and were DQI compliant. | | semivolatile fraction and were DQI compliant. |
| | | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | | | | | | |
| | | Surrogate Recoveries - all samples spiked | Laboratory Reports | Yes | | Yes | |
| | | where appropriate (e.g. chromatographic analysis of organics). | | | | | |
| | | Laboratory Control Samples - at least 1 per | Laboratory Reports | Yes | | Yes | |
| | | process batch. Matrix Spikes - at least 1 per matrix type per | Laboratory Reports | No | A matrix spike for PAH/Phenols and TRH-Semivolatile | No | A matrix spike for PAH/Phenols, dissolved metals and TRH- |
| | | process batch. | , | | fraction was not analysed, resulting in a non-conformance for | | Semivolatile fraction was not analysed, resulting in a non- |
| | | | | | frequency for this analysis. Not considered to impact upon | | conformance for frequency for this analysis. Not considered to |
| | | | | | assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory field duplicates were | | impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory |
| | | | | | analysed for PAH/ phenols and TRH semivolatile fraction and | | field duplicates were analysed for PAH/ phenol, dissolved |
| | | | | | were DQI compliant. | | metals and TRH semivolatile fraction and were DQI |
| Sample Preservation, | Samples appropriately preserved upon collection, stored and | In accordance with laboratory specific method | Laboratory Reports | Yes | | Yes | compliant. |
| Handling and Holding Times | transported, and analysed within holding times | requirements. | | | | | |
| | | Unless specific method indicates otherwise, soil and water samples should be stored, | | | | | |
| | | transported and received by the laboratory at < | | | | | |
| | | 6°C. | | | | | |
| Data Management | No errors in data transcription | Entry of field data verified by peer. | 10% check of electronically imported data (e.g. ESDAT). | Yes | | Yes | |
| | | | 100% check of manually entered data | | | | |
| | | | (e.g. field parameters, gauging data). | | | | |
| Data Useability | Limits of reporting less than investigation levels | Limits of reporting less than relevant | Results Tables | Yes | | Yes | |
| , | | investigation levels. | | | | | |
| | | | | | | | |
| Quality Control Process | Objectives & Measure | Acceptance Criteria | How? (i.e. ESDAT output, review | | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| Field (Intra-laboratory) | Field Duplicate samples used assess the variability in analyte | Analysed for same chemicals as primary | lab reports, review data etc) ESDAT generated summary of relative | Criteria Met? | Relevant intra-laboratory QC samples for this WME reported | Criteria Met? Yes | |
| Duplicate Sampling and | concentration between samples collected from the sample | sample. | percent difference (RPD) results for | N/A | in batch ES2304011. | 165 | |
| Analysis | location and the reproducibility of the laboratory analysis. | RPD <30% of mean conc. where both conc. | field duplicate samples. | | | | |
| | Where required, resubmission of previously analysed samples for chemicals within their holding times may be | >20 x LOR RPD <50% of mean conc. where both conc. | 1 | | | | |
| | - destrict to the feather and the second of | 10-20 x LOR | | | | | |
| , | undertaken to further assess precision level of precision. | | | | | | |
| | undertaken to further assess precision level of precision. | RPD No limit where both conc. < 10 x LOR | | | | | |
| Secondary Inter-laborator) | Results are accurate and free from laboratory error. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary | ESDAT generated summary of relative | N/A | | N/A | Relevant inter-laboratory QC samples for this WME reported |
| Duplicate Sampling and | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. | percent difference (RPD) results for | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 316159. | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 316159. |
| Duplicate Sampling and Analysis | Results are accurate and free from laboratory error. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. | | N/A | | N/A | |
| Duplicate Sampling and Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. | percent difference (RPD) results for | N/A | | N/A | |
| Duplicate Sampling and Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. | percent difference (RPD) results for | N/A | | N/A | |
| Duplicate Sampling and Analysis Field Rinsate Blank | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported | | |
| Duplicate Sampling and Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. | | in batch 316159. | | |
| Duplicate Sampling and Analysis Field Rinsate Blank | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 316159. Relevant intra-laboratory QC samples for this WME reported | | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. | | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance, in general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <60% of mean conc. where both conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance, In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory outrol samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance, in general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD < 30% of mean conc. where both conc. > 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes | in batch 316159. Mattix spike recovery not determined for manganese and |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes Yes | In batch 316159. Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance, in general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD < 30% of mean conc. where both conc. > 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes Yes | in batch 316159. Mattix spike recovery not determined for manganese and |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD < 30% of mean conc. where both conc. > 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD < 30% of mean conc. where both conc. > 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Ves | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Ves | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Ves | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory outrol samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed searchy like field samples. These blanks are used by the laboratory to assess contamination introduced during | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Ves | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory outrol samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed searchy like field samples. These blanks are used by the laboratory to assess contamination introduced during | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Ves | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes | Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory ontrol samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to example preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes NO Yes | In batch 316159. Mattix spike recovery not determined for manganese and nitrite as N as the background level greater than or equal to 4x |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation D Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory ontrol samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to example preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD < 30% of mean conc. where both conc. > 20 x LOR. RPD < 50% of mean conc. where both conc. 10 - 20 x LOR. RPD < LOR. RPD < LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes NO Yes | In batch 316159. Mattix spike recovery not determined for manganese and nitrie as N as the background level greater than or equal to 4: |
| Duplicate Sampling and Analysis Field Rinsate Blank Preparation 9 Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory duplicates are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. <10 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes Yes | in batch 316159. Relevant intra-laboratory QC samples for this WME reported in batch ES2304011. Relevant intra-laboratory QC samples for this WME reported | Yes Yes Yes N/A Yes NO Yes | In batch 316159. Mattix spike recovery not determined for manganese and nitrie as N as the background level greater than or equal to 4 |



| Job Number: | S20102 |
|---------------|--|
| Report Title: | Surface Water and Groundwater Monitoring |
| Client: | ReDirect Recycling |
| | |
| Completed By: | Bec Chapple |
| Completed By: | Bec Chapple 9-Aug-24 |
| | |

| SAMPLE DELIVERY GROUP (SDG): | 316159 | SAMPLE DELIVERY GROUP (SDG): | ES2326328 |
|------------------------------------|-----------|------------------------------------|-----------|
| Laboratory: | Envirolab | Laboratory: | ALS |
| Sample Dates: | 8-Feb-23 | Sample Dates: | 14-Aug-23 |
| Sample Media: | Water | Sample Media: | Water |

| March 1999/06 March 1999/0 | | | | | | | | |
|--|---|--|--|---|-------------------------|--|----------------------------------|--|
| Management Man | | Objectives & Measure | Acceptance Criteria | Source of Information | | Notes/Details of Nonconformance | | Notes/Details of Nonconformance |
| March Marc | | Standard field sampling procedures and forms used | No deviation from standard procedure and | Borelogs, field sheets COCs data | | | | |
| March Marc | | | forms used. | tables | | | | |
| Comment | Equipment Calibration | | | Calibration Certificates / Records | Yes | | Yes | |
| Marie | Testing Method | | | Laboratory Report | Yes | | Yes | |
| Command Continues Comm | Accreditation | | - | | | | | |
| March Marc | Quality Control Sampling | Field QC sampling frequency in accordance with AS4482.1- | | QA/QC register (within field book) | N/A | Primary laboratory received sample | Yes | QC102 |
| Application | Frequency | 2005 | | | | | | |
| Comment of the Comm | | | | | | | | |
| Part | | | | QA/QC register (within field book) | Yes | QC201 | N/A | Relevant inter-laboratory QC samples for this WME reported |
| April | | | | | | | | in batch 1020195. |
| Part Company Part | | | for PFAS investigations) | | | | | |
| April 1 1 1 1 1 1 1 1 1 | | | | QA/QC register (within field book) | N/A | Primary laboratory received sample | Yes | QC302 |
| The company of an international content of the company | | | oquip. 10.11 | | | | | |
| Commonweight Comm | | | | QA/QC register (within field book) | N/A | Primary laboratory received sample | Yes | QC502 |
| March Marc | | Laboratory QC analysis frequency in accordance with NEPC | | Laboratory Reports | Yes | | No | A laboratory duplicate for PAH/Phenols and TRH- |
| Part Company | | 2013 | | | | | | Semivolatile fraction was not analysed, resulting in a non- conformance for frequency for this analysis. Not considered to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenois and TRH semivolatile fraction and were DQI compliant. |
| Annual Content | | | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| March 1996 |] | | Surrogate Recoveries - all samples spiked | Laboratory Reports | Yes | | Yes | |
| ACCEPTAGE AND AC | | | | | | | | |
| Part | | | | Laboratory Reports | Yes | | Yes | |
| Seath research control of the contro | | | process batch. | | | | | |
| Section of the control of the contro | | | process batch. | | | | No | A matrix spike for PAH/Phenols and TRH-Semivolatile fraction was not analysed, resulting in a non-conformance for frequency for this analysis. Not considered to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH semivolatile fraction and were DQI compliant. |
| March 1997 1 | | | - | Laboratory Reports | No | | Yes | |
| Intercontant Accounts of the Control | | | | | | | | |
| Company Mark Mark Company | | | | | | | | |
| Similar Security Control Contr | | | | | | | | |
| Controlled Control Process | Data Management | No errors in data transcription | | 10% check of electronically imported | Yes | - Indiana in the control of the cont | Yes | |
| County County Opening Memory County County Opening Memory County County Opening Memory County County Opening Memory County Count | | | | 100% check of manually entered data | | | | |
| Coulty Court of Protection Selection Court of Protection Court of | Data Useability | Limits of reporting less than investigation levels | Limits of reporting less than relevant | Results Tables | Yes | | Yes | |
| Many distributions of the standard production and all assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess to the standard production and assess the s | <u> </u> | | . – | | | | | |
| Many distributions of the standard production and all assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess the worked by resident and extension to improve the standard production and assess to the standard production and assess the s | Quelity Control 7 | Objectives C Mass. | A acoustones Oritaria | Henry (i.e. FORAT- | Accept | Notes/Datella of Na | Accept | Nata (Patrilla of Na |
| The District incoming of the Control control of the | Quality Control Process | Objectives & Measure | Acceptance Criteria | | | Notes/Details of Nonconformance | | Notes/Details of Nonconformance |
| Declaration from the management of the managemen | Field (Intra-laboratory) | Field Duplicate samples used assess the variability in analyte | Analysed for same chemicals as primary | | | Primary laboratory received sample | No No | RPD exceeded for Zinc (49%) in primary sample MW3 and |
| When experience of processing any support of processing any support of processing and processing of processing and processing | Duplicate Sampling and | concentration between samples collected from the sample | sample. | percent difference (RPD) results for | | | | duplicate sample QC102. Both results above adopted |
| usurates to furnificate active the ordinary and the production of production to furnificate active the ordinary active. See 201-128. Generally Principle (Control of the production of the prod | | | | field duplicate samples. | | | | assessment criteria, therefore does not alter interpretation of |
| Controlly feel absoluted Controlly feel absolute Controlly feel absoluted Controlly feel absolut | | | RPD <50% of mean conc. where both conc. | 1 | | | | |
| Scool by Hors Library (1994) The control of the Co | | undertaken to further assess precision level of precision. | | | | | | |
| Desiration Equation (Control Analysis) Analysis (Control Analysis) Analysis (Control (Control (Control Analysis) Analysis (Control | | | | | | | | |
| Librariany Control Strategy Librariany Strategy Processors of the Strategy Processors of S | | | | | | | | |
| Prof. Discovery Control Starting Listocology Control Starting Accordance Associated an analysis of the control Starting Associated an analysis of the control Starting Associated an analysis of the control Starting Associated an analysis of the control Associated analysis of the control A | | | | | No | | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1020195 |
| Position Continue | Duplicate Sampling and | Secondary duplicate samples sent to a secondary laboratory | sample. RPD <30% of mean conc. where both conc. | percent difference (RPD) results for | No | MW3 and triplicate sample QC201. No adopted assessment | N/A | |
| Find Prince Basis Programation & Control Control Interview Control Control Interview Control C | Duplicate Sampling and Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | sample. RPD <30% of mean conc. where both conc. >20 x LOR. | percent difference (RPD) results for | No | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not | N/A | |
| Programment of Analysis according food and the courty over from earpring against an expension of Analysis and Section of the search of the sea | Duplicate Sampling and Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. | percent difference (RPD) results for | No | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not | N/A | |
| Fig. Blank Surpring and Cross contamination between named abose not occur in female agreement planning genociative. Librorativy Applications Librorativy Applications Librorativy Applications Librorativy control discretion View View View View Librorativy control discretion View Librorativy control discretion View Librorativy control discretion View View Librorativy control discretion View Librorativy control discretion View Librorativy control discretion View Librorativy control discretion View Librorativy control | Duplicate Sampling and Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. | sample. RPD<30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. | | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. | | |
| Assystis transfer or as an antificit of the sampling handing procedure. Laboratory Duplicates Laboratory duplicates are used to bett the procision of the absoratory massurements. Laboratory Duplicates Laboratory duplicates are used to bett the procision of the absoratory massurements. Laboratory Control Samples (LCS) are used to easies so visual method performance in general these aserptices are unline. Dynamic monowy limits as specified by incomposition of the absoratory control samples (LCS) are used to easies so visual method performance in general these aserptices are unline. Donation of the processor of the processor of the absoratory control samples (LCS) are used to easies so visual method performance in general these aserptices are unline. Donation of the processor of the processor of the processor of the absoratory of analyses. Surregular Recovery Danagian Recovery Danagian Recovery All and its Spike and its are arrived processor in a sample spike of the sample applied with a income concentration of the sample applied processor in a sample applied with a income concentration of the sample applied processor in a sample applied processor in a sample applied with a income concentration of the sample applied with a income concentration of the sample applied with a income concentration of the sample applied processor in a sample applied with a income concentration of the sample applied | Duplicate Sampling and Analysis Field Rinsate Blank | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between | sample. RPD<30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. | | |
| As all positions of the sampling banding procedure. Laboratory Cupilitatis Veet Opinion recovery initia as specified by laboratory. Veet Veet Opinion recovery initia as specified by laboratory. As specified by laboratory (specifis opinion recovery initia as specified by laboratory (specifis opini | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. RPD<30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. | | |
| As all positions of the sampling banding procedure. Laboratory Cupilitatis Veet Opinion recovery initials as specified by Maloratory very records opinion recovery initials as specified by Maloratory very cupilitatis As specified by Maloratory As pecified by Maloratory As pecifie | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. RPD<30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. | | |
| Liboratory Duplicates Liboratory Duplicates Liboratory Duplicates Liboratory Centrel Samples Liboratory Centrel Samples Liboratory centrel samples (LCS) are used to assess overal ontopolity of performance, in general flows samples are aimini in composition to amylyse of eliment organization to any overance amenges are aimini in composition to amylyse of eliment of Mit samples are sognic compounds that are similar in characteristic composition to amylyse and eliment of environment organization to a supples of eliment so environment and person amyles are allocated or evaluate maniar information or evaluate maniar information are anythe anythe environment on environment anythe grant elements on environment and replays and the environment and are applicated or evaluate maniar information and anything and the environment and evaluate anything and elements or excessing the laboratory and personal manifest and evaluation and environment and evaluation and environment and evaluation and ev | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | |
| Laboratory Control Samples Laboratory Control Samples Laboratory Control Samples Annotogatemance, in general free samples as entails in composition to environmental camples, and ordinals hose composition to environmental camples, and ordinals hose conscient of the analysis and remeal. Controlled Reference Material Analysis occurred to the Substitution Reference Material Controlled Reference Material NA NA NA NA NA NA NA NA NA N | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Liboratory Control Samples (LiGS) are used to assess overall processing and performance in great angles as senior in whethough performance in great angles as senior in which get formance in great angles as senior in which get formance in great angles as senior in the control formance in great angles as senior in the control formance in great angles and control formance in great angles angles and control formance in great angles and control formance in great angles and control formance in great and angles and control formance and are spiked in control formance in a sample processing in any spike in the great and angles in the great and and spike in the great and angles in the great angles in the g | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 |
| method performance in general these samples are similar in aboratory. Committed for environmental supples, and contain known amounts of the analyses of interest. Committed Reference Material Ciff Mamphies are usuad to monthly the laboratory. Surrogate Recovery Surrogate Recovery Surrogate Recovery A matrix spike is an aliqued of a sample spiked with a known concentration and surgive prior composition to analyse of interest and are spiked size or environmental surgives, prior cample preparation and analysis. Surrogate recovers are used to evaluate matrix electrocy on a sample spiked with a known concentration of the sample preparation and analysis, and the results are used to evaluate matrix. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory reports and analysis and the results are used to analyse or specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory reports and analysed scalely like field samples. These blanks are used to represent the sample matrix. Recovery 70 - 130% or dynamic limits if specified by laboratory reports and scale specified by laboratory reports and scale specified by laboratory and or reported during sample preparation activities. Recovery 70 - 130% or dynamic limits if specified limits are internally consistent, consistent with held measurements, and consistent with held measurements. | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| method performance, in general tease samples are similar in aboratory, composition to environmental supples, and contain known amounts of the analyses of interest. Confided Relevence Material CPM samples are used for mortisof the accuracy of analyses performed by the laboratory of performed and seasons of the samples of interest. Surregate Recovery Surregate Recovery Surregate Recovery A marks spike is an alloyor of a sample spiked with a known preference on a sample spiked with a known analysis. Surregate recording are used to evaluate matrix releference on a sample spiked with a known analysis, and the results are used to evaluate matrix. A marks spike is an alloyor of a sample spiked with a known analysis, and the results are used to evaluate matrix. A marks spike is an alloyor of a sample spiked with a known analysis, and the results are used to evaluate matrix. A marks spike is an alloyor of a sample spiked with a known assess the bias of an amethod in a given sample matrix. Laboratory Method Blains as closely as possible and preparation and analysis, and the results are used to the allowabout of spikes are used to represent the sample matrix. Analyse concentration of the given analyse and the results are used to the allowabout of spikes are used to represent the sample matrix. A consistent with experimental and analysed accurity like field samples. These blanks are used to the allowabout on assess contamination introduced during sample proparation activities. Analyse concentration below LORs. Analyse concentration below LORs. Analyse concentration below LORs. Analyse of controlling the controlling controlling the spike and propagation and manalysed accurity like field samples. These blanks are used to represent the sample matrix. Concentration to a specific data price. These blanks are used to represent the sample matrix. Analyse concentration of the controlling the spike of the spike | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| composition to environmental samples, and outside in adaptive of interior and an adaptive. Surrogate Recovery Surrogate are used to montor the accuracy of analyses of interior and an adaptive. Surrogate composition to analyse of interest and an adaptive. Surrogate composition to an aprile perspectation and analysis. Surrogate recovering an eu used to evaluate matrix interference on a sample specified beside. Matrix Spike Recovery A matrix spike is an aliquor of a sample spiked with a known concentration of starget analysis (business are used to evaluate matrix interference on a sample specified beside). Matrix Spike Recovery A matrix spike is an aliquor of a sample spiked with a known concentration of starget analysis (c). Spiking occurs prior to sample preparation and analysis, and the results are used to evaluate or according to the spiked and analysed exactly like field samples. These barks are prepared to represent the sample matrix as disease, as prepared to represent the sample matrix as disease, as possible and prepared extracted dispated and analysed exactly like field samples. These barks are used by the lebosotory to asses contamination induced during sample preparation activities. Potentially Anomalous Data in No discrepancies between field (laboratory and/or expected and of historical). Analytical results are internally consistent, consistent with field measurements, and consistent with selected and of historical. | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Contributed Reference Matterial Composition to analytics of interest and are spiked into environmental samples prior to sample representation and analysis. Surplate recoveries are used to evaluate matrix interference on a sample-specific basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analysis (a). Spiking cours prior to sample preparation and analysis, and the results are used to sample analysis and the results are used to sampled county in the fact of sample propers and analysis and the results are used to the blackboatch of a given sample matrix. Laboratory Method Blanks as closely as possible and prepared for represent the sample matrix is closely as possible and prepared interfaced digested and analysed county like field samples. These blanks are used to the blackboatch or bases contamination introduced during sample preparation activities. Note that the contribute Reference on the sample matrix Laboratory reports Yes Laboratory reports Yes Laboratory reports Yes Laboratory reports Yes Yes Potentially Anomalous Data analyses county like field samples. These blanks are internally consistent, or consistent with field measurements, and | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| performed by the laboratory. secovery limits). Usually not performed and assessed based on LOS results. Surrogate Recovery Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recovered are used to evaluate matrix interference on a sample-specific basis. Matrix Spike Recovery Amatrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample perparation and analysis, and the results are used to evaluate matrix. Laboratory Method Blanks Method blanks are prepared for represent the sample matrix. Laboratory Method Blanks as closely as possible and prepared/estracted/digested and analysed caucity like field samples. These blanks are used to the hallowork to assess ochemically and prepared/estracted/digested and analysed caucity like field samples. These blanks are used to the hallowork to assess contential introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected occusions with intermediate occusions with expected and for historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Surrogate Recovery Surrogate are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples perior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analysis, and the results are used to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Liaboratory Method Blanks Method blanks are prepared for represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the liaboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, liaboratory and/or expected must be a consistent with field measurements, and consistent with pedial measurements. Analytical results are internally consistent, consistent with pedial measurements, and consistent with expected and for historical of resolutions. | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| chemical composition to analyties of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analytie(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks are prepared to represent the sample matrix. Laboratory Method Blanks are prepared to represent the sample matrix. Analyte concentrations below LORs. a closely as possible and prepared/extracted/digested and analysed exactly (life field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data results are identified No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with septendiand/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of inferest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| chemical composition to analytes of interest and are spiked with a foreign preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of farget analytic(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix. as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory is assess contamination introduced during sample preparation activities. Potentially Anomalous Data results are identified No discrepancies between field, laboratory and/or expected and/or historical consistent with field measurements, and consistent with septected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of inferest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-spoidic basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analysis, 3, said not results are used to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data results are identified No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, with expected and consistent with expected and for historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD store. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes N/A | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Interference on a sample-specific basis. Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with prepared and or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD st.OR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes N/A | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Matrix Spike Recovery A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory reports Yes Yes Ves Yes Potentially Anomalous Data No discrepancies between field, laboratory and/or expected or results are identified Analytical results are internally consistent, on sistent with field measurements, and consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD st.OR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes N/A | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified No discrepancies between field, laboratory and/or expected consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRIM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD st.OR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes N/A | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified No discrepancies between field, laboratory and/or expected consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRIM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD st.OR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes N/A | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| assess the bias of a method in a given sample matrix. Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD < 50% of mean conc. where both conc. >20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| Laboratory Method Blanks Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified No discrepancies between field, laboratory and/or expected consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to | sample. RPD < 50% of mean conc. where both conc. >20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified results are identified and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to | sample. RPD < 50% of mean conc. where both conc. >20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to | sample. RPD < 50% of mean conc. where both conc. >20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| the laboratory to assess contamination introduced during sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Ves | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| sample preparation activities. Potentially Anomalous Data No discrepancies between field, laboratory and/or expected results are identified Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Ves | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| results are identified consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory deplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Ves | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| results are identified consistent with field measurements, and consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to exactly like field samples. These blanks are used to mahysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Ves | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| consistent with expected and/or historical | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to exactly like field samples. These blanks are used to mahysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Ves | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |
| | Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD < 50% of mean conc. where both conc. >20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD < 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes Yes | MW3 and triplicate sample QC201. No adopted assessment criteria for total phosphorus in groundwater, therefore does not impact upon interpretation of results. Primary laboratory received sample | Yes No Yes Yes Yes Yes Yes | in batch 1020195. QC302 was reported above LOR for Manganese (0.038 mg/L). This is not seen to impact on the data as this |



| Job Number: | S20102 |
|---------------|--|
| Report Title: | Surface Water and Groundwater Monitoring |
| Client: | ReDirect Recycling |
| Completed By: | Bec Chapple |
| Date: | 9-Aug-24 |
| Verified By: | Emma Walsh |
| Date: | 11-Sep-24 |

| | | 1 | |
|------------------------------------|-----------|------------------------------------|-----------|
| SAMPLE DELIVERY GROUP (SDG): | 1020195 | SAMPLE DELIVERY GROUP (SDG): | ES2403942 |
| Laboratory: | Eurofins | Laboratory: | ALS |
| Sample Dates: | 14-Aug-23 | Sample Dates: | 15-Feb-24 |
| Sample Media: | Water | Sample Media: | Water |

| | Objectives & Measure | Acceptance Criteria | Source of Information | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
|--|---|--|--|---------------------------|--|-------------------------------|--|
| Process Standard Procedures | Standard field sampling procedures and forms used | No deviation from standard procedure and | Borelogs, field sheets, COCs, data | Criteria Met? Yes | | Criteria Met? Yes | |
| | | forms used. | tables | | | Vac | |
| Equipment Calibration | All equipment calibrated in accordance with manufacturers specifications | All equipment calibrated in accordance with manufacturers specifications. | Calibration Certificates / Records | Yes | | Yes | |
| Testing Method Accreditation | NATA accredited methods used for all analyses determined | Primary and secondary laboratories to use NATA accredited methods for all analytes | Laboratory Report | Yes | | Yes | |
| | | determined. | | | | | |
| Quality Control Sampling Frequency | Field QC sampling frequency in accordance with AS4482.1- 2005 | Field (Intra-laboratory) Duplicates - ≥ 1 in 20 primary samples. | QA/QC register (within field book) | N/A | Primary laboratory received sample | N/A | Relevant intra-laboratory QC samples for this WME reported in batch ES2404239. |
| | | (note that PFAS NEMP recommends 1 in 10 | | | | | |
| | | for PFAS investigations) Secondary (inter-laboratory) duplicates - ≥ 1 in | QA/QC register (within field book) | Yes | QC202 | N/A | Relevant inter-laboratory QC samples for this WME reported |
| | | 20 primary samples. (note that PFAS NEMP recommends 1 in 10 | | | | | in batch 1067666. |
| | | for PFAS investigations) | 0.1/02 | N/+ | Dimentity | N/C | Dispate |
| | | Rinsate Blanks - ≥ 1 per day, per matrix per equipment. | QA/QC register (within field book) | N/A | Primary laboratory received sample | N/A | Rinsate not required as only surface water was collected straight into the bottles |
| | | | OA/OC radiator finishing | N/A | Primary Jahoraton reactived as well | Vec | |
| | | Trip Blanks - \geq 1 per esky containing samples for volatiles. | | N/A | Primary laboratory received sample | Yes | QC403 |
| | Laboratory QC analysis frequency in accordance with NEPC 2013 | Laboratory Duplicates - at least 1 in 10 analyses or 1 per process batch. | Laboratory Reports | Yes | | No | A laboratory duplicate for PAH/Phenols and TRH- Semivolatile fraction was not analysed, resulting in a non- |
| | | , comment | | | 1 | | conformance for frequency for this analysis. Not considered |
| | | | | | 1 | | to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory |
| | | | | | | | field duplicates were analysed for PAH/ phenols and TRH |
| | | | | | | | semivolatile fraction and were DQI compliant. |
| | | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | Surrogate Recoveries - all samples spiked | Laboratory Reports | Yes | | Yes | |
| | | where appropriate (e.g. chromatographic analysis of organics). | | | | | <u> </u> |
| | | Laboratory Control Samples - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | Matrix Spikes - at least 1 per matrix type per | Laboratory Reports | Yes | | No | A matrix spike for PAH/PhenoIs and TRH-Semivolatile |
| | | process batch. | | | | | fraction was not analysed, resulting in a non-conformance for frequency for this analysis. Not considered to impact |
| | | | | | | | upon assessment of accuracy, precision and comparability |
| | | | | | | | since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH |
| | | | | | | | semivolatile fraction and were DQI compliant. |
| Sample Preservation, | Samples appropriately preserved upon collection, stored and | | Laboratory Reports | Yes | | Yes | |
| Handling and Holding Times | | requirements. Unless specific method indicates otherwise, | | | | | |
| | | soil and water samples should be stored, | | | | | |
| L | | transported and received by the laboratory at $\!<\!6^{\circ}\text{C}.$ | | | | | <u> </u> |
| Data Management | No errors in data transcription | Entry of field data verified by peer. | 10% check of electronically imported | Yes | | Yes | |
| | | | data (e.g. ESDAT). 100% check of manually entered data | | 1 | | l i |
| | | | (e.g. field parameters, gauging data). | | | | |
| Data Useability | Limits of reporting less than investigation levels | Limits of reporting less than relevant | Results Tables | Yes | | Yes | |
| | | investigation levels. | | | | | |
| Quality Control Process | Objectives & Measure | Acceptance Criteria | How? (i.e. ESDAT output, review | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| | | | lab reports, review data etc) | Criteria Met? | | Criteria Met? | |
| Field (Intra-laboratory) | Field Duplicate samples used assess the variability in analyte | e Analysed for same chemicals as primary | | | | | |
| Duplicate Sameling | | | ESDAT generated summary of relative percent difference (RPD) results for | N/A | Primary laboratory received sample | N/A | Relevant intra-laboratory QC samples for this WME reported in batch ES2404239. |
| Duplicate Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. | sample. RPD <30% of mean conc. where both conc. | ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. | N/A | Primary laboratory received sample | N/A | Relevant intra-laboratory QC samples for this WME reported in batch ES2404239. |
| | concentration between samples collected from the sample | sample. | percent difference (RPD) results for | N/A | Primary laboratory received sample | N/A | |
| | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR | percent difference (RPD) results for | N/A | Primary laboratory received sample | N/A | |
| Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. | sample. RPD x30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR | percent difference (RPD) results for field duplicate samples. | | | | reported in batch ES2404239. |
| | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for | | Primary laboratory received sample RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc | N/A | |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative | | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted | | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory | sample. RPD 430% of mean conc. where both conc. >20 x LOR RPD 450% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD 430% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for | | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc | | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD To olimit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. |) No | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter | | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD To olimit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. |) No | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD To olimit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field |) No | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD To olimit where both conc. < 10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. |) No | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <80% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD to limit where both conc. <10 x LOR. Analyse concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR. RPD <30% of mean conc. where both conc. >20 x LOR. RPD 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | NO NO N/A | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <80% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD to limit where both conc. <10 x LOR. Analyse concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR. RPD <30% of mean conc. where both conc. >20 x LOR. RPD 50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | NO NO N/A | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <30% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyste concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | NO NO N/A | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | N/A N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. Analyse concentrations below LORs. Analyse concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR. RPD <30% of mean conc. where both conc. >20 x LOR. RPD on limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD do limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | N/A N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. <20 x LOR RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD oll limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD to Slow of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD to Slow of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD to limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix | sample. RPD <50% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD to Slow of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | N/A N/A Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD ×30% of mean conc. where both conc. >20 x LOR. RPD ×30% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory outplicates are used to test the precision of the laboratory measurements. Laboratory outplicates are used to test the precision of the laboratory on the laboratory of the laboratory. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD ×30% of mean conc. where both conc. >20 x LOR. RPD ×30% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes | reported in batch ES2404239. Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD ×30% of mean conc. where both conc. >20 x LOR. RPD ×30% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes N/A Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation of target analyte(s), spiking occurs prior to sample preparation of target analytes, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested and | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <30% of mean conc. where both conc. >10-20 x LOR. RPD on limit where both conc. <10 x LOR. Analyse concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A N/A N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <30% of mean conc. where both conc. >10-20 x LOR. RPD on limit where both conc. <10 x LOR. Analyse concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A N/A N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <30% of mean conc. where both conc. >10-20 x LOR. RPD on limit where both conc. <10 x LOR. Analyse concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A N/A N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to eases see the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No bimit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD down of mean conc. where both conc. 10-20 x LOR. RPD soll of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | No No N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes NO No Yes | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to eases see the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No ilmit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD to limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes N/A N/A N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analytes (5). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <50% of mean conc. where both conc. >220 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR Analysed for same chemicals as primary sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. >20 x LOR. RPD to solve of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | No No N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes NO No Yes | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |
| Analysis Secondary Inter-laborator) Duplicate Sampling and Analysis Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analytes (5). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No imit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD No limit where both conc. <10 x LOR Analysed for same chemicals as primary sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD stolk of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of relative percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | No No N/A N/A Yes Yes Yes | RPD exceedance for iron (84%) and zinc (35%) in primary sample in MW3 and triplicate sample QC202. Both zinc results above adopted assessment criteria, and no adopted assessment criteria for iron, therefore does not alter interpretation of results. Primary laboratory received sample | N/A N/A Yes Yes Yes NO No Yes | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. Rinsate not required as only surface water was collected straight into the bottles Mattix spike recovery not determined for zinc, nitrite and nitrate as N and total phosphorus as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this |



| Job Number: | S20102 |
|---------------|--|
| Report Title: | Surface Water and Groundwater Monitoring |
| Client: | ReDirect Recycling |
| Completed By: | Bec Chapple |
| Date: | 9-Aug-24 |
| | Emma Walsh |
| Verified By: | LITITIA VVAIGIT |

| SAMPLE DELIVERY GROUP (SDG): | ES2404239 | SAMPLE DELIVERY GROUP (SDG): | ES2404752 |
|------------------------------------|-----------|------------------------------------|-----------|
| Laboratory: | ALS | Laboratory: | ALS |
| Sample Dates: | 15-Feb-24 | Sample Dates: | 20-Feb-24 |
| Sample Media: | Water | Sample Media: | Water |

| Quality Assurance Process | | | | | | | |
|--|---|---|---|-----------------------------|---|-----------------------------|--|
| | Objectives & Measure | Acceptance Criteria | Source of Information | Acceptance Criteria Met? | Notes/Details of Nonconformance | Acceptance Criteria Met? | Notes/Details of Nonconformance |
| Standard Procedures | Standard field sampling procedures and forms used | No deviation from standard procedure and forms used. | Borelogs, field sheets, COCs, data tables | Yes | | Yes | |
| Equipment Calibration | All equipment calibrated in accordance with manufacturers | All equipment calibrated in accordance with | Calibration Certificates / Records | Yes | | Yes | |
| Testing Method | specifications NATA accredited methods used for all analyses determined | manufacturers specifications. Primary and secondary laboratories to use | Laboratory Report | Yes | | Yes | |
| Accreditation | | NATA accredited methods for all analytes determined. | | | | | |
| Quality Control Sampling | Field QC sampling frequency in accordance with AS4482.1- | Field (Intra-laboratory) Duplicates - ≥ 1 in 20 | QA/QC register (within field book) | Yes | QC103 | N/A | Relevant intra-laboratory QC samples for this WME |
| Frequency | 2005 | primary samples. (note that PFAS NEMP recommends 1 in 10 | | | | | reported in batch ES2304011. |
| | | for PFAS investigations) Secondary (inter-laboratory) duplicates - ≥ 1 in | OA/OC register (within field book) | N/A | Relevant inter-laboratory QC samples for this WME reported | N/Δ | Relevant inter-laboratory QC samples for this WME reported |
| | | 20 primary samples. | The second second | | in batch 1067666. | | in batch 1067666. |
| | | (note that PFAS NEMP recommends 1 in 10 for PFAS investigations) | | | | | |
| | | Rinsate Blanks - ≥ 1 per day, per matrix per equipment. | QA/QC register (within field book) | Yes | QC303 | N/A | Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day |
| | | ециртиент. | | | | | |
| | | Trip Blanks - ≥ 1 per esky containing samples for volatiles. | QA/QC register (within field book) | Yes | QC404 | N/A | No blank required due to limited number of samples (only MW4) |
| | Laboratory QC analysis frequency in accordance with NEPC 2013 | Laboratory Duplicates - at least 1 in 10 | Laboratory Reports | No | A laboratory duplicate for PAH/Phenols and TRH- | No | A laboratory duplicate for PAH/Phenols, TRH-Semivolatile |
| | 2013 | analyses or 1 per process batch. | | | Semivolatile fraction was not analysed, resulting in a non- conformance for frequency for this analysis. Not considered | | fraction and dissolved mercury was not analysed, resulting in a non-conformance for frequency for this analysis. Not |
| | | | | | to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory | | considered to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and |
| | | | | | field duplicates were analysed for PAH/ phenols and TRH | | inter-laboratory field duplicates were analysed for PAH/ |
| | | | | | semivolatile fraction and were DQI compliant. | | phenols and TRH semivolatile fraction and were DQI compliant. |
| | | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | Surrogate Recoveries - all samples spiked | Laboratory Reports | Yes | | Yes | |
| | | where appropriate (e.g. chromatographic analysis of organics). | | | | | |
| | | Laboratory Control Samples - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | Matrix Spikes - at least 1 per matrix type per | Laboratory Reports | No | A matrix spike for PAH/Phenols and TRH-Semivolatile | No | A matrix spike for PAH/Phenols, TRH-Semivolatile fraction |
| | | process batch. | | | fraction was not analysed, resulting in a non-conformance for frequency for this analysis. Not considered to impact | | and dissolved metals was not analysed, resulting in a non- conformance for frequency for this analysis. Not considered |
| | | | | | upon assessment of accuracy, precision and comparability | | to impact upon assessment of accuracy, precision and |
| | | | | | since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH | | comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH |
| | | | | | semivolatile fraction and were DQI compliant. | | semivolatile fraction and were DQI compliant. |
| Sample Preservation, | Samples appropriately preserved upon collection, stored and | In accordance with laboratory specific method | Laboratory Reports | Yes | | Yes | |
| Handling and Holding Times | transported, and analysed within holding times | requirements. Unless specific method indicates otherwise, | | | | | |
| | | soil and water samples should be stored, | | | | | |
| | | transported and received by the laboratory at < 6°C. | | | | | |
| Data Management | No errors in data transcription | Entry of field data verified by peer. | 10% check of electronically imported | Yes | | Yes | |
| | | | data (e.g. ESDAT). 100% check of manually entered data | | | | |
| | | | (e.g. field parameters, gauging data). | | | | |
| Data Useability | Limits of reporting less than investigation levels | Limits of reporting less than relevant | Results Tables | Yes | | Yes | |
| | | investigation levels. | | | | | |
| Quality Control Process | Objectives & Measure | Acceptance Criteria | How? (i.e. ESDAT output, review | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| Field (Intra-laboratory) | Field Duplicate samples used assess the variability in analyte | Analysed for same chamicals as primary | lab reports, review data etc) ESDAT generated summary of relative | Criteria Met? Yes | | Criteria Met? | Relevant intra-laboratory QC samples for this WME |
| Duplicate Sampling and | concentration between samples collected from the sample | sample. | percent difference (RPD) results for | 165 | | IN/A | reported in batch ES2304011. |
| Analysis | location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed | RPD <30% of mean conc. where both conc. >20 x LOR | field duplicate samples. | | | | |
| | samples for chemicals within their holding times may be | RPD <50% of mean conc. where both conc. 10-20 x LOR | | | | | |
| | undertaken to further assess precision level of precision. | RPD No limit where both conc. < 10 x LOR | | | | | |
| Secondary Inter-laborator) | Results are accurate and free from laboratory error. | A set send for come about leads on selection | ESDAT generated summary of relative | | | | |
| Duplicate Sampling and Analysis | | Analysed for same chemicals as primary | LODAT generated summary of relative | N/A | Relevant inter-laboratory QC samples for this WME reported | N/A | Relevant inter-laboratory QC samples for this WME reported |
| | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | sample. | percent difference (RPD) results for | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1067666. |
| | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. | | N/A | * ' ' ' | N/A | |
| | to assess the accuracy of the analyte concentrations reported | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. | percent difference (RPD) results for | N/A | * ' ' ' | N/A | |
| | to assess the accuracy of the analyte concentrations reported by the primary laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for | | in batch 1067666. | | in batch 1067666. |
| | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. | percent difference (RPD) results for field duplicate samples. | N/A | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | | |
| Field Rinsate Blank | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a | | in batch 1067666. Rinsate not required as only one sample collected using |
| Field Rinsate Blank Preparation & Analysis | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | No | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day |
| Field Rinsate Blank | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | | in batch 1067666. Rinsate not required as only one sample collected using |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | No | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | No | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | No Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of similar where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of similar where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment | N/A N/A Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1087666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. | N/A N/A Yes Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of similar where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. | N/A N/A Yes Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. | N/A N/A Yes Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment oriteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project | N/A N/A Yes Yes | in batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD store. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested and analysed scatcly like field samples. These blanks are used by the laboratory to assess contamination introduced during the laboratory to assess contamination introduced during | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD store. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD storm of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LOS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested and analysed scatcly like field samples. These blanks are used by the laboratory to assess contamination introduced during the laboratory to assess contamination introduced during | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD storm of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LOS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery Laboratory Method Blanks | to assess the accuracy of the analyte concentrations reponded by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory of seasess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes No Yes Yes Ves | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |
| Field Rinsate Blank Preparation & Analysis Trip Blank Sampling and Analysis Laboratory Duplicates Laboratory Control Samples Certified Reference Material Surrogate Recovery Matrix Spike Recovery | to assess the accuracy of the analyte concentrations reported by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes No Yes Yes Ves | in batch 1067666. Total phosphorus (as P) was above the LOR. Not seen as a significant impact to results as no adopted assessment criteria for phosphorus in groundwater. Mattix spike recovery not determined for ammonia (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of | N/A N/A Yes Yes N/A Yes | In batch 1067666. Rinsate not required as only one sample collected using hydrasleeves- dedicated sampling equipment for the day No blank required due to limited number of samples (only MW4) Mattix spike recovery not determined for manganese as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to impact the accuracy of the |



| Job Number: | S20102 |
|---------------|--|
| Report Title: | Surface Water and Groundwater Monitoring |
| Client: | ReDirect Recycling |
| | |
| Completed By: | Bec Chapple |
| Date: | 9-Aug-24 |
| Verified By: | Emma Walsh |
| Date: | 11-Sep-24 |

| SAMPLE DELIVERY GROUP (SDG): | 1067666 | SAMPLE DELIVERY GROUP (SDG): | ES2422553 |
|------------------------------------|-----------|------------------------------------|-----------|
| Laboratory: | Eurofins | Laboratory: | ALS |
| Sample Dates: | 12-Feb-24 | Sample Dates: | 16-Jul-24 |
| Sample Media: | Water | Sample Media: | Water |

| Quality Assurance | Objectives & Measure | Acceptance Criteria | Source of Information | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
|--|--|--|---|----------------------|------------------------------------|----------------------|--|
| Process Standard Procedures | Standard field sampling procedures and forms used | No deviation from standard procedure and | Borelogs, field sheets, COCs, data | Criteria Met? Yes | | Criteria Met? Yes | |
| | | forms used. | tables | | | Voc | |
| Equipment Calibration | All equipment calibrated in accordance with manufacturers specifications | All equipment calibrated in accordance with manufacturers specifications. | Calibration Certificates / Records | Yes | | Yes | |
| Testing Method Accreditation | NATA accredited methods used for all analyses determined | Primary and secondary laboratories to use NATA accredited methods for all analytes | Laboratory Report | Yes | | Yes | |
| | | determined. | | | | | |
| Quality Control Sampling Frequency | Field QC sampling frequency in accordance with AS4482.1- 2005 | Field (Intra-laboratory) Duplicates - ≥ 1 in 20 primary samples. | QA/QC register (within field book) | N/A | Primary laboratory received sample | N/A | Relevant intra-laboratory QC samples for this WME reported in batch ES2423038. |
| | | (note that PFAS NEMP recommends 1 in 10 | | | | | |
| | | for PFAS investigations) Secondary (inter-laboratory) duplicates - ≥ 1 in | QA/QC register (within field book) | Yes | QC203 | N/A | Relevant inter-laboratory QC samples for this WME reported |
| | | 20 primary samples. | | | | | in batch 1117968. |
| | | (note that PFAS NEMP recommends 1 in 10 for PFAS investigations) | | | | | |
| | | Rinsate Blanks - ≥ 1 per day, per matrix per equipment. | QA/QC register (within field book) | N/A | Primary laboratory received sample | N/A | Rinsate not required as only surface water was collected straight into the bottles |
| | | | | | | | |
| | | Trip Blanks - \geq 1 per esky containing samples for volatiles. | QA/QC register (within field book) | N/A | Primary laboratory received sample | Yes | QC405 |
| | Laboratory QC analysis frequency in accordance with NEPC | Laboratory Duplicates - at least 1 in 10 | Laboratory Reports | Yes | | No | A laboratory duplicate for PAH/Phenols and TRH- |
| | 2013 | analyses or 1 per process batch. | | | | | Semivolatile fraction was not analysed, resulting in a non- conformance for frequency for this analysis. Not considered to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH semivolatile fraction and were DQI compliant. |
| | | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | |
| | | Surrogate Decayories all complex spiked | Laboratory Reports | Voc | | Yes | |
| | | Surrogate Recoveries - all samples spiked where appropriate (e.g. chromatographic | Laboratory Reports | Yes | | Yes | |
| | | analysis of organics). Laboratory Control Samples - at least 1 per | Laboratory Reports | Yes | | Yes | |
| | | process batch. | | Yes | | Yes | |
| Spends Press 1 | | Matrix Spikes - at least 1 per matrix type per process batch. | Laboratory Reports | Yes | | No | A matrix spike for PAH/Phenols and TRH-Semivolatile fraction was not analysed, resulting in a non-conformance for frequency for this analysis. Not considered to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH semivolatile fraction and were DQI compliant. |
| Sample Preservation, Handling and Holding Times | Samples appropriately preserved upon collection, stored and transported, and analysed within holding times | In accordance with laboratory specific method requirements. | Laboratory Reports | Yes | | Yes | |
| | | Unless specific method indicates otherwise, soil and water samples should be stored, transported and received by the laboratory at < 6°C. | | | | | |
| Data Management | No errors in data transcription | Entry of field data verified by peer. | 10% check of electronically imported data (e.g. ESDAT). | Yes | | Yes | |
| | | | 100% check of manually entered data | | | | |
| | | | (e.g. field parameters, gauging data). | | | | |
| Data Useability | Limits of reporting less than investigation levels | Limits of reporting less than relevant | Results Tables | Yes | | Yes | |
| | | investigation levels. | <u> </u> | | <u></u> | | |
| Quality Control Process | Objectives & Measure | Acceptance Criteria | How? (i.e. ESDAT output, review | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| | | | lab reports, review data etc) | Criteria Met? | | Criteria Met? | |
| Field (Intra-laboratory) Duplicate Sampling and | Field Duplicate samples used assess the variability in analyte concentration between samples collected from the sample | Analysed for same chemicals as primary sample. | ESDAT generated summary of relative percent difference (RPD) results for | N/A | Primary laboratory received sample | N/A | Relevant intra-laboratory QC samples for this WME reported in batch ES2423038. |
| Analysis | Coation and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess precision level of precision. | RPD <30% of mean conc. where both conc. >20 x LOR RPD <50% of mean conc. where both conc. 10-20 x LOR RPD No limit where both conc. < 10 x LOR | field duplicate samples. | | | | |
| Secondary Inter-laborator) | Results are accurate and free from laboratory error. | Analysed for same chemicals as primary | ESDAT generated summary of relative | Yes | | N/A | Relevant inter-laboratory QC samples for this WME reported |
| Duplicate Sampling and Analysis | Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported | sample. RPD <30% of mean conc. where both conc. | percent difference (RPD) results for field duplicate samples. | | | | in batch 1117968. |
| | by the primary laboratory. | >20 x LOR. | | | | | |
| | | RPD <50% of mean conc. where both conc. 10-20 x LOR. | | | | | |
| Field Disease Plant | Cross contamination of samel-s does not | RPD no limit where both conc. < 10 x LOR. | ESDAT described | N/A | Driman laborator received country | N/A | Dineste not required as eath a set- |
| Field Rinsate Blank Preparation & Analysis | Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | Analyte concentrations below LORs. | ESDAT generated summary of field blank analytical results. | N/A | Primary laboratory received sample | N/A | Rinsate not required as only surface water was collected straight into the bottles |
| Trip Blank Sampling and | Cross contamination between samples does not occur in | Analyte concentrations below LORs. | ESDAT generated summary of field | N/A | Primary laboratory received sample | Yes | |
| Analysis | transit or as an artefact of the sampling handling procedure. | | blank analytical results. | | | | |
| Laboratory Duplicates | Laboratory duplicates are used to test the precision of the laboratory measurements. | As specified by laboratory. | Laboratory reports | Yes | | Yes | |
| Laboratory Control Samples | method performance. In general these samples are similar in composition to environmental samples, and contain known | Dynamic recovery limits as specified by laboratory. | Laboratory reports | Yes | | Yes | |
| Certified Reference Material | amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | Laboratory reports | N/A | | N/A | |
| Surrogate Recovery | Surrogates are organic compounds that are similar in | Dynamic recovery limits as specified by | Laboratory reports | Yes | | Yes | |
| | chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | laboratory. | | | | | |
| Matrix Spike Recovery | A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | Recovery 70 - 130% or dynamic limits if specified by laboratory. | Laboratory reports | Yes | | Yes | |
| Laboratory Method Blanks | Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | Analyte concentrations below LORs. | Laboratory reports | Yes | | Yes | |
| Potentially Anomalous Data | No discrepancies between field, laboratory and/or expected results are identified | Analytical results are internally consistent, consistent with field measurements, and consistent with expected and/or historical results based on CSM | Multiple sources | Yes | | Yes | |



| Job Number: | S20102 |
|------------------------|-------------------------------|
| | Surface Water and Groundwater |
| Report Title: | Monitoring |
| Client: | ReDirect Recycling |
| | |
| | |
| Completed By: | Bec Chapple |
| Completed By: Date: | Bec Chapple 9-Aug-24 |
| | |

| SAMPLE DELIVERY GROUP (SDG): | ES2423038 | SAMPLE DELIVERY GROUP (SDG): | 1117968 |
|------------------------------------|-----------|------------------------------------|-----------|
| Laboratory: | ALS | Laboratory: | Eurofins |
| Sample Dates: | 22-Jul-24 | Sample Dates: | 15-Jul-24 |
| Sample Media: | Water | Sample Media: | Water |

| Objectives & Measure | | | | | | |
|--|--|---|----------------------------|--|--------------------------|---|
| | Acceptance Criteria | Source of Information | Acceptance | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| | | | Criteria Met? | | Criteria Met? | |
| Standard field sampling procedures and forms used | No deviation from standard procedure and forms used. | Borelogs, field sheets, COCs, data tables | Yes | | Yes | V . |
| All equipment calibrated in accordance with manufacturers | All equipment calibrated in accordance with | tables Calibration Certificates / Records | Yes | | Yes | |
| specifications | manufacturers specifications. | | | | | |
| NATA accredited methods used for all analyses determined | Primary and secondary laboratories to use NATA accredited methods for all analytes | Laboratory Report | Yes | 1 | Yes | l l |
| Field OC compling for | determined. | 04/00 mminter () | Vec | 0010/ | N/A | Primary Johannia |
| Field QC sampling frequency in accordance with AS4482.1- 2005 | Field (Intra-laboratory) Duplicates - ≥ 1 in 20 primary samples. | QA/QC register (within field book) | Yes | QC104 | N/A | Primary laboratory received sample |
| | (note that PFAS NEMP recommends 1 in 10 | | | 1 | | 1 |
| | for PFAS investigations) Secondary (inter-laboratory) duplicates - ≥ 1 in | QA/QC register (within field book) | N/A | Relevant inter-laboratory QC samples for this WME reported | Yes | QC204 |
| | 20 primary samples. | | | in batch 1117968. | | l l |
| | (note that PFAS NEMP recommends 1 in 10 | | | 1 | | l l |
| | for PFAS investigations) Rinsate Blanks - ≥ 1 per day, per matrix per | QA/QC register (within field book) | Yes | QC304 | N/A | Primary laboratory received sample |
| | equipment. | | | 1 | | |
| | Trip Blanks - ≥ 1 per esky containing samples | QA/QC register (within field book) | Yes | QC405 | N/A | Primary laboratory received sample |
| Laborator 22 | for volatiles. | | | | | |
| Laboratory QC analysis frequency in accordance with NEPC 2013 | Laboratory Duplicates - at least 1 in 10 analyses or 1 per process batch. | Laboratory Reports | No | A laboratory duplicate for PAH/Phenols and TRH- Semivolatile fraction was not analysed, resulting in a non- | Yes | |
| | , , , | | | conformance for frequency for this analysis. Not considered | | İ |
| | | | | to impact upon assessment of accuracy, precision and comparability since the intra-laboratory and inter-laboratory | | l l |
| | | | | comparability since the intra-laboratory and inter-laboratory field duplicates were analysed for PAH/ phenols and TRH | | l l |
| | | | | semivolatile fraction and were DQI compliant. | | |
| | Method Blanks - at least 1 per process batch. | Laboratory Reports | Yes | | Yes | 1 |
| | | | | | | |
| | Surrogate Recoveries - all samples spiked where appropriate (e.g. chromatographic | Laboratory Reports | Yes | | Yes | |
| | analysis of organics). | | | | | |
| | Laboratory Control Samples - at least 1 per | Laboratory Reports | Yes | 1 | Yes | |
| | process batch. Matrix Spikes - at least 1 per matrix type per | Laboratory Reports | No | A matrix spike for PAH/Phenols and TRH-Semivolatile | Yes | |
| | process batch. | | | fraction was not analysed, resulting in a non-conformance | | |
| | | | | for frequency for this analysis. Not considered to impact upon assessment of accuracy, precision and comparability | | |
| | | | | since the intra-laboratory and inter-laboratory field | | l l |
| | | | | duplicates were analysed for PAH/ phenols and TRH semivolatile fraction and were DQI compliant. | | |
| | | | | DQI compliant. | | |
| Samples appropriately preserved upon collection, stored and transported, and analysed within holding times | | Laboratory Reports | Yes | | Yes | |
| transported, and analysed within holding times | requirements. Unless specific method indicates otherwise, | | | 1 | | |
| | soil and water samples should be stored, | | | 1 | | |
| | transported and received by the laboratory at < 6°C. | | | 1 | | |
| No errors in data transcription | 6°C. Entry of field data verified by peer. | 10% check of electronically imported | Yes | | Yes | |
| | | data (e.g. ESDAT). | | 1 | | |
| | | 100% check of manually entered data (e.g. field parameters, gauging data). | | 1 | | |
| | | | | | | |
| Limits of reporting less than investigation levels | Limits of reporting less than relevant investigation levels. | Results Tables | Yes | | Yes | |
| | | | | | | |
| Objectives & Measure | Acceptance Criteria | How? (i.e. ESDAT output, review | | Notes/Details of Nonconformance | Acceptance | Notes/Details of Nonconformance |
| | | lab reports, review data etc) | Criteria Met? | | Criteria Met? | |
| Field Duplicate samples used assess the variability in analyte concentration between samples collected from the sample | e Analysed for same chemicals as primary sample | ESDAT generated summary of relative | Yes | | N/A | Primary laboratory received sample |
| concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. | sample. RPD <30% of mean conc. where both conc. | percent difference (RPD) results for field duplicate samples. | | 1 | | |
| Where required, resubmission of previously analysed | >20 x LOR | 1 | | 1 | | |
| samples for chemicals within their holding times may be undertaken to further assess precision level of precision. | RPD <50% of mean conc. where both conc. 10-20 x LOR | | | 1 | | |
| | RPD No limit where both conc. < 10 x LOR |] | | 1 | | ų l |
| Results are accurate and free from laboratory error. | Analysed for same chemicals as primary | | | | | <u> </u> |
| Secondary duplicate samples sent to a secondary laboratory | | | N/A | Relevant inter-laboratory QC samples for this WME reported | Yes | |
| to accept the accurrent of the | sample. | percent difference (RPD) results for | N/A | Relevant inter-laboratory QC samples for this WME reported in batch 1117968. | Yes | |
| to assess the accuracy of the analyte concentrations reported by the primary laboratory. | sample. d RPD <30% of mean conc. where both conc. >20 x LOR. | | N/A | | Yes | |
| | sample. d RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. | percent difference (RPD) results for | N/A | | Yes | |
| | sample. d RPD <30% of mean conc. where both conc. >20 x LOR. | percent difference (RPD) results for | N/A | | Yes | |
| by the primary laboratory. Cross contamination of samples does not occur between | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | N/A | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total | Yes | Primary laboratory received sample |
| by the primary laboratory. | sample. JRPD 430% of mean conc. where both conc. >20 xLOR. RPD 450% of mean conc. where both conc. 10-20 xLOR. RPD no limit where both conc. < 10 xLOR. | percent difference (RPD) results for field duplicate samples. | | in batch 1117968. | | Primary laboratory received sample |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. JRPD 430% of mean conc. where both conc. >20 xLOR. RPD 450% of mean conc. where both conc. 10-20 xLOR. RPD no limit where both conc. < 10 xLOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a | | Primary laboratory received sample |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. | No | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling | sample. JRPD 430% of mean conc. where both conc. >20 xLOR. RPD 450% of mean conc. where both conc. 10-20 xLOR. RPD no limit where both conc. < 10 xLOR. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field | | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | | Primary laboratory received sample Primary laboratory received sample |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | No | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field | No | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. | sample. (RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the | sample. (RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known | sample. (RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. | No Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | No Yes Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A No Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | No Yes Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A No Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | No Yes Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A No Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of initial where both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A NO Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A NO Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples of interest and are spiked into environmental samples prior to sample preparation and | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted | N/A N/A NO Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <30% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of LOR. RPD of LOR. RPD not limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. | N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of third where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports | Yes Yes N/A | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. | N/A N/A NO Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 1117968. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. | N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used | N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes Yes Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of bill with the both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s), Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digestales are used to the laboratory to assess contamination introduced during the laboratory to assess contamination introduced during | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of bill with the both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory measurements. Laboratory measurements. Laboratory measurements. Laboratory measurements. Laboratory measurements. CRM samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of bill with the both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like feld samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of thirt where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes No Yes Yes Ves Ves | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A No Yes Yes Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to assess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during the laboratory to assess contamination introduced during | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of bill with the both conc. <10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes N/A Yes | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A NO Yes N/A Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to eassess the bias of a method in a given sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD of third where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analyte concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes No Yes Yes Ves Ves | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A No Yes Yes Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |
| by the primary laboratory. Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment. Cross contamination between samples does not occur in transit or as an artefact of the sampling handling procedure. Laboratory duplicates are used to test the precision of the laboratory measurements. Laboratory control samples (LCS) are used to assess overall method performance. In general these samples are similar in composition to environmental samples, and contain known amounts of the analytes of interest. CRM samples are used to monitor the accuracy of analyses performed by the laboratory. Surrogates are organic compounds that are similar in chemical composition to analytes of interest and are spiked into environmental samples prior to sample preparation and analysis. Surrogate recoveries are used to evaluate matrix interference on a sample-specific basis. A matrix spike is an aliquot of a sample spiked with a known concentration of target analyte(s). Spiking occurs prior to sample preparation and analysis, and the results are used to eassess the bias of a method in a given sample matrix. Method blanks are prepared to represent the sample matrix as closely as possible and prepared/extracted/digested and analysed exactly like field samples. These blanks are used by the laboratory to assess contamination introduced during sample preparation activities. | sample. RPD <50% of mean conc. where both conc. >20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD <50% of mean conc. where both conc. 10-20 x LOR. RPD no limit where both conc. < 10 x LOR. Analyte concentrations below LORs. Analyte concentrations below LORs. As specified by laboratory. Dynamic recovery limits as specified by laboratory. As specified by laboratory (generally dynamic recovery limits). Usually not performed and assessed based on LCS results. Dynamic recovery limits as specified by laboratory. Recovery 70 - 130% or dynamic limits if specified by laboratory. Analytic concentrations below LORs. | percent difference (RPD) results for field duplicate samples. ESDAT generated summary of field blank analytical results. ESDAT generated summary of field blank analytical results. Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports Laboratory reports | Yes Yes No Yes Yes Ves Ves | in batch 111796B. Total oxidised nitrogen, total kjeldahl nitrogen and total nitrogen (as N) were reported above LOR. Not seen as a significant impact to interpretation of results asno adopted assessment criteria for these analytes in groundwater. Mattix spike recovery not determined for manganese and nitrite and nitrate (as N) as the background level greater than or equal to 4x spike level. An ananymous sample was used from a different project and therefore this is not expected to | N/A N/A No Yes Yes Yes | Primary laboratory received sample The RPD reported for nickel (32%) passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of |



| | | Location Code | | MW3 | | MW3 | MW3 | | MW3 | MW3 | | MW3 | MW3 | 1 | MW3 | MW3 | | MW3 | MW3 | | MW3 | MW3 | | MW3 | MW3 | |
|--|------------------|------------------|----------------------|---------------------|--|-------------------|---------------------|----------|--|--|--|-------------------|---------------------|-----|--|--|--|----------------------|---------------------|----------|-------------------|---------------------|---------|-------------------|---------------------|--------------|
| | | Field ID Date | | QC101 08/02/2023 | - | MW3 08/02/2023 | QC201 08/02/2023 | 1 | MW3 14/08/2023 | QC102 14/08/2023 | - | MW3 14/08/2023 | QC202 14/08/2023 | 1 | MW3 09/02/2024 | QC103 09/02/2024 | - | MW3 09/02/2024 | QC203 09/02/2024 | | MW3 11/07/2024 | QC104 11/07/2024 | - | MW3 11/07/2024 | QC204 11/07/2024 | - |
| | | Sample Type | | Field_D | | Normal | Interlab_D | 1 | Normal | Field_D | 1 | Normal | Interlab_D | 1 | Normal | Field_D | 1 | Normal | Interlab_D | | Normal | Field_D | | Normal | Interlab_D | |
| | 1 | Lab Report No. | ES2304011 | ES2304011 | RPD | ES2304011 | 316159 | RPD | ES2327328 | ES2327328 | RPD | ES2327328 | 1020195 | RPD | ES2404239 | ES2404239 | RPD | ES2404239 | 1067666 | RPD | ES2423038 | ES2423038 | RPD | ES2423038 | 1117968 | RPD |
| | Unit | EQL | | | | | | | | | | | | | | | | | | | | | | | | |
| Physical Parameters | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Electrical Conductivity | μS/cm | 1 | 34,200 | - | | 34,200 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Total Dissolved Solids pH (Lab) | mg/L pH Units | 0.01 | 22,200 7.09 | - | - | 22,200 7.09 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Metals | prionito | 0.01 | 7.03 | | <u> </u> | 7.05 | | | | | | | | | 1 | | | | | - | | | - | | _ | |
| Arsenic (filtered) | mg/L | 0.001 | < 0.010 | < 0.010 | 0 | < 0.010 | 0.004 | 0 | < 0.010 | < 0.010 | 0 | < 0.010 | 0.002 | 0 | < 0.010 | < 0.010 | 0 | < 0.010 | 0.011 | 0 | 0.003 | 0.003 | 0 | 0.003 | <0.01 | 0 |
| Cadmium (filtered) Chromium (filtered) | mg/L | 0.0001 | <0.0010 | <0.0010 | 0 | <0.0010 | 0.0001 | 0 | <0.0010 | <0.0010 | 0 | <0.0010 | <0.0002 | 0 | <0.0010 | <0.0010 | 0 | <0.0010 | <0.0002 | 0 | <0.0001 | <0.0001 | 0 | <0.0001 | <0.002 | 0 |
| Copper (filtered) | mg/L mg/L | 0.001 0.001 | <0.010 <0.010 | <0.010 <0.010 | 0 | <0.010 | 0.002 <0.001 | 0 | <0.010 | <0.010 | 0 | <0.010 | 0.002 0.002 | 0 | <0.010 | <0.010 | 0 | <0.010 | 0.005 0.006 | 0 | <0.001 0.002 | <0.001 <0.001 | 0 67 | <0.001 0.002 | <0.01 | 0 |
| Iron (filtered) | mg/L | 0.01 | 5.05 | 5.15 | 2 | 5.05 | 5.7 | 12 | 5.64 | 6.04 | 7 | 5.64 | 2.3 | 84 | 8.01 | 8.08 | 1 | 8.01 | - | - | 7.01 | 6.96 | 1 | 7.01 | 7.4 | 5 |
| Lead (filtered) | mg/L | 0.001 | <0.010 | <0.010 | 0 | <0.010 | 0.001 | 0 | <0.010 | < 0.010 | 0 | <0.010 | 0.002 | 0 | <0.010 | < 0.010 | 0 | <0.010 | 0.005 | 0 | <0.001 | <0.001 | 0 | <0.001 | <0.01 | 0 |
| Manganese (filtered) Mercury (filtered) | mg/L mg/L | 0.001 0.00005 | 5.99 <0.0001 | 6.15 <0.0001 | 0 | 5.99 <0.0001 | 5.8 <0.00005 | 0 | 6.39 <0.0001 | 6.57 <0.0001 | 0 | 6.39 <0.0001 | 5.9 <0.0001 | 8 | 7.00 <0.0001 | 7.08 <0.0001 | 0 | 7.00 < 0.0001 | <0.0001 | - 0 | 6.79 <0.0001 | 6.80 <0.0001 | 0 | 6.79 <0.0001 | 7.4 <0.001 | 9 |
| Nickel (filtered) | mg/L | 0.000 | 0.191 | 0.167 | 13 | 0.191 | 0.18 | 6 | 0.207 | 0.205 | 1 | 0.207 | 0.18 | 14 | 0.191 | 0.197 | 3 | 0.191 | 0.16 | 4 | 0.200 | 0.200 | 0 | 0.200 | 0.20 | 0 |
| Zinc (filtered) | mg/L | 0.001 | 0.225 | 0.196 | 14 | 0.225 | 0.23 | 2 | 0.122 | 0.074 | 49 | 0.122 | 0.086 | 35 | 0.247 | 0.253 | 2 | 0.247 | 0.18 | 8 | 0.243 | 0.239 | 2 | 0.243 | 0.25 | 3 |
| Inorganics | / | 2.24 | | | | 2.00 | | | | | 1 | 2.00 | | | 0.00 | | ļ | 0.00 | | | 2.22 | | | 0.00 | | |
| Ammonia (as N) Nitrate (as N) | mg/L mg/L | 0.01 0.01 | 0.22 <0.01 | - | + : | 0.22 <0.01 | - | - | 0.29 <0.01 | - | + : | 0.29 <0.01 | <0.02 | - 0 | 0.29 | | | 0.29 | 0.10 | 108 | 0.28 0.01 | - | - | 0.28 | <0.02 | - 0 |
| Nitrite (as N) | mg/L | 0.01 | <0.01 | - | - | <0.01 | - | - | <0.01 | - | - | <0.01 | < 0.02 | 0 | <0.01 | - | - | <0.01 | <0.02 | 0 | <0.01 | - | - | <0.01 | <0.02 | 0 |
| Total Oxidised Nitrogen (as N) | mg/L | 0.01 | <0.01 | 0.02 | 67 | <0.01 | - | - | <0.01 | - | - | <0.01 | < 0.05 | 0 | 0.03 | 0.02 | 40 | 0.03 | 0.11 | 114 | 0.01 | 0.02 | 67 | 0.01 | < 0.05 | 0 |
| Total Kjeldahl Nitrogen Total Nitrogen (as N) | mg/L mg/L | 0.1 0.1 | 1.0 | 1.3 | 26 26 | 1.0 | 0.5 | - 67 | 0.4 | - | - | 0.4 | 0.5 0.5 | 22 | 0.9 | 0.9 | 0 | 0.9 | 1.3 1.4 | 36 43 | 0.6 | 0.6 0.6 | 0 | 0.6 | 1.0 | 50 50 |
| Phosphorus (as P) | mg/L | 0.01 | 0.12 | 0.10 | 18 | 0.12 | 0.8 | 148 | 0.02 | - | | 0.4 | - | - | 0.9 | 0.09 | 12 | 0.08 | 1.4 | - | 0.04 | 0.04 | 0 | 0.04 | - | - |
| Phosphate (as P) | mg/L | 0.01 | - | - | | - | - | - | - | - | - | - | 0.03 | - | - | - | - | | 0.05 | - | - | - | - | - | 0.03 | - |
| Ortho-phosphate (as P) Fluoride | mg/L | 0.01 | <0.01 | - | - | <0.01 | - | | - | - | - | - | - | - | - | - | - | | - | | - | - | | - | - | ↓ - □ |
| Sodium Absorption Ratio (filtered) | mg/L | 0.1 0.01 | 1.2 37.8 | - | - | 1.2 37.8 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | + - |
| Major lons | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Calcium (filtered) | mg/L | 1 | 181 | - | - | 181 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Chloride Magnesium (filtered) | mg/L mg/L | 1 | 11,900 1,040 | - | 1 - | 11,900 1,040 | - | - | - | - | 1 - | - | - | - | - | - | - | - | - | | - | - | - | - | - | 1 |
| Potassium (filtered) | mg/L | 1 | 1,040 | - | | 1,040 | - | - | | | | | - | 1 | | | 1 | | - | | - | - | - | | | + |
| Sulfate (as SO4) (filtered) | mg/L | 1 | 907 | - | - | 907 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Sodium (filtered) Anions Total | mg/L | 1 | 5,980 | - | - | 5,980 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Cations Total | meq/L meq/L | 0.01 0.01 | 359 355 | - | - | 359 355 | - | - | - | - | - | - | - | - | - | | - | - | - | - | - | - | - | | - | - |
| Ionic Balance | % | 0.01 | 0.55 | - | - | 0.55 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Alkalinity | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Bicarbonate Alkalinity (as CaCO3) Carbonate Alkalinity (as CaCO3) | mg/L mg/L | 1 1 | 222 <1 | - | - | 222 <1 | - | - | - | - | - | - | - | - | - | | - | - | - | - | - | - | - | - | - | - |
| Hydroxide Alkalinity (as CaCO3) | mg/L | 1 | <1 | - | - | <1 | - | - | - | - | - | - | - | - | - | - | | - | - | - | - | - | - | | - | + - |
| Total Alkalinity (as CaCO3) | mg/L | 1 | 222 | - | - | 222 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | | - | - |
| Hardness (as CaCO3) (filtered) | mg/L | 1 | 4,730 | - | - | 4,730 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| BTEX Benzene | μg/L | 1 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 | <1 | <1 | 0 |
| Toluene | μg/L | 1 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 |
| Ethylbenzene | μg/L | 1 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 | <2 | <2 | 0 | <2 | <1 | 0 |
| Xylene (m & p) Xylene (o) | μg/L μg/L | 2 | <2 <2 | <2 <2 | 0 | <2 <2 | <2 <1 | 0 | <2 <2 | <2 <2 | 0 | <2 <2 | <2 <1 | 0 | <2 <2 | <2 <2 | 0 | <2 <2 | <2 <1 | 0 | <2 <2 | <2 <2 | 0 | <2 <2 | <2 <1 | 0 |
| Total Xylene | μg/L | 2 | <2 | <2 | 0 | <2 | - | - | <2 | <2 | 0 | <2 | <3 | 0 | <2 | <2 | 0 | <2 | <3 | 0 | <2 | <2 | 0 | <2 | <3 | 0 |
| Total BTEX | μg/L | 1 | <1 | <1 | 0 | <1 | - | - | <1 | <1 | 0 | <1 | - | - | <1 | <1 | 0 | <1 | - | - | <1 | <1 | 0 | <1 | - | - |
| Total Petroleum Hydrocarbons C6-C9 Fraction | | 40 | 20 | 20 | | -00 | 40 | _ | 20 | 00 | | 20 | 20 | | 20 | 20 | | -00 | 20 | | 00 | 00 | 0 | 00 | -00 | |
| C10-C14 Fraction | μg/L μg/L | 10 50 | <20 <50 | <20 <50 | 0 | <20 <50 | <10 <50 | 0 | <20 <50 | <20 <50 | 0 | <20 <50 | <20 <50 | 0 | <20 <50 | <20 <50 | 0 | <20 <50 | <20 <50 | 0 | <20 <50 | <20 <50 | 0 | <20 <50 | <20 90 | 0 57 |
| C15-C28 Fraction | μg/L | 100 | <100 | <100 | 0 | <100 | 140 | 33 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | 810 | 800 | 1 | 810 | 400 | 68 |
| C29-C36 Fraction | μg/L | 50 | <50 | <50 | 0 | <50 | <100 | 0 | <50 | <50 | 0 | <50 | <100 | 0 | <50 | <50 | 0 | <50 | <100 | 0 | <50 | <50 | 0 | <50 | 300 | 143 |
| C10-C36 Fraction (Sum) Total Recoverable Hydrocarbons | μg/L | 50 | <50 | <50 | 0 | <50 | 140 | 95 | <50 | <50 | 0 | <50 | <100 | 0 | <50 | <50 | 0 | <50 | <100 | 0 | 810 | 800 | 1 | 810 | 790 | 2 |
| C6-C10 Fraction | μg/L | 10 | <20 | <20 | 0 | <20 | <10 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 |
| C6-C10 Fraction minus BTEX (F1) | μg/L | 10 | <20 | <20 | 0 | <20 | <10 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 | <20 | <20 | 0 |
| >C10-C16 Fraction >C10-C16 Fraction minus naphthalene (F2) | μg/L μg/L | 50 50 | <100 <100 | <100 <100 | 0 | <100 <100 | 130 130 | 26 26 | <100 <100 | <100 <100 | 0 | <100 <100 | <50 <50 | 0 | <100 <100 | <100 <100 | 0 | <100 <100 | <50 <50 | 0 | <100 <100 | <100 <100 | 0 | <100 <100 | 110 110 | 10 10 |
| >C16-C34 Fraction | μg/L | 100 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | 400 | 480 | 18 | 400 | 600 | 40 |
| >C34-C40 Fraction | μg/L | 100 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | 200 | 67 |
| >C10-C40 Fraction (Sum) PAHs | μg/L | 50 | <100 | <100 | 0 | <100 | 130 | 26 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | <100 | <100 | 0 | 400 | 480 | 18 | 400 | 910 | 78 |
| Acenaphthene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Acenaphthylene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Anthracene Benz(a)anthracene | μg/L μg/L | 1 1 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 |
| Benzo(a)pyrene | μg/L μg/L | 0.5 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 |
| Benzo(b+j)fluoranthene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | - | - | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Benzo(g,h,i)perylene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Benzo(b+j+k)fluoranthene Benzo(k)fluoranthene | μg/L μg/L | 2 | <1.0 | <1.0 | - 0 | <1.0 | <2 | - | <1.0 | <1.0 | - 0 | <1.0 | <1 | - 0 | <1.0 | <1.0 | - 0 | <1.0 | - <1 | - 0 | <1.0 | <1.0 | - 0 | <1.0 | - <1 | - 0 |
| Chrysene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Dibenz(a,h)anthracene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Fluorene Fluorene | μg/L μg/L | 1 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 |
| Indeno(1,2,3-c,d)pyrene | μg/L μg/L | 1 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 | <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 <1.0 | <1 | 0 | <1.0 <1.0 | <1.0 <1.0 | 0 | <1.0 | <1 <1 | 0 |
| Naphthalene | μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Naphthalene (VOC) | μg/L | 5 | - | | - | - | - | - | | | - | - | - | - | - | - | - | - | - | | <5 | <5 | 0 | <5 | <10 | 0 |
| Phenanthrene Pyrene | μg/L μg/L | 1 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 <1 | 0 | <1.0 | <1.0 | 0 | <1.0 <1.0 | <1 <1 | 0 | <1.0 | <1.0 | 0 | <1.0 | <1 | 0 |
| Benzo(a)pyrene TEQ (Zero) | μg/L μg/L | 0.5 | <1.0 <0.5 | <1.0 <0.5 | 0 | <1.0 <0.5 | <1 | - | <1.0 <0.5 | <1.0 <0.5 | 0 | <1.0 <0.5 | <1 | - | <1.0 <0.5 | <1.0 <0.5 | 0 | <1.0 <0.5 | <1 | - | <1.0 <0.5 | <1.0 <0.5 | 0 | <1.0 <0.5 | <1 | - |
| Sum of Polycyclic aromatic hydrocarbons (PAH) |) μg/L | 0.5 | <0.5 | <0.5 | 0 | <0.5 | - | - | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 | <0.5 | <0.5 | 0 | <0.5 | <1 | 0 |
| Benzo(a)pyrene TEQ | μg/L | 5 | - | - | - | - | <5 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Total Positive PAHs Phenols | μg/L | 1 | - | - | - | - | 0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | + |
| 2-Methylphenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - 1 | - | - | - | - | - | - |
| | | | | | | | | | | | | | | | | | | | | | | | | | | |

Project: Surface Water and Groundwater Monitoring 2023 and 2024 Client: ReDirect Recycling Site Address: 24 Davis Road, Wetherill Park NSW

Table D2: RPD Analytical Results

| | J |
|--------|----|
| s⊘nv⊘r | sa |

| | | Location Code | MW3 | MW3 | |
|--------------------------------|------|----------------|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|
| | | Field ID | MW3 | QC101 | 1 | MW3 | QC201 | 1 1 | MW3 | QC102 | 1 | MW3 | QC202 | 1 | MW3 | QC103 | | MW3 | QC203 | 1 | MW3 | QC104 | 1 | MW3 | QC204 | 1 |
| | | Date | 08/02/2023 | 08/02/2023 | 1 | 08/02/2023 | 08/02/2023 | 1 1 | 14/08/2023 | 14/08/2023 | 1 | 14/08/2023 | 14/08/2023 | 1 | 09/02/2024 | 09/02/2024 | | 09/02/2024 | 09/02/2024 | 1 | 11/07/2024 | 11/07/2024 | 1 | 11/07/2024 | 11/07/2024 | 1 |
| | | Sample Type | Normal | Field_D | 1 | Normal | Interlab_D | 1 [| Normal | Field_D | 1 | Normal | Interlab_D | 1 | Normal | Field_D | | Normal | Interlab_D | 1 | Normal | Field_D | 1 | Normal | Interlab_D | 1 |
| | | Lab Report No. | ES2304011 | ES2304011 | RPD | ES2304011 | 316159 | RPD | ES2327328 | ES2327328 | RPD | ES2327328 | 1020195 | RPD | ES2404239 | ES2404239 | RPD | ES2404239 | 1067666 | RPD | ES2423038 | ES2423038 | RPD | ES2423038 | 1117968 | RPI |
| | Unit | EQL | | | | | | | | | | | | | | | | | | | | | | | | |
| 2-Nitrophenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | T |
| 2,4-Dimethylphenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - 1 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 3-&4-Methylphenol (m&p-cresol) | μg/L | 2 | <2.0 | - | - | <2.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 4-Chloro-3-methylphenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - 1 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Phenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | _ |
| alogenated Phenols | | | | | 1 | | | | | | | | | | | | | | | | | | | | 1 | 1 |
| 2,4,5-Trichlorophenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 2,4,6-Trichlorophenol | μg/L | 1 | <1.0 | - | ٠. | <1.0 | - | - 1 | - | | - | - | - | - | - | - | - | - | - | - | - | - | - | | - | 1 |
| 2,4-Dichlorophenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | T - |
| 2,6-Dichlorophenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 2-Chlorophenol | μg/L | 1 | <1.0 | - | - | <1.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| Pentachlorophenol | μg/L | 2 | <2.0 | - | - | <2.0 | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | 1 - |

^{*}RPDs have only been considered where a concentration is greater than 1 times the EQL.

**Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 1000 (1 - 10 x EQL); 50 (10 - 20 x EQL); 30 (> 20 x EQL))

***Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

Site Address: 24 Davis Road, Wetherill Park NSW

Table D3: Rinsate Analytical Table



Field ID QC301 QC302 QC303 QC304 Date 08/02/2023 09/02/2024 11/07/2024 14/08/2023 Sample Type Rinsate Rinsate Rinsate Rinsate Lab Report No. ES2304011 ES2327328 ES2404239 ES2423038 Unit FOI Arsenic (filtered) mg/L 0.001 Cadmium (filtered) mg/L 0.0001 < 0.0001 Chromium (filtered) ma/L 0.001 Copper (filtered) mg/L 0.001 Iron (filtered) mg/L 0.05 Lead (filtered) mg/L 0.001 Manganese (filtered) mg/L 0.001 0.038 Mercury (filtered) mg/L 0.0001 Nickel (filtered) mg/L 0.001 mg/L 0.005 norganics
Total Oxidised Nitrogen (as N) mg/L 0.01 Total Kjeldahl Nitrogen mg/L 0.1 0.1 Total Nitrogen (as N) mg/L 3.4 0.1 Phosphorus (as P) mg/L 0.02 BTEX Benzene μq/L μg/L Ethylbenzene μg/L Xylene (m & p) μg/L Xvlene (o) μg/L Total Xylene μg/L μg/L 1 otal Petroleum Hydrocarbons μg/L C6-C9 Fraction 20 C10-C14 Fraction μg/L 50 C15-C28 Fraction μg/L 100 C29-C36 Fraction μg/L 50 C10-C36 Fraction (Sum) μg/L 50 otal Recoverable Hydrocarbons C6-C10 Fraction μg/L 20 μg/L 20 >C10-C16 Fraction μg/L 100 >C10-C16 Fraction minus naphthalene (F2) μg/L 100 >C16-C34 Fraction μg/L 100 >C34-C40 Fraction >C10-C40 Fraction (Sum) μg/L 100 μg/L 100 <100 <100 PAHs Acenaphthene ua/L Acenaphthylene μg/L Anthracene μg/L <1.0 Benz(a)anthracene μg/L Benzo(a)pyrene μg/L 0.5 Benzo(b+j)fluoranthene μg/L Benzo(g,h,i)perylene μg/L Benzo(k)fluoranthene μg/L Chrysene μg/L 1 Dibenz(a,h)anthracene μg/L Fluoranthene μg/L Fluorene μg/L Indeno(1,2,3-c,d)pyrene μg/L Naphthalene μg/L Naphthalene (VOC) μg/L 5 Phenanthrene μg/L μg/L Benzo(a)pyrene TEQ (Zero) μg/L 0.5 < 0.5 < 0.5 < 0.5 Sum of Polycyclic aromatic hydrocarbons (PAH) μg/L 0.5



| | | Field ID Date le Type port No. | 6/02/2023 Trip Spike | TSC | % Recovery | QC402 1/08/2023 Trip Spike ES2327328 | TSC | % Recovery | QC503 5/02/2024 Trip Spike ES2403942 | TSC | % Recovery | QC504 5/02/2024 Trip Spike ES2404239 | TSC | % Recovery | QC505 8/07/2024 Trip Spike ES2423038 | TSC | % Recovery | QC505 9/07/2024 Trip Spike ES2422553 | TSC | % Recovery |
|----------------|------|---|-------------------------|-----|------------|---|-----|------------|---|-----|------------|---|-----|------------|---|-----|------------|---|-----|------------|
| | Unit | EQL | | | | | | | | | | | | | | | | | | |
| BTEXN | | | | | | | | | | | | | | | | | | | | |
| Benzene | μg/L | 1 | 16 | 20 | 80 | 16 | 20 | 80 | 20 | 20 | 100 | 15 | 20 | 75 | 14 | 20 | 70 | 14 | 20 | 70 |
| Toluene | μg/L | 2 | 15 | 20 | 75 | 16 | 20 | 80 | 18 | 20 | 90 | 16 | 20 | 80 | 15 | 20 | 75 | 15 | 20 | 75 |
| Ethylbenzene | μg/L | 2 | 14 | 20 | 70 | 17 | 20 | 85 | 17 | 20 | 85 | 16 | 20 | 80 | 14 | 20 | 70 | 16 | 20 | 80 |
| Xylene (m & p) | μg/L | 2 | 15 | 20 | 75 | 18 | 20 | 90 | 16 | 20 | 80 | 17 | 20 | 85 | 14 | 20 | 70 | 16 | 20 | 80 |
| Xylene (o) | μg/L | 2 | 15 | 20 | 75 | 19 | 20 | 95 | 16 | 20 | 80 | 17 | 20 | 85 | 16 | 20 | 80 | 14 | 20 | 70 |
| Total Xylene | μg/L | 2 | 30 | 40 | 75 | 37 | 40 | 93 | 32 | 40 | 80 | 34 | 40 | 85 | 30 | 40 | 75 | 30 | 40 | 75 |
| Total BTEX | μg/L | 1 | 75 | 100 | 75 | 86 | 100 | 86 | 87 | 100 | 87 | 81 | 100 | 81 | 73 | 100 | 73 | 75 | 100 | 75 |
| Naphthalene | μg/L | 1 | 15 | 20 | 75 | 17 | 20 | 85 | 16 | 20 | 80 | 18 | 20 | 90 | 19 | 20 | 95 | 21 | 20 | 105 |

Appendix E: Laboratory Reports



Chain of Custody Documentation

| Senversa Pt | v Ltd | | | Laboratory: | ALS NSW | | | | | | | | | Analysis R | equired | | |
|--------------|----------------------------|----------------------|-------------------|--------------------------------|--------------------------------------|---------------------|-------------|------------------------------|-------------------|-------------------------------|----------------|--------------|--------------|------------|---------|-------|---|
| ABN 89 132 | a.com.au | | | Address: Contact: Phone: | Sample Receipt | | | | , | | | | | | | | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. |
| Job Number | : | S2 | 0102 | Purchase Order: | | | 1 | TALS | METALS | AND | | | | | | | Environmental Divisi |
| Project Nam | e: | Wetherill | Park WME | Quote No: | EN/103/21 | | 1 | 3 ME | 3 ME | NS A | ja . | | | | | | Sydney Work Order Reference ES230401 |
| Sampled By | | Bec | Chapple | Turn Around Time: | Standard 7 D | Days | | AH/ | /PAH/8 | ANIONS | | | | (NW | | | Work Order Reference |
| Project Man | | | a Walsh | Page: | 1 | of 1 | EXN | EXF | Ĭ. | NS, | | | | AND | ~ | | ES230401 |
| Email Repor | | Bec.Chapple@ | senversa.com.au; | Phone/Mobile: | 0408038593, 040 | | (TRH/BTEXN) | W-26 (TRH/BTEX/PAH/8 METALS) | RH/BT LS) | NT-14 (CATIONS, NUTRIENTS) | NT-11 (TN, TP) | EA015H (TDS) | EA025H (TSS) | (FE A | | | |
| Етпан Керог | 110. | Sample Information | | Priorie/Mobile. | Container Infor | | 1 E | 3 (TF | E NON | 4 (C | T) . | 15H | 25H | | | | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | W-18 | N-2(| W-27 (T PHENOI | 155 | 1 | EA0 | EA0 | EG005F | | НОГР | |
| 1 | QC401 | W | 8/02/2023 | AM | VOA | 1 | Х | | | | - | | | | | + | |
| 2 | QC501 | W | 8/02/2023 | AM | VOA | 1 | X | | | | | | | | | 1 | - - IIII III III III III III III III II |
| 3 | QC301 | w | 8/02/2023 | AM | VS x2, N, UA, VSA | 5 | | Х | | | X | | | X | | 1 | Telephone: +61-2-8784 8555 |
| 4 | MW1 | W | 8/02/2023 | AM | P, VS x2, N, UA, VSA | 6 | | | X | X | | | | X | | | _ |
| 5 | MW2 | W | 8/02/2023 | AM | P, VS x2, N, UA, VSA | 6 | | | × | X | | | | X | | | |
| 6 | MW3 | W | 8/02/2023 | AM | P, VS x2, N, UA, VSA | 6 | | | × | X | | | | X | | | |
| 7 | MW4 | W | 8/02/2023 | AM ` | P, VS x2, N, UA, VSA | 6 | | | X | X | | | | Х | | | |
| 8 | MW6 | W | 8/02/2023 | AM | P, VS x2, N, UA, VSA | 6 | | | X | X | | | | Х | | | |
| 9 | QC101 | W | 8/02/2023 | AM | VS x2, N, UA, VSA | 5 | | X | | | Х | | | X | | | |
| X | QC201 | W | 8/02/2023 | AM | VS x2, N, UA, VSA | 5 | | | | | | | | | | | Please forward to Envirolab |
| | | | | | | | | | | | - | | | | | | |
| | | | | | | - | | - | - | | | | | - | | | |
| | | | | - | | - | _ | - | - | - | | | | | 7 | | |
| | | | | | | - | - | - | - | | | | | | | + | |
| - | | - | | - | | 1 | - | - | - | - | _ | | - | - | | | |
| | | | | | - | - | | - | - | - | | | - | - | | | |
| | | | | | | - | - | - | - | - | | | | - | _ | - | |
| Total | | | | | | 47 | | | | | | | | | | | |
| Total | test that proper field sam | unling propedures in | negordanae with S | nuveres standard pro- | and was and far project | 47 Sampler Name: | | Pag (| Chapple | | Signate | | 4 | 2111 | | Date: | 8/02/2023 |
| | is were used during the c | | | enversa standard proc | secures and/or project | Sampler Name. | | Dec c | ларріе | | Signati | | 区区 | 巻り | | Date | 6/02/2023 |
| Relinquishe | d By: | | | | Method of Shipment (if ap | plicable): | | | Receiv | red by: | | ~ | _ | | | | |
| Name/Signat | ure: | Bec Chapple | | Date: 8/2/23 | Carrier / Reference #: | | | | Name/ | Signature | e: | TA | ンシ | fg | | | Date: 812123 |
| Of: | | | | Time: 12:00 PM | Date/Time: | | | - | Of: | · · | | - | 4 | /_ | | | Time: 12-29 |
| Name/Signati | ure: | | | Date: Time: | Carrier / Reference #: Date/Time: | | | | Name/ | Signature | 9: | | | | | | Date: |
| Name/Signat | Ite. | | | Date: | Carrier / Reference #: | | | | Name/ | Signature | 2· | | | | | | Date: |
| o. | | | | Time: | Date/Time: | | | | r territer | -igriatule | ,, | | | | | | Time: |



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2304011

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Helen Simpson

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : helen.simpson@alsqlobal.com

Telephone : 02 8252 0000 Telephone : +61 2 8784 8555 Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 3

 Order number
 : --- Quote number
 : ES2022SENVER0004 (SY/103/22)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Site : ----

Sampler : Bec Chapple

Dates

Date Samples Received : 08-Feb-2023 12:20 Issue Date : 08-Feb-2023 Client Requested Due : 15-Feb-2023 Scheduled Reporting Date : 15-Feb-2023

Date

Delivery Details

Mode of Delivery : Client Drop Off Security Seal : Intact.

No. of coolers/boxes : 2 Temperature : 15.56'C - Ice present

Receipt Detail : No. of samples received / analysed : 9 / 9

General Comments

- This report contains the following information:
 - Sample Container(s)/Preservation Non-Compliances
 - Summary of Sample(s) and Requested Analysis
 - Proactive Holding Time Report
 - Requested Deliverables
- Sample ID QC201 will be forwarded to Envirolab as per COC.
- samples 1 and 2 had a sample date of 08/02 on the COC but the bottle had 06/02.
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Sample(s) requiring volatile organic compound analysis received in airtight containers (ZHE).
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

Issue Date : 08-Feb-2023

Page

2 of 3 ES2304011 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

| process necessatasks. Packages as the determining tasks, that are included in the sampling default 00:00 on its provided, the laboratory and component Matrix: WATER Laboratory sample | ry for the executi may contain ad ation of moisture uded in the package. time is provided, the date of sampling sampling date wi displayed in bra | be part of a laboratory on of client requested ditional analyses, such content and preparation the sampling time will g. If no sampling date II be assumed by the ckets without a time | WATER - EG005F Dissolved Metals by ICPAES | WATER - EP080 BTEXN | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - NT-14 Extended Water Suite B | WATER - W-18 TRH(C6 - C9)/BTEXN | WATER - W-26 TRH/BTEXN/PAH/8 Metals | WATER - W-27 TRH/BTEXN/PAH/Phenols/8 Metals |
|--|---|--|--|------------------------|--|---|------------------------------------|--|--|
| ID ES2304011-001 | <i>time</i> 06-Feb-2023 00:00 | QC401 | ≥ □ | <u> </u> | ≶ ⊢ | <u> ≤ iii</u> | ≤ F | ≶ F | SF |
| ES2304011-001 | 06-Feb-2023 00:00 | QC501 | - | √ | | | _ | | |
| | | | | Ľ | | | | | |
| ES2304011-003 | 08-Feb-2023 00:00 | QC301 | ✓ | | ✓ | | | ✓ | |
| ES2304011-004 | 08-Feb-2023 00:00 | MW1 | ✓ | | | ✓ | | | ✓ |
| ES2304011-005 | 08-Feb-2023 00:00 | MW2 | ✓ | | | ✓ | | | ✓ |
| ES2304011-006 | 08-Feb-2023 00:00 | MW3 | ✓ | | | 1 | | | ✓ |
| ES2304011-007 | 08-Feb-2023 00:00 | MW4 | ✓ | | | ✓ | | | ✓ |
| ES2304011-008 | 08-Feb-2023 00:00 | MW6 | 1 | | | ✓ | | | ✓ |
| ES2304011-009 | 08-Feb-2023 00:00 | QC101 | ✓ | | ✓ | | | ✓ | |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

: 08-Feb-2023 Issue Date

Page

3 of 3 ES2304011 Amendment 0 Work Order Client : SENVERSA PTY LTD



Requested Deliverables

| Angus | Dibl | ey |
|-------|------|----|
|-------|------|----|

| - *AU Certificate of Analysis - NATA (COA) | Email | angus.dibley@senversa.com.au |
|--|-------|------------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | angus.dibley@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | angus.dibley@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | angus.dibley@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | angus.dibley@senversa.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | angus.dibley@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | angus.dibley@senversa.com.au |

BEC CHAPPLE

| - *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
|--|-------|-----------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| - EDI Format - XTab (XTAB) | Email | bec.chapple@senversa.com.au |

EMMA WALSH

| - *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
|--|-------|----------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - XTab (XTAB) | Email | Emma.Walsh@senversa.com.au |

GRAEME MILLER

| OTALINE INICELIT | | |
|--|-------|-------------------------------|
| - *AU Certificate of Analysis - NATA (COA) | Email | graeme.miller@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | graeme.miller@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | graeme.miller@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | graeme.miller@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | graeme.miller@senversa.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | graeme.miller@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | graeme.miller@senversa.com.au |
| | | |

| , | | g |
|--|-------|-----------------------|
| lan Wilson | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | ian.wilson@pkc.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | ian.wilson@pkc.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | ian.wilson@pkc.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | ian.wilson@pkc.com.au |
| - Chain of Custody (CoC) (COC) | Email | ian.wilson@pkc.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | ian.wilson@pkc.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | ian.wilson@pkc.com.au |
| | | |

JIM BAILEY

| - *AU Certificate of Analysis - NATA (COA) | Email | jim.bailey@pkc.com.au |
|--|-------|-----------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | jim.bailey@pkc.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | jim.bailey@pkc.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | jim.bailey@pkc.com.au |
| - Chain of Custody (CoC) (COC) | Email | jim.bailey@pkc.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | jim.bailey@pkc.com.au |
| - EDI Format - XTab (XTAB) | Email | jim.bailey@pkc.com.au |

JONATHAN MANN

| OUTATION MAIN | | |
|--|-------|-------------------------------|
| - *AU Certificate of Analysis - NATA (COA) | Email | jonathan.mann@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | jonathan.mann@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | jonathan.mann@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | jonathan.mann@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | jonathan.mann@senversa.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | jonathan.mann@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | jonathan.mann@senversa.com.au |
| | | |

SUPPLIER ACCOUNTS

| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
|-----------------------------|-------|---------------------------------|
|-----------------------------|-------|---------------------------------|



CERTIFICATE OF ANALYSIS

Work Order : ES2304011

: SENVERSA PTY LTD

Contact : EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number : ----

Client

C-O-C number : ----

Sampler : Bec Chapple

Site : --

Quote number : SY/103/22

No. of samples received : 9
No. of samples analysed : 9

Page : 1 of 11

Laboratory : Environmental Division Sydney

Contact : Helen Simpson

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61 2 8784 8555

Date Samples Received : 08-Feb-2023 12:20

Date Analysis Commenced : 08-Feb-2023

Issue Date : 14-Feb-2023 17:19



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Edwandy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EK067G: LOR raised for Total P on sample 4 due to sample matrix.
- EK059G: LOR raised for NOx on sample 4 due to sample matrix.
- EK058G: LOR raised for Nitrate on sample 4 due to sample matrix.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a.h)anthracene (1.0), Benzo(g.h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- As per QWI EN55-3 Data Interpreting Procedures, Ionic balances are typically calculated using Major Anions Chloride, Alkalinity and Sulfate; and Major Cations Calcium, Magnesium, Potassium and Sodium. Where applicable and dependent upon sample matrix, the Ionic Balance may also include the additional contribution of Ammonia, Dissolved Metals by ICPMS and H+ to the Cations and Nitrate, SiO2 and Fluoride to the Anions
- EG020: LORs have been raised for some samples due to matrix interference (High sample salinity)
- EK057G: LOR raised for Nitrite due to sample matrix
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.
- EA016: Calculated TDS is determined from Electrical conductivity using a conversion factor of 0.65.
- Sodium Adsorption Ratio (where reported): Where results for Na, Ca or Mg are <LOR, a concentration at half the reported LOR is incorporated into the SAR calculation. This represents a conservative approach for Na relative to the assumption that <LOR = zero concentration and a conservative approach for Ca & Mg relative to the assumption that <LOR is equivalent to the LOR concentration.

Page : 3 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC401 | QC501 | QC301 | MW1 | MW2 |
|---------------------------------------|------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 06-Feb-2023 00:00 | 06-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2304011-001 | ES2304011-002 | ES2304011-003 | ES2304011-004 | ES2304011-005 |
| | | | | Result | Result | Result | Result | Result |
| EA005P: pH by PC Titrator | | | | | | | | |
| pH Value | | 0.01 | pH Unit | | | | 7.74 | 7.70 |
| EA006: Sodium Adsorption Ratio (SA | R) | | | | | | | |
| ^ Sodium Adsorption Ratio | | 0.01 | - | | | | 30.4 | 31.6 |
| EA010P: Conductivity by PC Titrator | | | | | | | | |
| Electrical Conductivity @ 25°C | | 1 | μS/cm | | | | 25800 | 25700 |
| EA016: Calculated TDS (from Electric | al Conductivity) | | | | | | | |
| Total Dissolved Solids (Calc.) | | 1 | mg/L | | | | 16800 | 16700 |
| EA065: Total Hardness as CaCO3 | | | | | | | | |
| Total Hardness as CaCO3 | | 1 | mg/L | | | | 4020 | 3980 |
| ED037P: Alkalinity by PC Titrator | | | | | | | | |
| Hydroxide Alkalinity as CaCO3 | DMO-210-001 | 1 | mg/L | | | | <1 | <1 |
| Carbonate Alkalinity as CaCO3 | 3812-32-6 | 1 | mg/L | | | | <1 | <1 |
| Bicarbonate Alkalinity as CaCO3 | 71-52-3 | 1 | mg/L | | | | 916 | 815 |
| Total Alkalinity as CaCO3 | | 1 | mg/L | | | | 916 | 815 |
| ED041G: Sulfate (Turbidimetric) as SC | 04 2- by DA | | | | | | | |
| Sulfate as SO4 - Turbidimetric | 14808-79-8 | 1 | mg/L | | | | 691 | 756 |
| ED045G: Chloride by Discrete Analyse | | | | | | | | |
| Chloride | 16887-00-6 | 1 | mg/L | | | | 8840 | 8800 |
| ED093F: Dissolved Major Cations | | | | | | | | |
| Calcium | 7440-70-2 | 1 | mg/L | | | | 273 | 232 |
| Magnesium | 7439-95-4 | 1 | mg/L | | | | 810 | 826 |
| Sodium | 7440-23-5 | 1 | mg/L | | | | 4430 | 4590 |
| Potassium | 7440-09-7 | 1 | mg/L | | | | 25 | 21 |
| EG005(ED093)F: Dissolved Metals by | ICP-AES | | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | | | <0.05 | 4.97 | 0.40 |
| Manganese | 7439-96-5 | 0.01 | mg/L | | | <0.01 | 0.92 | 0.96 |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | | | <0.001 | 0.011 | 0.004 |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | | | <0.0001 | <0.0001 | <0.0001 |
| Chromium | 7440-47-3 | 0.001 | mg/L | | | <0.001 | <0.001 | <0.001 |
| Copper | 7440-50-8 | 0.001 | mg/L | | | <0.001 | 0.015 | 0.011 |
| Lead | 7439-92-1 | 0.001 | mg/L | | | <0.001 | <0.001 | <0.001 |
| Nickel | 7440-02-0 | 0.001 | mg/L | | | <0.001 | 0.023 | 0.006 |

Page : 4 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC401 | QC501 | QC301 | MW1 | MW2 |
|---|--------------------------|--------|-----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ing date / time | 06-Feb-2023 00:00 | 06-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2304011-001 | ES2304011-002 | ES2304011-003 | ES2304011-004 | ES2304011-005 |
| · · | | | | Result | Result | Result | Result | Result |
| EG020F: Dissolved Metals by ICF | P-MS - Continued | | | | | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | | | <0.005 | 0.012 | 0.008 |
| EG035F: Dissolved Mercury by F | IMS | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | | | <0.0001 | <0.0001 | <0.0001 |
| EK040P: Fluoride by PC Titrator | | | | | | | | |
| Fluoride | 16984-48-8 | 0.1 | mg/L | | | | 0.8 | 0.7 |
| EK055G: Ammonia as N by Discr | | | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | | | | 0.71 | 0.52 |
| EK057G: Nitrite as N by Discrete | | | J. J. | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | | | | <0.10 | <0.01 |
| | | 0.0. | g | | | | 0.10 | 0.01 |
| EK058G: Nitrate as N by Discrete Nitrate as N | 14797-55-8 | 0.01 | mg/L | | | | <0.10 | 0.03 |
| | | | Hig/L | | | | ~0.10 | 0.03 |
| EK059G: Nitrite plus Nitrate as N Nitrite + Nitrate as N | (NOx) by Discrete Anal | | ma/l | | | <0.01 | <0.10 | 0.03 |
| | | 0.01 | mg/L | | | <0.01 | <0.10 | 0.03 |
| EK061G: Total Kjeldahl Nitrogen | By Discrete Analyser | | | | | 2.4 | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | | | <0.1 | 0.9 | 1.0 |
| EK062G: Total Nitrogen as N (TK | N + NOx) by Discrete An | | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | | | <0.1 | 0.9 | 1.0 |
| EK067G: Total Phosphorus as P | by Discrete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | | | <0.01 | <0.05 | 0.06 |
| EK071G: Reactive Phosphorus as | s P by discrete analyser | | | | | | | |
| Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | | | | <0.01 | 0.02 |
| EN055: Ionic Balance | | | | | | | | |
| ø Total Anions | | 0.01 | meq/L | | | | 282 | 280 |
| ø Total Cations | | 0.01 | meq/L | | | | 274 | 280 |
| ø Ionic Balance | | 0.01 | % | | | | 1.52 | 0.09 |
| EP075(SIM)A: Phenolic Compour | nds | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | | | | <2.0 | <2.0 |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | | | | <1.0 | <1.0 |

Page : 5 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC401 | QC501 | QC301 | MW1 | MW2 |
|---------------------------------------|----------------------|-----------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 06-Feb-2023 00:00 | 06-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2304011-001 | ES2304011-002 | ES2304011-003 | ES2304011-004 | ES2304011-005 |
| | | | | Result | Result | Result | Result | Result |
| EP075(SIM)A: Phenolic Compound | ds - Continued | | | | | | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | | | | <1.0 | <1.0 |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | | | | <2.0 | <2.0 |
| EP075(SIM)B: Polynuclear Aromat | ic Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Fluorene | 86-73-7 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Anthracene | 120-12-7 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Pyrene | 129-00-0 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Chrysene | 218-01-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| ^ Sum of polycyclic aromatic hydrocar | rbons | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| EP080/071: Total Petroleum Hydro | carbons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | | <20 | <20 | <20 |
| C10 - C14 Fraction | | 50 | μg/L | | | <50 | <50 | <50 |
| C15 - C28 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| C29 - C36 Fraction | | 50 | μg/L | | | <50 | <50 | <50 |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | | | <50 | <50 | <50 |
| EP080/071: Total Recoverable Hyd | rocarbons - NEPM 201 | 3 Fractio | ns | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | | <20 | <20 | <20 |
| ^ C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | | <20 | <20 | <20 |

Page : 6 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC401 | QC501 | QC301 | MW1 | MW2 |
|---|-------------------|-----------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 06-Feb-2023 00:00 | 06-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2304011-001 | ES2304011-002 | ES2304011-003 | ES2304011-004 | ES2304011-005 |
| | | | | Result | Result | Result | Result | Result |
| EP080/071: Total Recoverable Hydroca | arbons - NEPM 201 | 3 Fractio | ns - Continued | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| >C16 - C34 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| >C34 - C40 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | | | <100 | <100 | <100 |
| ^ >C10 - C16 Fraction minus Naphthalene (F2) | | 100 | μg/L | | | <100 | <100 | <100 |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | 16 | <1 | <1 | <1 |
| Toluene | 108-88-3 | 2 | μg/L | <2 | 15 | <2 | <2 | <2 |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 14 | <2 | <2 | <2 |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | 15 | <2 | <2 | <2 |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 15 | <2 | <2 | <2 |
| ^ Total Xylenes | | 2 | μg/L | <2 | 30 | <2 | <2 | <2 |
| ^ Sum of BTEX | | 1 | μg/L | <1 | 75 | <1 | <1 | <1 |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | 17 | <5 | <5 | <5 |
| EP075(SIM)S: Phenolic Compound Su | rrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | | | 28.1 | 31.6 | 30.2 |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | | | 58.8 | 62.4 | 60.6 |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | | | 60.4 | 70.1 | 71.9 |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | | | 69.7 | 68.8 | 73.0 |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | | | 94.2 | 98.3 | 93.6 |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | | | 80.2 | 85.6 | 80.2 |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 99.7 | 97.1 | 89.9 | 96.8 | 100 |
| Toluene-D8 | 2037-26-5 | 2 | % | 99.4 | 98.2 | 94.5 | 99.5 | 98.4 |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 97.7 | 94.7 | 88.7 | 96.6 | 96.5 |

Page : 7 of 11
Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW4 | MW6 | QC101 | |
|--|-----------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|---|
| | | Sampli | ng date / time | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2304011-006 | ES2304011-007 | ES2304011-008 | ES2304011-009 | |
| | | | | Result | Result | Result | Result | |
| EA005P: pH by PC Titrator | | | | | | | | |
| pH Value | | 0.01 | pH Unit | 7.09 | 7.72 | 8.06 | | |
| EA006: Sodium Adsorption Ratio (SAR | 2) | | | | | | | |
| ^ Sodium Adsorption Ratio | | 0.01 | - | 37.8 | 21.4 | 6.70 | | |
| EA010P: Conductivity by PC Titrator | | | | | | | | |
| Electrical Conductivity @ 25°C | | 1 | μS/cm | 34200 | 19900 | 2310 | | |
| EA016: Calculated TDS (from Electrical | L Conductivity) | | | | | | | |
| Total Dissolved Solids (Calc.) | | 1 | mg/L | 22200 | 12900 | 1500 | | |
| EA065: Total Hardness as CaCO3 | | | | | | | | |
| Total Hardness as CaCO3 | | 1 | mg/L | 4730 | 3980 | 586 | | |
| ED037P: Alkalinity by PC Titrator | | | _ | | | | | 1 |
| Hydroxide Alkalinity as CaCO3 | DMO-210-001 | 1 | mg/L | <1 | <1 | <1 | | |
| Carbonate Alkalinity as CaCO3 | 3812-32-6 | 1 | mg/L | <1 | <1 | <1 | | |
| Bicarbonate Alkalinity as CaCO3 | 71-52-3 | 1 | mg/L | 222 | 1110 | 834 | | |
| Total Alkalinity as CaCO3 | | 1 | mg/L | 222 | 1110 | 834 | | |
| ED041G: Sulfate (Turbidimetric) as SO- | 4.2- by DA | | | | | | | |
| Sulfate as SO4 - Turbidimetric | 14808-79-8 | 1 | mg/L | 907 | 280 | 44 | | |
| ED045G: Chloride by Discrete Analyse | | | J | | | | | |
| Chloride | 16887-00-6 | 1 | mg/L | 11900 | 6680 | 341 | | |
| ED093F: Dissolved Major Cations | .000. 00 0 | | J | | | | | |
| Calcium | 7440-70-2 | 1 | mg/L | 181 | 299 | 50 | | |
| Magnesium | 7439-95-4 | 1 | mg/L | 1040 | 786 | 112 | | |
| Sodium | 7440-23-5 | 1 | mg/L | 5980 | 3100 | 373 | | |
| Potassium | 7440-09-7 | 1 | mg/L | 14 | 35 | 6 | | |
| EG005(ED093)F: Dissolved Metals by I | CP-AES | | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 5.05 | 1.22 | <0.05 | 5.15 | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 5.99 | 5.45 | 0.04 | 6.15 | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | <0.010 | 0.005 | <0.001 | <0.010 | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0010 | <0.0001 | <0.0001 | <0.0010 | |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.010 | <0.001 | <0.001 | <0.010 | |
| Copper | 7440-50-8 | 0.001 | mg/L | <0.010 | 0.005 | 0.003 | <0.010 | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.010 | <0.001 | <0.001 | <0.010 | |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.191 | 0.021 | <0.001 | 0.167 | |

Page : 8 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW4 | MW6 | QC101 | |
|--------------------------------------|------------------------|---------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Samplii | ng date / time | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2304011-006 | ES2304011-007 | ES2304011-008 | ES2304011-009 | |
| | | | | Result | Result | Result | Result | |
| EG020F: Dissolved Metals by ICP-M | IS - Continued | | | | | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.225 | <0.005 | <0.005 | 0.196 | |
| EG035F: Dissolved Mercury by FIM | S | | | | | | | |
| Mercury | | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0001 | <0.0001 | |
| EK040P: Fluoride by PC Titrator | | | | | | | | |
| Fluoride | 16984-48-8 | 0.1 | mg/L | 1.2 | 1.6 | 1.8 | | |
| EK055G: Ammonia as N by Discrete | Analyser | | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.22 | 0.34 | 0.02 | | |
| EK057G: Nitrite as N by Discrete A | | | | | | <u> </u> | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.25 | | |
| EK058G: Nitrate as N by Discrete A | nalyser | | | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | <0.01 | 0.01 | 1.00 | | |
| EK059G: Nitrite plus Nitrate as N (N | | vser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.01 | 1.25 | 0.02 | |
| EK061G: Total Kjeldahl Nitrogen By | Discrete Analyser | | J | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.0 | 1.1 | 0.4 | 1.3 | |
| EK062G: Total Nitrogen as N (TKN + | NOv) by Discrete An | alveor | J | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 1.0 | 1.1 | 1.6 | 1.3 | |
| EK067G: Total Phosphorus as P by | | | 3 | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.12 | 0.09 | 0.09 | 0.10 | |
| EK071G: Reactive Phosphorus as P |) by discrete englyser | | 9-2 | •··- | | U.U. | • | |
| Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | <0.01 | <0.01 | | |
| EN055: Ionic Balance | 14203-44-2 | 0.01 | mg/L | -0.01 | -0.01 | 10.01 | | |
| Ø Total Anions | | 0.01 | meg/L | 359 | 216 | 27.2 | | |
| Ø Total Cations | | 0.01 | meg/L | 355 | 215 | 28.1 | | |
| Ø Ionic Balance | | 0.01 | % | 0.55 | 0.25 | 1.61 | | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | <2.0 | <2.0 | <2.0 | | |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |

Page : 9 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW4 | MW6 | QC101 | |
|--|---------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2304011-006 | ES2304011-007 | ES2304011-008 | ES2304011-009 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)A: Phenolic Compounds | - Continued | | | | | | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | <2.0 | <2.0 | <2.0 | | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| ^ Sum of polycyclic aromatic hydrocarb | ons | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| EP080/071: Total Petroleum Hydroca | arbons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | <20 | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| C15 - C28 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| EP080/071: Total Recoverable Hydro | ocarbons - NEPM 201 | 3 Fraction | ns | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | <20 | |
| ^ C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | <20 | |

Page : 10 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW4 | MW6 | QC101 | |
|---|-------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | 08-Feb-2023 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2304011-006 | ES2304011-007 | ES2304011-008 | ES2304011-009 | |
| | | | | Result | Result | Result | Result | |
| EP080/071: Total Recoverable Hydroca | arbons - NEPM 201 | 3 Fraction | ns - Continued | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| ^ >C10 - C16 Fraction minus Naphthalene | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| (F2) | | | | | | | | |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | <1 | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | <2 | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | <2 | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | <2 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | <2 | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | <2 | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | <1 | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | <5 | |
| EP075(SIM)S: Phenolic Compound Su | rrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 32.6 | 29.6 | 31.5 | 27.0 | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 62.2 | 60.2 | 63.8 | 52.4 | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 84.6 | 78.9 | 70.0 | 62.2 | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 77.8 | 77.0 | 81.0 | 66.8 | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 109 | 102 | 104 | 84.4 | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 94.4 | 88.4 | 87.4 | 70.8 | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 99.0 | 102 | 95.4 | 98.2 | |
| Toluene-D8 | 2037-26-5 | 2 | % | 104 | 100.0 | 95.8 | 101 | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 98.9 | 95.0 | 92.1 | 95.6 | |

Page : 11 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery | Limits (%) |
|---|------------|----------|------------|
| Compound | CAS Number | Low | High |
| EP075(SIM)S: Phenolic Compound Surrogates | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 |
| EP075(SIM)T: PAH Surrogates | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 |
| Anthracene-d10 | 1719-06-8 | 27 | 113 |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 |
| EP080S: TPH(V)/BTEX Surrogates | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 71 | 137 |
| Toluene-D8 | 2037-26-5 | 79 | 131 |
| 4-Bromofluorobenzene | 460-00-4 | 70 | 128 |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2304011** Page : 1 of 11

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Telephone : +61 2 8784 8555

Project : S20102 Wetherill Park WME Date Samples Received : 08-Feb-2023

Site :--- Issue Date : 14-Feb-2023
Sampler : Bec Chapple No. of samples received : 9

Sampler : Bec Chapple No. of samples received : 9
Order number : ---- No. of samples analysed : 9

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- Matrix Spike outliers exist please see following pages for full details.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

• NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 11 Work Order : ES2304011

 Client
 : SENVERSA PTY LTD

 Project
 · S20102 Wetherill Park WME

Outliers : Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: WATER

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|--|----------------------|------------------|--------------|------------|------------|--------|----------------------------------|
| Matrix Spike (MS) Recoveries | | | | | | | |
| EG005(ED093)F: Dissolved Metals by ICP-AES | ES2304011007 | MW4 | Manganese | 7439-96-5 | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |
| EK057G: Nitrite as N by Discrete Analyser | ES2303855001 | Anonymous | Nitrite as N | 14797-65-0 | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |

Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | С | ount | Rate | e (%) | Quality Control Specification |
|-----------------------------|----|---------|--------|----------|--------------------------------|
| Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | |
| PAH/Phenols (GC/MS - SIM) | 0 | 7 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 7 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | |
| Dissolved Metals by ICP-AES | 1 | 24 | 4.17 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | 0 | 7 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 7 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive <u>or</u> Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

| Evaluation: 🗴 | = Holding | itime breach : | ; ✓ = Within | holding time. |
|---------------|-----------|----------------|--------------|---------------|
|---------------|-----------|----------------|--------------|---------------|

| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
|--------------------------------------|-------|-------------|----------------|------------------------|------------|---------------|------------------|------------|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EA005P: pH by PC Titrator | | | | | | | | |
| Clear Plastic Bottle - Natural (EA00 | 95-P) | | | | | | | |
| MW1, | MW2, | 08-Feb-2023 | | | | 08-Feb-2023 | 08-Feb-2023 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6 | | | | | | | | |

Page : 3 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: **x** = Holding time breach ; ✓ = Within holding time. Method Sample Date Extraction / Preparation Analysis Container / Client Sample ID(s) Date extracted Due for extraction Evaluation Date analysed Due for analysis Evaluation EA006: Sodium Adsorption Ratio (SAR) Clear Plastic Bottle - Nitric Acid; Filtered (ED093F) 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW1, MW2. MW3. MW4, MW6 EA010P: Conductivity by PC Titrator Clear Plastic Bottle - Natural (EA010-P) MW1. MW2. 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW3. MW4. MW6 EA065: Total Hardness as CaCO3 Clear Plastic Bottle - Nitric Acid; Filtered (ED093F) 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW1, MW2, MW3. MW4, MW6 ED037P: Alkalinity by PC Titrator Clear Plastic Bottle - Natural (ED037-P) MW1, MW2, 08-Feb-2023 08-Feb-2023 22-Feb-2023 MW3, MW4, MW6 ED041G: Sulfate (Turbidimetric) as SO4 2- by DA Clear Plastic Bottle - Natural (ED041G) 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW1. MW2, MW3. MW4. MW6 ED045G: Chloride by Discrete Analyser Clear Plastic Bottle - Natural (ED045G) MW1, MW2, 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW3, MW4, MW6 ED093F: Dissolved Major Cations Clear Plastic Bottle - Nitric Acid; Filtered (ED093F) 08-Feb-2023 08-Feb-2023 08-Mar-2023 MW1, MW2, MW3. MW4, MW6 EG005(ED093)F: Dissolved Metals by ICP-AES Clear Plastic Bottle - Nitric Acid; Filtered (EG005F) 07-Aug-2023 QC301, MW1, 08-Feb-2023 09-Feb-2023 MW2. MW3. MW4. MW6. QC101

Page : 4 of 11 Work Order : ES2304011



| Matrix: WATER | | | | | Evaluation | : x = Holding time | breach ; ✓ = Withi | in holding time |
|---|---------|-------------|----------------|------------------------|------------|--------------------|--------------------|-----------------|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | | | | 08-Feb-2023 | 07-Aug-2023 | ✓ |
| MW2, | MW3, | | | | | | | |
| MW4, | MW6, | | | | | | | |
| QC101 | | | | | | | | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | | | | 09-Feb-2023 | 08-Mar-2023 | ✓ |
| MW2, | MW3, | | | | | | | |
| MW4, | MW6, | | | | | | | |
| QC101 | , | | | | | | | |
| EK040P: Fluoride by PC Titrator | | | | | | | | |
| Clear Plastic Bottle - Natural (EK040P) | | | | | | | | |
| MW1, | MW2, | 08-Feb-2023 | | | | 08-Feb-2023 | 08-Mar-2023 | ✓ |
| MW3, | MW4, | | | | | | | <u> </u> |
| MW6 | , | | | | | | | |
| EK055G: Ammonia as N by Discrete Analyser | | | | | | | | ! |
| Clear Plastic Bottle - Sulfuric Acid (EK055G) | | | | | | | | |
| MW1, | MW2, | 08-Feb-2023 | | | | 13-Feb-2023 | 08-Mar-2023 | 1 |
| MW3, | MW4, | | | | | | | ' |
| MW6 | , | | | | | | | |
| EK057G: Nitrite as N by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Natural (EK057G) | | | | | | | | |
| MW1, | MW2, | 08-Feb-2023 | | | | 08-Feb-2023 | 10-Feb-2023 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6 | | | | | | | | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete A | nalyser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | | | | 13-Feb-2023 | 08-Mar-2023 | ✓ |
| MW2, | MW3, | | | | | | | |
| MW4, | MW6, | | | | | | | |
| QC101 | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | | | | | | | | |
| MW6 | | 08-Feb-2023 | 09-Feb-2023 | 08-Mar-2023 | ✓ | 09-Feb-2023 | 08-Mar-2023 | ✓ |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | 09-Feb-2023 | 08-Mar-2023 | ✓ | 10-Feb-2023 | 08-Mar-2023 | ✓ |
| MW2, | MW3, | | | | | | | |
| MW4 | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | | | | | | | | |
| QC101 | | 08-Feb-2023 | 10-Feb-2023 | 08-Mar-2023 | √ | 11-Feb-2023 | 08-Mar-2023 | ✓ |

Page : 5 of 11
Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: x = Holding time breach ; √ = Within holding time. Method Sample Date Extraction / Preparation Analysis Container / Client Sample ID(s) Pute systematical Properties Fireflycition Pute systematical Properties Fireflycition

| ntainer / Client Sample ID(s) 67G: Total Phosphorus as P by Discrete Analyser Plastic Bottle - Sulfuric Acid (EK067G) | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
|--|-------------|----------------|--------------------|------------|---------------|------------------|------------|
| Plastic Bottle - Sulfuric Acid (EK067G) | | | | | | | |
| Plastic Bottle - Sulfuric Acid (EK067G) | | | | | | | |
| | | | | | | | |
| C301, MW1, | 08-Feb-2023 | 09-Feb-2023 | 08-Mar-2023 | ✓ | 10-Feb-2023 | 08-Mar-2023 | ✓ |
| W2, MW3, | | | | | | | |
| W4, MW6 | | | | | | | |
| Plastic Bottle - Sulfuric Acid (EK067G) | | | | | | | |
| C101 | 08-Feb-2023 | 10-Feb-2023 | 08-Mar-2023 | ✓ | 11-Feb-2023 | 08-Mar-2023 | ✓ |
| 71G: Reactive Phosphorus as P by discrete analyser | | | | | | | |
| Plastic Bottle - Natural (EK071G) | | | | | | | |
| W1, MW2, | 08-Feb-2023 | | | | 08-Feb-2023 | 10-Feb-2023 | ✓ |
| W3, MW4, | | | | | | | |
| W6 | | | | | | | |
| 75(SIM)A: Phenolic Compounds | | | | | | | |
| er Glass Bottle - Unpreserved (EP075(SIM)) | | | | | | | |
| W1, MW2, | 08-Feb-2023 | 10-Feb-2023 | 15-Feb-2023 | 1 | 13-Feb-2023 | 22-Mar-2023 | ✓ |
| W3, MW4, | | | | | | | |
| W6 | | | | | | | |
| 75(SIM)B: Polynuclear Aromatic Hydrocarbons | | | | | | | |
| er Glass Bottle - Unpreserved (EP075(SIM)) | | | | | | | |
| C301, MW1, | 08-Feb-2023 | 10-Feb-2023 | 15-Feb-2023 | ✓ | 13-Feb-2023 | 22-Mar-2023 | ✓ |
| W2, MW3, | | | | | | | |
| W4, MW6, | | | | | | | |
| C101 | | | | | | | |
| 80/071: Total Petroleum Hydrocarbons | | | | | | | |
| er Glass Bottle - Unpreserved (EP071) | | | | | | | |
| C301, MW1, | 08-Feb-2023 | 10-Feb-2023 | 15-Feb-2023 | ✓ | 13-Feb-2023 | 22-Mar-2023 | ✓ |
| W2, MW3, | | | | | | | |
| W4, MW6, | | | | | | | |
| C101 | | | | | | | |
| er VOC Vial - Sulfuric Acid (EP080) | | | | | | | |
| C401 | 06-Feb-2023 | 09-Feb-2023 | 20-Feb-2023 | ✓ | 09-Feb-2023 | 20-Feb-2023 | ✓ |
| er VOC Vial - Sulfuric Acid (EP080) | | | | | | | |
| C301, MW1, | 08-Feb-2023 | 09-Feb-2023 | 22-Feb-2023 | ✓ | 09-Feb-2023 | 22-Feb-2023 | ✓ |
| W2, MW3, | | | | | | | |
| | | I | | | I | | |
| W4, MW6, | | | | | | | |

Page : 6 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: **WATER**Evaluation: × = Holding time breach; ✓ = Within holding time.

| Method | | Sample Date | E | xtraction / Preparation | | Analysis | | | |
|--|----------------------------|-------------|----------------|-------------------------|------------|---------------|------------------|------------|--|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EP080/071: Total Recoverable Hydrocarb | bons - NEPM 2013 Fractions | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP07 | 1) | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | 10-Feb-2023 | 15-Feb-2023 | ✓ | 13-Feb-2023 | 22-Mar-2023 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW4, | MW6, | | | | | | | | |
| QC101 | | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC401 | | 06-Feb-2023 | 09-Feb-2023 | 20-Feb-2023 | ✓ | 09-Feb-2023 | 20-Feb-2023 | ✓ | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | 09-Feb-2023 | 22-Feb-2023 | ✓ | 09-Feb-2023 | 22-Feb-2023 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW4, | MW6, | | | | | | | | |
| QC101 | | | | | | | | | |
| EP080: BTEXN | | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC401, | QC501 | 06-Feb-2023 | 09-Feb-2023 | 20-Feb-2023 | ✓ | 09-Feb-2023 | 20-Feb-2023 | ✓ | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC301, | MW1, | 08-Feb-2023 | 09-Feb-2023 | 22-Feb-2023 | ✓ | 09-Feb-2023 | 22-Feb-2023 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW4, | MW6, | | | | | | | | |
| QC101 | | | | | | | | | |

Page : 7 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: WATER

Evaluation: x = Quality Control frequency not within specification: √ = Quality Control frequency within specification

| Matrix: WATER Evaluation: ★ = Quality Control frequency not within specification; ✓ = Quality Control frequency within specification; | | | | | | | | | | | |
|---|------------|----|---------|--------|----------|------------|--------------------------------|--|--|--|--|
| Quality Control Sample Type | | C | ount | | Rate (%) | | Quality Control Specification | | | | |
| Analytical Methods | Method | ОC | Regular | Actual | Expected | Evaluation | | | | | |
| Laboratory Duplicates (DUP) | | | | | | | | | | | |
| Alkalinity by Auto Titrator | ED037-P | 2 | 14 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Ammonia as N by Discrete analyser | EK055G | 3 | 21 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Chloride by Discrete Analyser | ED045G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Conductivity by Auto Titrator | EA010-P | 5 | 43 | 11.63 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 8 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Metals by ICP-AES | EG005F | 3 | 24 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 8 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Fluoride by Auto Titrator | EK040P | 2 | 14 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Major Cations - Dissolved | ED093F | 3 | 23 | 13.04 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 4 | 34 | 11.76 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Nitrite as N by Discrete Analyser | EK057G | 2 | 10 | 20.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 7 | 0.00 | 10.00 | 3£ | NEPM 2013 B3 & ALS QC Standard | | | | |
| pH by Auto Titrator | EA005-P | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Reactive Phosphorus as P-By Discrete Analyser | EK071G | 3 | 22 | 13.64 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Sulfate (Turbidimetric) as SO4 2- by Discrete Analyser | ED041G | 1 | 8 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 22 | 13.64 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Total Phosphorus as P By Discrete Analyser | EK067G | 4 | 23 | 17.39 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| TRH - Semivolatile Fraction | EP071 | 0 | 7 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard | | | | |
| TRH Volatiles/BTEX | EP080 | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Laboratory Control Samples (LCS) | | | | | | | | | | | |
| Alkalinity by Auto Titrator | ED037-P | 2 | 14 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 21 | 9.52 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Chloride by Discrete Analyser | ED045G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Conductivity by Auto Titrator | EA010-P | 4 | 43 | 9.30 | 8.33 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Metals by ICP-AES | EG005F | 2 | 24 | 8.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Fluoride by Auto Titrator | EK040P | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Major Cations - Dissolved | ED093F | 2 | 23 | 8.70 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 3 | 34 | 8.82 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 10 | 10.00 | 5.00 | √ | NEPM 2013 B3 & ALS QC Standard | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| pH by Auto Titrator | EA005-P | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Reactive Phosphorus as P-By Discrete Analyser | EK071G | 2 | 22 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Sulfate (Turbidimetric) as SO4 2- by Discrete Analyser | ED041G | 2 | 8 | 25.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 6 | 22 | 27.27 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | | | |
| | | | - | | | | • | | | | |

Page : 8 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: **WATER**Evaluation: **×** = Quality Control frequency not within specification; ✓ = Quality Control frequency within specification.

| Quality Control Sample Type | | Co | ount | Rate (%) | | | Quality Control Specification |
|--|------------|----|---------|----------|----------|------------|--------------------------------|
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | |
| Laboratory Control Samples (LCS) - Continued | | | | | | | |
| Total Phosphorus as P By Discrete Analyser | EK067G | 6 | 23 | 26.09 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 21 | 9.52 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Chloride by Discrete Analyser | ED045G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Conductivity by Auto Titrator | EA010-P | 1 | 43 | 2.33 | 1.67 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 2 | 24 | 8.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Fluoride by Auto Titrator | EK040P | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Major Cations - Dissolved | ED093F | 2 | 23 | 8.70 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 3 | 34 | 8.82 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Reactive Phosphorus as P-By Discrete Analyser | EK071G | 2 | 22 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Sulfate (Turbidimetric) as SO4 2- by Discrete Analyser | ED041G | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 22 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 23 | 8.70 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 21 | 9.52 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Chloride by Discrete Analyser | ED045G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 24 | 4.17 | 5.00 | se | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Fluoride by Auto Titrator | EK040P | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 3 | 34 | 8.82 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 7 | 0.00 | 5.00 | 3£ | NEPM 2013 B3 & ALS QC Standard |
| Reactive Phosphorus as P-By Discrete Analyser | EK071G | 2 | 22 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Sulfate (Turbidimetric) as SO4 2- by Discrete Analyser | ED041G | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 22 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 23 | 8.70 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 7 | 0.00 | 5.00 | se | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |

Page : 9 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|--|----------|--------|---|
| pH by Auto Titrator | EA005-P | WATER | In house: Referenced to APHA 4500 H+ B. This procedure determines pH of water samples by automated ISE. This method is compliant with NEPM Schedule B(3) |
| Conductivity by Auto Titrator | EA010-P | WATER | In house: Referenced to APHA 2510 B. This procedure determines conductivity by automated ISE. This method is compliant with NEPM Schedule B(3) |
| Calculated TDS (from Electrical Conductivity) | EA016 | WATER | In house: Calculation from Electrical Conductivity (APHA 2510 B) using a conversion factor specified in the analytical report. This method is compliant with NEPM Schedule B(3) |
| Alkalinity by Auto Titrator | ED037-P | WATER | In house: Referenced to APHA 2320 B This procedure determines alkalinity by automated measurement (e.g. PC Titrate) on a settled supernatant aliquot of the sample using pH 4.5 for indicating the total alkalinity end-point. This method is compliant with NEPM Schedule B(3) |
| Sulfate (Turbidimetric) as SO4 2- by Discrete Analyser | ED041G | WATER | In house: Referenced to APHA 4500-SO4. Dissolved sulfate is determined in a 0.45um filtered sample. Sulfate ions are converted to a barium sulfate suspension in an acetic acid medium with barium chloride. Light absorbance of the BaSO4 suspension is measured by a photometer and the SO4-2 concentration is determined by comparison of the reading with a standard curve. This method is compliant with NEPM Schedule B(3) |
| Chloride by Discrete Analyser | ED045G | WATER | In house: Referenced to APHA 4500 CI - G.The thiocyanate ion is liberated from mercuric thiocyanate through sequestration of mercury by the chloride ion to form non-ionised mercuric chloride. In the presence of ferric ions the liberated thiocynate forms highly-coloured ferric thiocynate which is measured at 480 nm. |
| Major Cations - Dissolved | ED093F | WATER | In house: Referenced to APHA 3120 and 3125; USEPA SW 846 - 6010 and 6020; Cations are determined by either ICP-AES or ICP-MS techniques. This method is compliant with NEPM Schedule B(3) Sodium Adsorption Ratio is calculated from Ca, Mg and Na which determined by ALS in house method QWI-EN/ED093F. This method is compliant with NEPM Schedule B(3) Hardness parameters are calculated based on APHA 2340 B. This method is compliant with NEPM Schedule B(3) |
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered samples, emitting a characteristic spectrum which is compared against matrix matched standards. This method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Fluoride by Auto Titrator | EK040P | WATER | In house: Referenced to APHA 4500-F C: CDTA is added to the sample to provide a uniform ionic strength background, adjust pH, and break up complexes. Fluoride concentration is determined by either manual or automatic ISE measurement. This method is compliant with NEPM Schedule B(3) |

Page : 10 of 11 Work Order : ES2304011



| Analytical Methods | Method | Matrix | Method Descriptions |
|---|--------------|--------|---|
| Ammonia as N by Discrete analyser | EK055G | WATER | In house: Referenced to APHA 4500-NH3 G Ammonia is determined by direct colorimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrite as N by Discrete Analyser | EK057G | WATER | In house: Referenced to APHA 4500-NO2- B. Nitrite is determined by direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrate as N by Discrete Analyser | EK058G | WATER | In house: Referenced to APHA 4500-NO3- F. Nitrate is reduced to nitrite by way of a chemical reduction followed by quantification by Discrete Analyser. Nitrite is determined seperately by direct colourimetry and result for Nitrate calculated as the difference between the two results. This method is compliant with NEPM Schedule B(3) |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Reactive Phosphorus as P-By Discrete Analyser | EK071G | WATER | In house: Referenced to APHA 4500-P F Ammonium molybdate and potassium antimonyl tartrate reacts in acid medium with othophosphate to form a heteropoly acid -phosphomolybdic acid - which is reduced to intensely coloured molybdenum blue by ascorbic acid. Quantification is by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Ionic Balance by PCT DA and Turbi SO4 DA | * EN055 - PG | WATER | In house: Referenced to APHA 1030F. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| Preparation Methods | Method | Matrix | Method Descriptions |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |

Page : 11 of 11 Work Order : ES2304011



| Preparation Methods | Method | Matrix | Method Descriptions |
|---|---------|--------|--|
| Separatory Funnel Extraction of Liquids | ORG14 | WATER | In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated and concentrated for analysis. This method is compliant with NEPM Schedule B(3). ALS default excludes sediment which may be resident in the container. |
| Volatiles Water Preparation | ORG16-W | WATER | A 5 mL aliquot or 5 mL of a diluted sample is added to a 40 mL VOC vial for purging. |



QUALITY CONTROL REPORT

Page

: 1 of 11

Work Order : **ES2304011**

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Helen Simpson

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : +61 2 8784 8555

Project : S20102 Wetherill Park WME Date Samples Received : 08-Feb-2023

Order number Date Analysis Commenced : 08-Feb-2023

Order number : ---- Date Analysis Commenced : 08-Feb-2023

C-O-C number ---- Issue Date 14-Feb-2023

Sampler ; Bec Chapple

Site · ----

Quote number : SY/103/22

No. of samples received : 9
No. of samples analysed : 9

Accreditation No. 825
Accredited for compliance with ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Edwandy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|-------------------------|--|-------------|------|---------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | solved Metals by ICP-A | NES (QC Lot: 4859853) | | | | | | | |
| ES2303873-004 | Anonymous | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.04 | 115 | No Limit |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 0.10 | 0.10 | 0.0 | No Limit |
| ES2303873-014 | Anonymous | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 0.28 | 0.30 | 4.1 | 0% - 20% |
| EG005(ED093)F: Dis | solved Metals by ICP-A | NES (QC Lot: 4859854) | | | | | | | |
| ES2304011-006 | MW3 | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 5.99 | 6.28 | 4.6 | 0% - 20% |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 5.05 | 5.29 | 4.7 | 0% - 20% |
| EA005P: pH by PC T | itrator (QC Lot: 48599 | 14) | | | | | | | |
| ES2303998-001 | Anonymous | EA005-P: pH Value | | 0.01 | pH Unit | 8.25 | 8.21 | 0.5 | 0% - 20% |
| ES2304011-007 | MW4 | EA005-P: pH Value | | 0.01 | pH Unit | 7.72 | 7.73 | 0.1 | 0% - 20% |
| EA010P: Conductivi | ty by PC Titrator (QC L | ot: 4859910) | | | | | | | |
| ES2303998-001 | Anonymous | EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | 488 | 486 | 0.4 | 0% - 20% |
| ES2303799-001 | Anonymous | EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | 1240 | 1230 | 0.6 | 0% - 20% |
| ES2303915-029 | Anonymous | EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | 112 | 111 | 1.0 | 0% - 20% |
| ES2303915-015 | Anonymous | EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | 3 | 3 | 0.0 | No Limit |
| ES2304011-007 | MW4 | EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | 19900 | 19800 | 0.4 | 0% - 20% |
| ED037P: Alkalinity b | y PC Titrator (QC Lot: | 4859913) | | | | | | | |
| ES2303934-009 | Anonymous | ED037-P: Hydroxide Alkalinity as CaCO3 | DMO-210-001 | 1 | mg/L | <1 | <1 | 0.0 | No Limit |
| | | ED037-P: Carbonate Alkalinity as CaCO3 | 3812-32-6 | 1 | mg/L | <1 | <1 | 0.0 | No Limit |
| | | ED037-P: Bicarbonate Alkalinity as CaCO3 | 71-52-3 | 1 | mg/L | 14 | 13 | 10.4 | 0% - 50% |
| | | ED037-P: Total Alkalinity as CaCO3 | | 1 | mg/L | 14 | 13 | 10.4 | 0% - 50% |
| ES2304011-007 | MW4 | ED037-P: Hydroxide Alkalinity as CaCO3 | DMO-210-001 | 1 | mg/L | <1 | <1 | 0.0 | No Limit |
| | | ED037-P: Carbonate Alkalinity as CaCO3 | 3812-32-6 | 1 | mg/L | <1 | <1 | 0.0 | No Limit |
| | | ED037-P: Bicarbonate Alkalinity as CaCO3 | 71-52-3 | 1 | mg/L | 1110 | 927 | 17.7 | 0% - 20% |

Page : 3 of 11
Work Order : ES2304011



| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|---------------------------|--|------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| ED037P: Alkalinity | by PC Titrator (QC Lot: | 4859913) - continued | | | | | | | |
| ES2304011-007 | MW4 | ED037-P: Total Alkalinity as CaCO3 | | 1 | mg/L | 1110 | 927 | 17.7 | 0% - 20% |
| ED041G: Sulfate (T | urbidimetric) as SO4 2- b | by DA (QC Lot: 4860151) | | | | | | | |
| ES2304009-005 | Anonymous | ED041G: Sulfate as SO4 - Turbidimetric | 14808-79-8 | 1 | mg/L | 10 | 10 | 0.0 | 0% - 50% |
| ED045G: Chloride b | by Discrete Analyser (Q | C Lot: 4860150) | | | | | | | |
| ES2304029-010 | Anonymous | ED045G: Chloride | 16887-00-6 | 1 | mg/L | 13 | 13 | 0.0 | 0% - 50% |
| ES2304009-005 | Anonymous | ED045G: Chloride | 16887-00-6 | 1 | mg/L | 5 | 5 | 0.0 | No Limit |
| ED093F: Dissolved | Major Cations (QC Lot: | 4859850) | | | _ | | | | |
| ES2303873-013 | Anonymous | ED093F: Calcium | 7440-70-2 | 1 | mg/L | 30 | 30 | 0.0 | 0% - 20% |
| | , | ED093F: Magnesium | 7439-95-4 | 1 | mg/L | 41 | 40 | 0.0 | 0% - 20% |
| | | ED093F: Sodium | 7440-23-5 | 1 | mg/L | 493 | 482 | 2.1 | 0% - 20% |
| | | ED093F: Potassium | 7440-09-7 | 1 | mg/L | 38 | 38 | 0.0 | 0% - 20% |
| ES2303640-001 | Anonymous | ED093F: Calcium | 7440-70-2 | 1 | mg/L | 248 | 252 | 1.3 | 0% - 20% |
| | | ED093F: Magnesium | 7439-95-4 | 1 | mg/L | 66 | 66 | 0.0 | 0% - 20% |
| | | ED093F: Sodium | 7440-23-5 | 1 | mg/L | 1 | 1 | 0.0 | No Limit |
| | | ED093F: Potassium | 7440-09-7 | 1 | mg/L | 8 | 8 | 0.0 | No Limit |
| ED093F: Dissolved | Major Cations (QC Lot: | 4859855) | | | | | | | |
| ES2304011-008 | MW6 | ED093F: Calcium | 7440-70-2 | 1 | mg/L | 50 | 48 | 3.3 | 0% - 20% |
| | | ED093F: Magnesium | 7439-95-4 | 1 | mg/L | 112 | 111 | 0.0 | 0% - 20% |
| | | ED093F: Sodium | 7440-23-5 | 1 | mg/L | 373 | 389 | 4.2 | 0% - 20% |
| | | ED093F: Potassium | 7440-09-7 | 1 | mg/L | 6 | 5 | 0.0 | No Limit |
| EG020F: Dissolved | Metals by ICP-MS (QC I | Lot: 4859851) | | | | | | | |
| ES2303640-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | 0.538 | 0.546 | 1.6 | 0% - 20% |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.034 | 0.034 | 0.0 | 0% - 20% |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 782 | 786 | 0.5 | 0% - 20% |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | 2.72 | 2.74 | 0.7 | 0% - 20% |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 2.03 | 2.04 | 0.7 | 0% - 20% |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 134 | 129 | 3.5 | 0% - 20% |
| EG035F: Dissolved | Mercury by FIMS (QC L | .ot: 4859849) | | | | | | | |
| ES2304011-004 | MW1 | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| EK040P: Fluoride b | y PC Titrator (QC Lot: 4 | 859908) | | | | | | | |
| ES2303799-001 | Anonymous | EK040P: Fluoride | 16984-48-8 | 0.1 | mg/L | 0.1 | <0.1 | 0.0 | No Limit |
| ES2304011-007 | MW4 | EK040P: Fluoride | 16984-48-8 | 0.1 | mg/L | 1.6 | 1.6 | 0.0 | 0% - 50% |
| EK055G: Ammonia | as N by Discrete Analys | ser (QC Lot: 4861210) | | | | | | | |
| ES2303838-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.01 | 0.01 | 0.0 | No Limit |
| ES2303866-007 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.07 | 0.02 | 93.9 | No Limit |
| EK055G: Ammonia | as N by Discrete Analys | | | | | | | | |
| ES2304011-008 | MW6 | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.02 | 0.02 | 0.0 | No Limit |

Page : 4 of 11
Work Order : ES2304011



| Sub-Matrix: WATER | | | | | | Laboratory L | Ouplicate (DUP) Report | | |
|----------------------|------------------------------|--------------------------------------|------------|------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EK057G: Nitrite as N | by Discrete Analyser (QC I | | | | | | | | |
| ES2303855-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | 4.04 | 4.01 | 0.9 | 0% - 20% |
| EW2300593-011 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK059G: Nitrite plus | Nitrate as N (NOx) by Disc | rete Analyser (QC Lot: 4861209) | | | | | | | |
| ES2303838-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.61 | 0.61 | 0.0 | 0% - 20% |
| ES2303866-007 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.01 | 0.01 | 0.0 | No Limit |
| EK059G: Nitrite plus | Nitrate as N (NOx) by Disc | rete Analyser (QC Lot: 4861211) | | | | | | | |
| ES2304011-008 | MW6 | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 1.25 | 1.18 | 6.0 | 0% - 20% |
| EK059G: Nitrite plus | Nitrate as N (NOx) by Disc | rete Analyser (QC Lot: 4864095) | | | | | | | |
| ME2300270-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.01 | 0.02 | 0.0 | No Limit |
| EK061G: Total Kjelda | ahl Nitrogen By Discrete Ana | alyser (QC Lot: 4861208) | | | | | | | |
| ES2304011-003 | QC301 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | <0.1 | 0.0 | No Limit |
| EK061G: Total Kjelda | ahl Nitrogen By Discrete Ana | , , | | | | | | | |
| ES2303697-005 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 4.7 | 4.6 | 2.7 | 0% - 20% |
| EW2300556-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 232 | 246 | 5.7 | 0% - 50% |
| EK067G: Total Phos | phorus as P by Discrete Ana | lyser (QC Lot: 4861207) | | | | | | | |
| ES2303955-004 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 296 | 316 | 6.4 | 0% - 20% |
| ES2304011-003 | QC301 | EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 0.03 | 101 | No Limit |
| EK067G: Total Phos | phorus as P by Discrete Ana | lyser (QC Lot: 4864091) | | | | | | | |
| ES2303697-005 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.46 | 0.44 | 4.7 | 0% - 20% |
| EW2300556-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 37.0 | 37.9 | 2.6 | 0% - 20% |
| EK071G: Reactive Pl | hosphorus as P by discrete a | analyser (QC Lot: 4860147) | | | | | | | |
| ES2303957-001 | Anonymous | EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| ES2304009-005 | Anonymous | EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK071G: Reactive Pl | hosphorus as P by discrete a | analyser (QC Lot: 4860152) | | | | | | | |
| ES2304011-008 | MW6 | EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | 0.02 | 67.3 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbons (QC I | Lot: 4860105) | | | | | | | |
| ES2303866-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2304011-003 | QC301 | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Red | coverable Hydrocarbons - NI | EPM 2013 Fractions (QC Lot: 4860105) | | | | | | | |
| ES2303866-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2304011-003 | QC301 | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 4860105) | | | | | | | | |
| ES2303866-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | _ | _ | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |

Page : 5 of 11 Work Order : ES2304011



| Sub-Matrix: WATER | ub-Matrix: WATER | | | Laboratory Duplicate (DUP) Report | | | | | | |
|----------------------|---------------------------|----------------------------|------------|-----------------------------------|------|-----------------|------------------|---------|--------------------|--|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) | |
| EP080: BTEXN (QC | Lot: 4860105) - continued | | | | | | | | | |
| ES2303866-001 | Anonymous | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit | |
| ES2304011-003 | QC301 | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit | |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | |
| | | | 106-42-3 | | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit | |

Page : 6 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | | |
|---|------------|--------|---------|-------------------|---------------------------------------|--------------------|--------------|--------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 48 | 359853) | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 93.5 | 82.0 | 114 |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 100 | 81.0 | 113 |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 48 | 359854) | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 97.0 | 82.0 | 114 |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 98.2 | 81.0 | 113 |
| EA005P: pH by PC Titrator (QCLot: 4859914) | | | | | - | | | |
| EA005-P: pH Value | | | pH Unit | | 4 pH Unit | 101 | 98.8 | 101 |
| ZAGGOT : pri valido | | | | | 7 pH Unit | 100 | 99.2 | 101 |
| EA010P: Conductivity by PC Titrator (QCLot: 4859910) | | | | | | | | |
| EA010-P: Electrical Conductivity @ 25°C | | 1 | μS/cm | <1 | 220 µS/cm | 93.5 | 89.9 | 110 |
| EACTO-1 : Electrical Conductivity @ 25 C | | · | μο/οπ | <1 | 2100 µS/cm | 101 | 90.2 | 111 |
| ED037P: Alkalinity by PC Titrator (QCLot: 4859913) | | | | | | | | |
| ED037-P: Total Alkalinity as CaCO3 | | | mg/L | | 200 mg/L | 92.9 | 81.0 | 111 |
| ED037-F. Total Alkalillity as CaCO3 | | | mg/L | | 50 mg/L | 111 | 80.0 | 120 |
| EDO440: Culfata (Tunkidimatuia) aa CO40 ku DA (OC) at | 4000454) | | | | 00g/_ | | 00.0 | |
| ED041G: Sulfate (Turbidimetric) as SO4 2- by DA (QCLot: | 14808-79-8 | 1 | ma/l | <1 | 25 mg/L | 108 | 82.0 | 122 |
| ED041G: Sulfate as SO4 - Turbidimetric | 14000-79-0 | ' | mg/L | <1 | 500 mg/L | 108 | 82.0 | 122 |
| | | | | 1 | 300 mg/L | 107 | 02.0 | 122 |
| ED045G: Chloride by Discrete Analyser (QCLot: 4860150) | | 4 | | -11 | 50 mm m // | 405 | 00.0 | 407 |
| ED045G: Chloride | 16887-00-6 | 1 | mg/L | <1 <1 | 50 mg/L 1000 mg/L | 105 100 | 80.9 80.9 | 127 127 |
| | | | | <u> </u> | 1000 Hig/L | 100 | 60.9 | 127 |
| ED093F: Dissolved Major Cations (QCLot: 4859850) | | | | | | | | |
| ED093F: Calcium | 7440-70-2 | 1 | mg/L | <1 | 50 mg/L | 97.3 | 80.0 | 114 |
| ED093F: Magnesium | 7439-95-4 | 1 | mg/L | <1 | 50 mg/L | 102 | 90.0 | 116 |
| ED093F: Sodium | 7440-23-5 | 1 | mg/L | <1 | 50 mg/L | 105 | 82.0 | 120 |
| ED093F: Potassium | 7440-09-7 | 1 | mg/L | <1 | 50 mg/L | 104 | 85.0 | 113 |
| ED093F: Dissolved Major Cations (QCLot: 4859855) | | | | | | | | |
| ED093F: Calcium | 7440-70-2 | 1 | mg/L | <1 | 50 mg/L | 97.2 | 80.0 | 114 |
| ED093F: Magnesium | 7439-95-4 | 1 | mg/L | <1 | 50 mg/L | 104 | 90.0 | 116 |
| ED093F: Sodium | 7440-23-5 | 1 | mg/L | <1 | 50 mg/L | 112 | 82.0 | 120 |
| ED093F: Potassium | 7440-09-7 | 1 | mg/L | <1 | 50 mg/L | 104 | 85.0 | 113 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 4859851) | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 97.9 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 96.6 | 84.0 | 110 |

Page : 7 of 11
Work Order : ES2304011



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|--|-----------------------|--------|--------|-------------------|---|------------------------------|--------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 485 | 9851) - continued | | | | | | | |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.0 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 94.7 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.2 | 83.0 | 111 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.0 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 96.4 | 81.0 | 117 |
| EG035F: Dissolved Mercury by FIMS (QCLot: 4859 | 849) | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 99.6 | 83.0 | 105 |
| EK040P: Fluoride by PC Titrator (QCLot: 4859908) | | | | | | | | |
| EK040P: Fluoride | 16984-48-8 | 0.1 | mg/L | <0.1 | 5 mg/L | 96.4 | 82.0 | 116 |
| EK055G: Ammonia as N by Discrete Analyser (QC | Lot: 4861210) | | | | | | | |
| EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | <0.01 | 1 mg/L | 110 | 90.0 | 114 |
| EK055G: Ammonia as N by Discrete Analyser (QC | ot: 4861212) | | | | | | | |
| EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | <0.01 | 1 mg/L | 110 | 90.0 | 114 |
| EK057G: Nitrite as N by Discrete Analyser (QCLot | · 4860146) | | | | | | | |
| EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 98.4 | 82.0 | 114 |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discret | | | ···g- | | *************************************** | 2211 | | |
| EK059G: Nitrite + Nitrate as N | e Analyser (QCLOL 46 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 103 | 91.0 | 113 |
| | | | 9.2 | 0.01 | 0.0 mg/2 | .00 | 00 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discret EK059G: Nitrite + Nitrate as N | e Analyser (QCLot: 48 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 104 | 91.0 | 113 |
| | | | IIIg/L | ~0.01 | 0.5 mg/L | 104 | 91.0 | 113 |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discret | | | | <0.01 | 0.5 | 05.0 | 04.0 | 440 |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 95.9 | 91.0 | 113 |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analy | | • | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 <0.1 | 10 mg/L | 97.9 105 | 69.0 70.0 | 101 118 |
| | | | | <0.1 | 1 mg/L 5 mg/L | 105 | 70.0 | 130 |
| | (00) ((00) | | | 40.1 | J Hig/L | 103 | 70.0 | 130 |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analy | ser (QCLot: 4864092) | 0.1 | ma/l | <0.1 | 10 mg/l | 100 | 69.0 | 101 |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 <0.1 | 10 mg/L 1 mg/L | 100 | 69.0 70.0 | 101 118 |
| | | | | <0.1 | 5 mg/L | 104 | 70.0 | 130 |
| EVOCTO: Tatal Phase have as B by Bisserts Aught | (OCL -t: 4004007) | | | .0.1 | o mg/L | 101 | 7 0.0 | 100 |
| EK067G: Total Phosphorus as P by Discrete Analyse EK067G: Total Phosphorus as P | ser (QCLot: 4861207) | 0.01 | mg/L | <0.01 | 4.42 mg/L | 97.7 | 71.3 | 126 |
| EROOTO. Total Phosphorus as P | | 0.01 | IIIg/L | <0.01 | 0.442 mg/L | 96.4 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 98.7 | 71.3 | 126 |
| EK067G: Total Phosphorus as P by Discrete Analys | ser (OCL of: 4864094) | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 92.7 | 71.3 | 126 |
| Enter 6. Total i Hospitoras as i | | | | <0.01 | 0.442 mg/L | 95.2 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 99.5 | 71.3 | 126 |

Page : 8 of 11 Work Order : ES2304011



| Sub-Matrix: WATER | | | | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | | | |
|--|--------------------------|------|------|-------------------|---------------------------------------|--------------------|------------|------------|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EK071G: Reactive Phosphorus as P by discrete a | analyser (QCLot: 486014) | 7) | | | | | | | |
| EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 98.8 | 85.0 | 117 | |
| EK071G: Reactive Phosphorus as P by discrete a | analyser (QCLot: 486015 | 2) | | | | | | | |
| EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 100.0 | 85.0 | 117 | |
| EP075(SIM)A: Phenolic Compounds (QCLot: 485 | 59814) | | | | | | | | |
| EP075(SIM): Phenol | 108-95-2 | 1 | μg/L | <1.0 | 5 μg/L | 30.5 | 24.5 | 61.9 | |
| EP075(SIM): 2-Chlorophenol | 95-57-8 | 1 | μg/L | <1.0 | 5 μg/L | 61.4 | 52.0 | 90.0 | |
| EP075(SIM): 2-Methylphenol | 95-48-7 | 1 | μg/L | <1.0 | 5 μg/L | 55.1 | 51.0 | 91.0 | |
| EP075(SIM): 3- & 4-Methylphenol | 1319-77-3 | 2 | μg/L | <2.0 | 10 μg/L | 51.2 | 44.0 | 88.0 | |
| EP075(SIM): 2-Nitrophenol | 88-75-5 | 1 | μg/L | <1.0 | 5 μg/L | 67.1 | 48.0 | 100 | |
| EP075(SIM): 2.4-Dimethylphenol | 105-67-9 | 1 | μg/L | <1.0 | 5 μg/L | 49.3 | 49.0 | 99.0 | |
| EP075(SIM): 2.4-Dichlorophenol | 120-83-2 | 1 | μg/L | <1.0 | 5 μg/L | 68.7 | 53.0 | 105 | |
| EP075(SIM): 2.6-Dichlorophenol | 87-65-0 | 1 | μg/L | <1.0 | 5 μg/L | 68.7 | 57.0 | 105 | |
| EP075(SIM): 4-Chloro-3-methylphenol | 59-50-7 | 1 | μg/L | <1.0 | 5 μg/L | 64.3 | 53.0 | 99.0 | |
| EP075(SIM): 2.4.6-Trichlorophenol | 88-06-2 | 1 | μg/L | <1.0 | 5 μg/L | 71.6 | 50.0 | 106 | |
| EP075(SIM): 2.4.5-Trichlorophenol | 95-95-4 | 1 | μg/L | <1.0 | 5 μg/L | 70.2 | 51.0 | 105 | |
| EP075(SIM): Pentachlorophenol | 87-86-5 | 2 | μg/L | <2.0 | 10 μg/L | 84.8 | 10.0 | 95.0 | |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbo | ns (QCLot: 4859814) | | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 69.4 | 50.0 | 94.0 | |
| EP075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 71.4 | 63.6 | 114 | |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 71.4 | 62.2 | 113 | |
| EP075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 74.4 | 63.9 | 115 | |
| EP075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 100 | 62.6 | 116 | |
| EP075(SIM): Anthracene | 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 94.9 | 64.3 | 116 | |
| EP075(SIM): Fluoranthene | 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 100 | 63.6 | 118 | |
| EP075(SIM): Pyrene | 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 101 | 63.1 | 118 | |
| EP075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 80.2 | 64.1 | 117 | |
| EP075(SIM): Chrysene | 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 84.4 | 62.5 | 116 | |
| EP075(SIM): Benzo(b+j)fluoranthene | 205-99-2 | 1 | μg/L | <1.0 | 5 μg/L | 91.4 | 61.7 | 119 | |
| | 205-82-3 | | | | | | | | |
| EP075(SIM): Benzo(k)fluoranthene | 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 72.8 | 63.0 | 115 | |
| EP075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 79.4 | 63.3 | 117 | |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 81.2 | 59.9 | 118 | |
| EP075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 81.6 | 61.2 | 117 | |
| EP075(SIM): Benzo(g.h.i)perylene | 191-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 81.4 | 59.1 | 118 | |
| EP080/071: Total Petroleum Hydrocarbons (QCL | .ot: 4859815) | | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 84.4 | 53.7 | 97.0 | |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 78.0 | 63.3 | 107 | |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 91.6 | 58.3 | 120 | |

Page : 9 of 11 Work Order : ES2304011

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | | | |
|--|--------------------|--------------|------|-------------------|---------------------------------------|--------------------|------------|------------|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 48 | 60105) | | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 98.9 | 75.0 | 127 | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2 | 013 Fractions (QCL | ot: 4859815) | | | | | | | |
| EP071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 83.4 | 53.9 | 95.5 | |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 80.3 | 57.8 | 110 | |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 93.6 | 50.5 | 115 | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2 | 013 Fractions (QCL | ot: 4860105) | | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 101 | 75.0 | 127 | |
| EP080: BTEXN (QCLot: 4860105) | | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 107 | 70.0 | 122 | |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 108 | 69.0 | 123 | |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 109 | 70.0 | 120 | |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 104 | 69.0 | 121 | |
| | 106-42-3 | | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 106 | 72.0 | 122 | |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 109 | 70.0 | 120 | |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| ub-Matrix: WATER | | | | Matrix Spike (MS) Report | | | | |
|---------------------|---|--|------------|--------------------------|---------------------|--------------|------------|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable l | Limits (%) | |
| aboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | |
| G005(ED093)F: [| Dissolved Metals by ICP-AES (QCLot: 4859854) | | | | | | | |
| ES2304011-007 | MW4 | EG005F: Manganese | 7439-96-5 | 1 mg/L | # Not Determined | 70.0 | 130 | |
| D041G: Sulfate (| Turbidimetric) as SO4 2- by DA (QCLot: 4860151) | | | | | | | |
| ES2304009-005 | Anonymous | ED041G: Sulfate as SO4 - Turbidimetric | 14808-79-8 | 10 mg/L | 117 | 70.0 | 130 | |
| D045G: Chloride | by Discrete Analyser (QCLot: 4860150) | | | | | | | |
| ES2304009-005 | Anonymous | ED045G: Chloride | 16887-00-6 | 50 mg/L | 107 | 70.0 | 130 | |
| G020F: Dissolve | d Metals by ICP-MS (QCLot: 4859851) | | | | | | | |
| ES2304011-005 | MW2 | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 108 | 70.0 | 130 | |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 92.8 | 70.0 | 130 | |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 93.8 | 70.0 | 130 | |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 101 | 70.0 | 130 | |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 107 | 70.0 | 130 | |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 101 | 70.0 | 130 | |

Page : 10 of 11 Work Order : ES2304011



| Sub-Matrix: WATER | Alatrix: WATER | | | | Matrix Spike (MS) Report | | | | | |
|---------------------|---|--------------------------------------|------------|---------------|--------------------------|----------------|----------|--|--|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable Lii | mits (%) | | | |
| aboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | | | |
| EG020F: Dissolve | d Metals by ICP-MS (QCLot: 4859851) - continued | | | | | | | | | |
| ES2304011-005 | MW2 | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 95.7 | 70.0 | 130 | | | |
| G035F: Dissolve | d Mercury by FIMS (QCLot: 4859849) | | | | | | | | | |
| ES2304011-003 | QC301 | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 96.4 | 70.0 | 130 | | | |
| K040P: Fluoride | by PC Titrator (QCLot: 4859908) | | | | | | | | | |
| ES2303799-001 | Anonymous | EK040P: Fluoride | 16984-48-8 | 5 mg/L | 93.2 | 70.0 | 130 | | | |
| K055G: Ammoni | a as N by Discrete Analyser (QCLot: 4861210) | | | | | | | | | |
| ES2303838-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 1 mg/L | 118 | 70.0 | 130 | | | |
| K055G: Ammoni | a as N by Discrete Analyser (QCLot: 4861212) | | | | | | | | | |
| ES2304011-008 | MW6 | EK055G: Ammonia as N | 7664-41-7 | 1 mg/L | 109 | 70.0 | 130 | | | |
| K057G: Nitrite a | s N by Discrete Analyser (QCLot: 4860146) | | | | | | | | | |
| ES2303855-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.5 mg/L | # Not | 70.0 | 130 | | | |
| | | | | | Determined | | | | | |
| K059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCLot: 486 | 1209) | | | | | | | | |
| ES2303838-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 117 | 70.0 | 130 | | | |
| K059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCLot: 486 | 1211) | | | | | | | | |
| ES2304011-008 | MW6 | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 93.2 | 70.0 | 130 | | | |
| K059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCLot: 486 | 4095) | | | | | | | | |
| ME2300270-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 99.6 | 70.0 | 130 | | | |
| K061G: Total Kje | Idahl Nitrogen By Discrete Analyser (QCLot: 4861208) | | | | | | | | | |
| ES2304011-004 | MW1 | EK061G: Total Kjeldahl Nitrogen as N | | 25 mg/L | 101 | 70.0 | 130 | | | |
| K061G: Total Kje | Idahl Nitrogen By Discrete Analyser (QCLot: 4864092) | | | | | | | | | |
| ES2304009-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 5 mg/L | 100 | 70.0 | 130 | | | |
| K067G: Total Pho | osphorus as P by Discrete Analyser (QCLot: 4861207) | | | | | | | | | |
| ES2304011-004 | MW1 | EK067G: Total Phosphorus as P | | 5 mg/L | 99.5 | 70.0 | 130 | | | |
| K067G: Total Pho | osphorus as P by Discrete Analyser (QCLot: 4864091) | | | | | | | | | |
| ES2304009-001 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 95.6 | 70.0 | 130 | | | |
| K071G: Reactive | Phosphorus as P by discrete analyser (QCLot: 4860147 |) | | | | | | | | |
| ES2303957-001 | Anonymous | EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.5 mg/L | 93.3 | 70.0 | 130 | | | |
| K071G: Reactive | Phosphorus as P by discrete analyser (QCLot: 4860152 | | | | | | | | | |
| ES2304011-008 | MW6 | EK071G: Reactive Phosphorus as P | 14265-44-2 | 0.5 mg/L | 98.1 | 70.0 | 130 | | | |
| P080/071: Total F | Petroleum Hydrocarbons (QCLot: 4860105) | | | | | | | | | |
| ES2303866-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 80.0 | 70.0 | 130 | | | |
| P080/071: Total F | Recoverable Hydrocarbons - NEPM 2013 Fractions (QCL | ot: 4860105) | | | | | | | | |

Page : 11 of 11 Work Order : ES2304011



| Sub-Matrix: WATER | | Matrix Spike (MS) Report | | | | | |
|----------------------|---|----------------------------|------------|---------------|------------------|------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP080/071: Total | Recoverable Hydrocarbons - NEPM 2013 Fractions (QCL | ot: 4860105) - continued | | | | | |
| ES2303866-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 81.9 | 70.0 | 130 |
| EP080: BTEXN (C | QCLot: 4860105) | | | | | | |
| ES2303866-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 81.6 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 83.9 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 87.4 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 82.4 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 86.5 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 87.2 | 70.0 | 130 |

Chain of Custody Documentation

ABN 89 132 231 380 Senversa Pty Ltd Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project Job Number: specifications were used during the collection of these samples: Relinquished By: Lab ID roject Manager mail Report To: oject Name: mpled By: me/Signature: ne/Signature: | Time: | Date/Time: | Date/Tim Sample ID SW2 SW1 Bec Chapple Sample Information Matrix * 8 × Wetherill Park WME Emma Walsh Bec Chapple S20102 10/02/2023 10/02/2023 Date Address: Contact: Page: Phone/Mobile Quote No: Purchase Order: Date Turn Around Time Laboratory: Date: 10/2/23 Time: 9:15 AM Time AM AM ALS NSW Sample Receipt Method of Shipment (if applicable) Date/Time:)ate/Time: arrier / Reference # arrier / Reference rier / Reference #: Type / Code EN/103/21 0408038593, 0404011544 Container Information Standard 7 Days of 1 Sampler Name: **Total Bottles** 12 W-27 (TRH/BTEX/PAH/8 METALS/ PHENOLS) Bec Chapple NT-11 (TN, TP) Received by: EA015H (TDS) EA025H (TSS) Signature: EG005F (FE AND MN) P Analysis Required Sydney
Work Order Reference
ES2304342 Telephone: +61-2-8784 8555 **Environmental Division** Date: HOLD Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. Date: 10/1/23 roxig

Checked by: V = VOA Vial Hydochloric Acid (HCI) Preserved; VS = VOA Vial Sulphuric Preserved. VSA = Sulphuric Preserved Amber Glass; H = HCI Preserved Plastic; HS = HCI Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottle; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's odine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass

ved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2304342

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Khaleda Ataei

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : khaleda.ataei@alsqlobal.com

Telephone : 02 8252 0000 Telephone : + 61 2 8784 8555
Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 2

 Order number
 : --- Quote number
 : EM2020SENVER0016 (EN/103/21)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Site : ----

Sampler : BEC CHAPPLE

Dates

Date Samples Received : 10-Feb-2023 10:30 Issue Date : 10-Feb-2023 Client Requested Due : 16-Feb-2023 Scheduled Reporting Date : 16-Feb-2023

Date

Delivery Details

Mode of Delivery : Client Drop Off Security Seal : Intact.

No. of coolers/boxes : --- Temperature : 8.6' C - Ice present

Receipt Detail : FOAM ESKY No. of samples received / analysed : 2 / 2

General Comments

- This report contains the following information:
 - Sample Container(s)/Preservation Non-Compliances
 - Summary of Sample(s) and Requested Analysis
 - Proactive Holding Time Report
 - Requested Deliverables
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Sample(s) requiring volatile organic compound analysis received in airtight containers (ZHE).
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 10-Feb-2023 Issue Date

Page

2 of 2 ES2304342 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

| process necessatasks. Packages as the determin tasks, that are inclif no sampling default 00:00 on | ry for the execution may contain ad ation of moisture uded in the package. Itime is provided, the date of sampling date wi | ditional analyses, such content and preparation the sampling time will g. If no sampling date II be assumed by the ckets without a time | WATER - EA015H Total Dissolved Solids - Standard Level | WATER - EA025H Suspended Solids - Standard Level | WATER - EG005F Dissolved Metals by ICPAES | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - W-27 TRH/BTEXN/PAH/Phenols/8 Metals |
|--|--|---|---|---|--|--|--|
| ES2304342-001 | 10-Feb-2023 00:00 | SW1 | ✓ | ✓ | 1 | ✓ | ✓ |
| ES2304342-002 | 10-Feb-2023 00:00 | sw2 | ✓ | ✓ | ✓ | ✓ | ✓ |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

BEC CHAPPLE

| BEO OTIAL I EE | | |
|---|-------|---------------------------------|
| *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
| *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| EMMA WALSH | | |
| *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
| *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Tax Invoice (INV) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ENMRG (ENMRG) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| SUPPLIER ACCOUNTS | | |
| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
| | | u |
| | | |



CERTIFICATE OF ANALYSIS

Work Order : **ES2304342**

: SENVERSA PTY LTD

Contact : EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number : ---C-O-C number : ----

Client

Sampler · BEC CHAPPLE

Site : ---

Quote number : EN/103/21

No. of samples received : 2
No. of samples analysed : 2

Page : 1 of 6

Laboratory : Environmental Division Sydney

Contact : Khaleda Ataei

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : + 61 2 8784 8555

Date Samples Received : 10-Feb-2023 10:30

Date Analysis Commenced : 13-Feb-2023

Issue Date : 16-Feb-2023 16:15



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Edwandy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 6

Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.

Page : 3 of 6
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | sw2 | | |
|--|---------------------|--------|-----------------|-------------------|-------------------|------|--|
| | | Sampli | ing date / time | 10-Feb-2023 00:00 | 10-Feb-2023 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2304342-001 | ES2304342-002 | | |
| | | | | Result | Result | | |
| EA015: Total Dissolved Solids dried at | t 180 ± 5 °C | | | | | | |
| Total Dissolved Solids @180°C | | 10 | mg/L | 240 | 352 | | |
| EA025: Total Suspended Solids dried | at 104 ± 2°C | | | | | | |
| Suspended Solids (SS) | | 5 | mg/L | 86 | 69 | | |
| EG005(ED093)F: Dissolved Metals by | ICP-AES | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 0.06 | 0.06 | | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 0.01 | <0.01 | | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.001 | <0.001 | | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | | |
| Chromium | 7440-47-3 | 0.001 | mg/L | 0.002 | 0.002 | | |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.006 | 0.003 | | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | | |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.001 | 0.001 | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | | |
| EK059G: Nitrite plus Nitrate as N (NO | x) by Discrete Ana | lyser | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.36 | 0.50 | | |
| EK061G: Total Kjeldahl Nitrogen By D | iscrete Analyser | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.3 | 1.0 | | |
| EK062G: Total Nitrogen as N (TKN + N | IOx) by Discrete An | alyser | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 0.7 | 1.5 | | |
| EK067G: Total Phosphorus as P by Di | screte Analyser | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.06 | 0.19 | | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | <2.0 | <2.0 | | |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | <1.0 | <1.0 | | |

Page : 4 of 6
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | sw2 | | |
|---|--------------------|------------|----------------|-------------------|-------------------|------|--|
| | | Samplii | ng date / time | 10-Feb-2023 00:00 | 10-Feb-2023 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2304342-001 | ES2304342-002 | | |
| | | | | Result | Result | | |
| EP075(SIM)A: Phenolic Compounds | - Continued | | | | | | |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | <1.0 | <1.0 | | |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | <2.0 | <2.0 | | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | | |
| ^ Sum of polycyclic aromatic hydrocarbo | ons | 0.5 | μg/L | <0.5 | <0.5 | | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | | |
| EP080/071: Total Petroleum Hydroca | rbons | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | | |
| C15 - C28 Fraction | | 100 | μg/L | <100 | <100 | | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | <50 | | |
| EP080/071: Total Recoverable Hydro | carbons - NEPM 201 | 3 Fraction | ns | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | | |
| ^ C6 - C10 Fraction minus BTEX | C6_C10-BTEX | 20 | μg/L | <20 | <20 | | |
| (F1) | | | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | | |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | <100 | | |

Page : 5 of 6
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | Sample ID | | | SW1 | sw2 | | |
|---|-------------------|-----------|----------------|-------------------|-------------------|------|--|
| | | Sampli | ng date / time | 10-Feb-2023 00:00 | 10-Feb-2023 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2304342-001 | ES2304342-002 | | |
| | | | | Result | Result | | |
| EP080/071: Total Recoverable Hydroca | arbons - NEPM 201 | 3 Fractio | ns - Continued | | | | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | <100 | | |
| ^ >C10 - C16 Fraction minus Naphthalene (F2) | | 100 | μg/L | <100 | <100 | | |
| EP080: BTEXN | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | | |
| EP075(SIM)S: Phenolic Compound Su | rrogates | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 30.1 | 26.2 | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 59.0 | 53.2 | | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 55.7 | 58.8 | | |
| EP075(SIM)T: PAH Surrogates | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 65.1 | 62.5 | | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 74.7 | 79.4 | | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 68.6 | 85.7 | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 132 | 117 | | |
| Toluene-D8 | 2037-26-5 | 2 | % | 124 | 113 | | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 120 | 106 | | |

Page : 6 of 6
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME

ALS

Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery | Limits (%) | | | | | | |
|---|------------|----------|------------|--|--|--|--|--|--|
| Compound | CAS Number | Low | High | | | | | | |
| EP075(SIM)S: Phenolic Compound Surrogates | | | | | | | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 | | | | | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 | | | | | | |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 | | | | | | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 | | | | | | |
| Anthracene-d10 | 1719-06-8 | 27 | 113 | | | | | | |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 | | | | | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 71 | 137 | | | | | | |
| Toluene-D8 | 2037-26-5 | 79 | 131 | | | | | | |
| 4-Bromofluorobenzene | 460-00-4 | 70 | 128 | | | | | | |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2304342** Page : 1 of 7

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

 Contact
 : EMMA WALSH
 Telephone
 : + 61 2 8784 8555

 Project
 : S20102 Wetherill Park WME
 Date Samples Received
 : 10-Feb-2023

 Site
 : --- Issue Date
 : 16-Feb-2023

Site :---- Issue Date
Sampler : BEC CHAPPLE No. of samples received

Sampler : BEC CHAPPLE No. of samples received : 2
Order number : ---- No. of samples analysed : 2

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- NO Matrix Spike outliers occur.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

NO Analysis Holding Time Outliers exist.

Outliers : Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 7
Work Order : ES2304342

 Client
 : SENVERSA PTY LTD

 Project
 : S20102 Wetherill Park WME



Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | Count Rate (%) Qua | | e (%) | Quality Control Specification | |
|-----------------------------|--------------------|---------|--------|-------------------------------|--------------------------------|
| Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | |
| PAH/Phenols (GC/MS - SIM) | 0 | 2 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 5 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | |
| PAH/Phenols (GC/MS - SIM) | 0 | 2 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 5 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

| Matrix: WATER | | | | | Evaluation | : × = Holding time | breach ; ✓ = Withi | n holding time. |
|---|--------|-------------|----------------|------------------------|------------|--------------------|--------------------|-----------------|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA015H) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 14-Feb-2023 | 17-Feb-2023 | ✓ |
| EA025: Total Suspended Solids dried at 104 ± 2°C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA025H) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 14-Feb-2023 | 17-Feb-2023 | ✓ |
| EG005(ED093)F: Dissolved Metals by ICP-AES | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG005F) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 16-Feb-2023 | 09-Aug-2023 | ✓ |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 15-Feb-2023 | 09-Aug-2023 | ✓ |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 16-Feb-2023 | 10-Mar-2023 | ✓ |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Ana | alyser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | | | | 15-Feb-2023 | 10-Mar-2023 | ✓ |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 14-Feb-2023 | 10-Mar-2023 | ✓ | 15-Feb-2023 | 10-Mar-2023 | ✓ |

Page : 3 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: **WATER**Evaluation: × = Holding time breach; ✓ = Within holding time.

| WAILK | | | | | Lvaldatioi | i. • - Holding time | Dicacii, • - with | ir noluling till |
|---|-------------------|-------------|----------------|-------------------------|------------|---------------------|-------------------|------------------|
| Method | | Sample Date | Ex | ktraction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EK067G: Total Phosphorus as P by Discrete Analy | yser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK067G) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 14-Feb-2023 | 10-Mar-2023 | ✓ | 15-Feb-2023 | 10-Mar-2023 | ✓ |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 13-Feb-2023 | 17-Feb-2023 | ✓ | 14-Feb-2023 | 25-Mar-2023 | ✓ |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons | s | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 13-Feb-2023 | 17-Feb-2023 | ✓ | 14-Feb-2023 | 25-Mar-2023 | ✓ |
| EP080/071: Total Petroleum Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 13-Feb-2023 | 17-Feb-2023 | ✓ | 14-Feb-2023 | 25-Mar-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| SW1 | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | ✓ | 14-Feb-2023 | 24-Feb-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| sw2 | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | ✓ | 15-Feb-2023 | 24-Feb-2023 | ✓ |
| EP080/071: Total Recoverable Hydrocarbons - NEF | PM 2013 Fractions | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| SW1, | sw2 | 10-Feb-2023 | 13-Feb-2023 | 17-Feb-2023 | ✓ | 14-Feb-2023 | 25-Mar-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | 40 5 1 0000 | 44 5 1 0000 | 04 5-6 0000 | | 44 5 1 0000 | 04 5-6 0000 | |
| SW1 | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | ✓ | 14-Feb-2023 | 24-Feb-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | | 15-Feb-2023 | 24-Feb-2023 | |
| sw2 | | 10-Feb-2023 | 14-Feb-2023 | 24-160-2023 | ✓ | 15-reb-2023 | 24-Feb-2023 | ✓ |
| EP080: BTEXN | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | 40 = 1 | 44 5 1 0000 | 04 5-6 0000 | | 44.5.1.0000 | 04 5-6 0000 | |
| SW1 | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | ✓ | 14-Feb-2023 | 24-Feb-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | 40 F-h 0000 | 44 Fab 2000 | 24-Feb-2023 | | 45 Feb 2000 | 24-Feb-2023 | |
| sw2 | | 10-Feb-2023 | 14-Feb-2023 | 24-Feb-2023 | √ | 15-Feb-2023 | 24-Feb-2023 | ✓ |

Page : 4 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

| Quality Control Sample Type | | 0 | ount | | Rate (%) | | Quality Control Specification |
|---|------------|------|---------|--------|----------|------------|--------------------------------|
| Analytical Methods | Method | oc - | Regular | Actual | Expected | Evaluation | quality control opcomeditori |
| aboratory Duplicates (DUP) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 2 | 19 | 10.53 | 10.00 | | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 2 | 50.00 | 10.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 20 | 10.00 | 10.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 16 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| AH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 2 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Dissolved Solids (High Level) | EA015H | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 17 | 11.76 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Phosphorus as P By Discrete Analyser | EK067G | 2 | 16 | 12.50 | 10.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| RH - Semivolatile Fraction | EP071 | 0 | 5 | 0.00 | 10.00 | se | NEPM 2013 B3 & ALS QC Standard |
| RH Volatiles/BTEX | EP080 | 2 | 17 | 11.76 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| aboratory Control Samples (LCS) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 19 | 5.26 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| issolved Metals by ICP-AES | EG005F | 1 | 2 | 50.00 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| issolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| litrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 16 | 6.25 | 5.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 2 | 50.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 3 | 20 | 15.00 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Dissolved Solids (High Level) | EA015H | 3 | 20 | 15.00 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 17 | 17.65 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Phosphorus as P By Discrete Analyser | EK067G | 3 | 16 | 18.75 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| RH - Semivolatile Fraction | EP071 | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| RH Volatiles/BTEX | EP080 | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| issolved Mercury by FIMS | EG035F | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| issolved Metals by ICP-AES | EG005F | 1 | 2 | 50.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| litrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 2 | 50.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| suspended Solids (High Level) | EA025H | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Dissolved Solids (High Level) | EA015H | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| otal Phosphorus as P By Discrete Analyser | EK067G | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| RH - Semivolatile Fraction | EP071 | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| RH Volatiles/BTEX | EP080 | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |

Page : 5 of 7 Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: * = Quality Control frequency not within specification; * = Quality Control frequency within specification. Quality Control Sample Type Count Rate (%) Quality Control Specification Evaluation Method QC Analytical Methods Regular Actual Expected Matrix Spikes (MS) - Continued Dissolved Mercury by FIMS 19 5.26 5.00 NEPM 2013 B3 & ALS QC Standard EG035F 1 Dissolved Metals by ICP-AES 1 2 NEPM 2013 B3 & ALS QC Standard 50.00 5.00 1 EG005F Dissolved Metals by ICP-MS - Suite A 1 20 EG020A-F 5.00 5.00 1 NEPM 2013 B3 & ALS QC Standard Nitrite and Nitrate as N (NOx) by Discrete Analyser 1 16 6.25 5.00 NEPM 2013 B3 & ALS QC Standard EK059G 1 2 PAH/Phenols (GC/MS - SIM) 0 EP075(SIM) 0.00 5.00 NEPM 2013 B3 & ALS QC Standard × Total Kjeldahl Nitrogen as N By Discrete Analyser 1 17 5.88 NEPM 2013 B3 & ALS QC Standard 5.00 EK061G 1 Total Phosphorus as P By Discrete Analyser 1 16 6.25 5.00 1 NEPM 2013 B3 & ALS QC Standard EK067G TRH - Semivolatile Fraction 0 5 NEPM 2013 B3 & ALS QC Standard 0.00 5.00 EP071 × TRH Volatiles/BTEX 1 17 NEPM 2013 B3 & ALS QC Standard EP080 5.88 5.00 1

Page : 6 of 7 Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|---|----------|--------|---|
| Total Dissolved Solids (High Level) | EA015H | WATER | In house: Referenced to APHA 2540C. A gravimetric procedure that determines the amount of 'filterable' residue in an aqueous sample. A well-mixed sample is filtered through a glass fibre filter (1.2um). The filtrate is evaporated to dryness and dried to constant weight at 180+/-5C. This method is compliant with NEPM Schedule B(3) |
| Suspended Solids (High Level) | EA025H | WATER | In house: Referenced to APHA 2540D. A gravimetric procedure employed to determine the amount of 'non-filterable' residue in a aqueous sample. The prescribed GFC (1.2um) filter is rinsed with deionised water, oven dried and weighed prior to analysis. A well-mixed sample is filtered through a glass fibre filter (1.2um). The residue on the filter paper is dried at 104+/-2C. This method is compliant with NEPM Schedule B(3) |
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered samples, emitting a characteristic spectrum which is compared against matrix matched standards. This method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |

Page : 7 of 7 Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Analytical Methods | Method | Matrix | Method Descriptions |
|--------------------------------------|-----------------------|-----------------|---|
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode |
| | | | and quantification is by comparison against an established 5 point calibration curve. This method is compliant |
| | | | with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary |
| | | | GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a |
| | | | sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This |
| | | | method is compliant with the QC requirements of NEPM Schedule B(3) |
| | | | |
| Preparation Methods | Method | Matrix | Method Descriptions |
| Preparation Methods TKN/TP Digestion | Method EK061/EK067 | Matrix WATER | Method Descriptions In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| | | | |
| | | | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated |



QUALITY CONTROL REPORT

Work Order : **ES2304342** Page : 1 of 7

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Khaleda Ataei

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : + 61 2 8784 8555

Project : \$20102 Wetherill Park WME Date Samples Received : 10-Feb-2023

Order number Date Analysis Commenced : 13-Feb-2023

C-O-C number ---- Issue Date · 16-Feb-2023

Sampler ; BEC CHAPPLE

Site · ----

Quote number : EN/103/21

No. of samples received : 2

No. of samples analysed : 2

Accreditation No. 825
Accredited for compliance with ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Edwandy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory L | Duplicate (DUP) Report | | |
|----------------------|-------------------------|---------------------------------------|------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | ssolved Metals by ICP-A | AES (QC Lot: 4869828) | | | | | | | |
| ES2304342-002 | sw2 | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 0.06 | 0.06 | 0.0 | No Limit |
| EA015: Total Dissol | ved Solids dried at 180 | ± 5 °C (QC Lot: 4870476) | | | | | | | |
| ES2304252-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 4550 | 4510 | 0.8 | 0% - 20% |
| ES2304358-003 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 38100000 μg/L | 38800 | 2.0 | 0% - 20% |
| EA025: Total Suspe | nded Solids dried at 10 | 4 ± 2°C (QC Lot: 4870477) | | | | | | | |
| ES2304252-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 11 | 14 | 26.3 | No Limit |
| ES2304358-003 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 270000 μg/L | 262 | 3.0 | 0% - 20% |
| EG020F: Dissolved | Metals by ICP-MS (QC | Lot: 4869827) | | | | | | | |
| ES2304488-011 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.003 | 0.003 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| ES2304342-002 | sw2 | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | 0.002 | 0.002 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 0.003 | 0.003 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |

Page : 3 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|-----------------------|--|----------------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG035F: Dissolved | Mercury by FIMS (Q | C Lot: 4869826) | | | | | | | |
| ES2304488-008 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | 0.0004 | 0.0004 | 0.0 | No Limit |
| ES2304342-002 | sw2 | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| EK059G: Nitrite plu | ıs Nitrate as N (NOx) | by Discrete Analyser (QC Lot: 4869687) | | | | | | | |
| ES2304232-003 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 1.25 | 1.22 | 2.9 | 0% - 20% |
| ES2304352-002 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK061G: Total Kjeld | dahl Nitrogen By Disc | crete Analyser (QC Lot: 4869684) | | | | | | | |
| ES2304342-002 | sw2 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.0 | 0.8 | 14.3 | No Limit |
| ES2304232-003 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 4.8 | 4.4 | 10.0 | 0% - 20% |
| EK067G: Total Phos | sphorus as P by Disc | rete Analyser (QC Lot: 4869683) | | | | | | | |
| ES2304342-002 | sw2 | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.19 | 0.17 | 7.2 | 0% - 50% |
| ES2304232-003 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 3.85 | 3.79 | 1.4 | 0% - 20% |
| EP080/071: Total Pe | etroleum Hydrocarbo | ns (QC Lot: 4868872) | | | | | | | |
| ES2304473-004 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2304473-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Re | ecoverable Hydrocar | bons - NEPM 2013 Fractions (QC Lot: 4868872) | | | | | | | |
| ES2304473-004 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2304473-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 4868872) | | | | | | | | |
| ES2304473-004 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| ES2304473-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 4 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|-----------------|---------|------|-------------------|---------------|------------------------------|------------|--------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 48 | 69828) | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 108 | 82.0 | 114 |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 91.1 | 81.0 | 113 |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C (QCLot: | 4870476) | | | | | | | |
| EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | <10 | 2000 mg/L | 94.6 | 87.0 | 109 |
| | | | | <10 | 293 mg/L | 99.5 | 75.2 | 126 |
| | | | | <10 | 2340 mg/L | 102 | 83.0 | 124 |
| EA025: Total Suspended Solids dried at 104 ± 2°C (QCLot | 4870477) | | | | | | | |
| EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | 150 mg/L | 95.3 | 83.0 | 129 |
| | | | | <5 | 1000 mg/L | 96.3 | 82.0 | 110 |
| | | | | <5 | 987 mg/L | 93.6 | 83.0 | 118 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 4869827) | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 89.6 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 89.5 | 84.0 | 110 |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 88.3 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 88.4 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 90.6 | 83.0 | 111 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 86.8 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 88.2 | 81.0 | 117 |
| EG035F: Dissolved Mercury by FIMS (QCLot: 4869826) | | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 95.2 | 83.0 | 105 |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analy | ser (QCLot: 48 | (69687) | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 106 | 91.0 | 113 |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser(Q0 | CL of: 4869684) | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 96.4 | 69.0 | 101 |
| 2.100.0. rotal rystal in ridingen as it | | | | <0.1 | 1 mg/L | 94.4 | 70.0 | 118 |
| | | | | <0.1 | 5 mg/L | 102 | 70.0 | 130 |
| EK067G: Total Phosphorus as P by Discrete Analyser(QC | Lot: 4869683) | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 93.5 | 71.3 | 126 |
| | | | | <0.01 | 0.442 mg/L | 90.4 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 98.7 | 71.3 | 126 |
| EP075(SIM)A: Phenolic Compounds (QCLot: 4866073) | | | | | | | | |
| EP075(SIM): Phenol | 108-95-2 | 1 | μg/L | <1.0 | 5 μg/L | 34.6 | 24.5 | 61.9 |
| EP075(SIM): 2-Chlorophenol | 95-57-8 | 1 | μg/L | <1.0 | 5 μg/L | 74.2 | 52.0 | 90.0 |
| EP075(SIM): 2-Methylphenol | 95-48-7 | 1 | μg/L | <1.0 | 5 μg/L | 66.3 | 51.0 | 91.0 |

Page : 5 of 7
Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | aboratory Control Spike (LCS) Report | |
|---|--------------------------|-----|-------|-------------------|----------------------|------------------------------|--------------------------------------|--------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| P075(SIM)A: Phenolic Compounds (QCLot: 48) | 66073) - continued | | | | | | | |
| P075(SIM): 3- & 4-Methylphenol | 1319-77-3 | 2 | μg/L | <2.0 | 10 μg/L | 62.1 | 44.0 | 88.0 |
| P075(SIM): 2-Nitrophenol | 88-75-5 | 1 | μg/L | <1.0 | 5 μg/L | 72.5 | 48.0 | 100 |
| P075(SIM): 2.4-Dimethylphenol | 105-67-9 | 1 | μg/L | <1.0 | 5 μg/L | 67.3 | 49.0 | 99.0 |
| P075(SIM): 2.4-Dichlorophenol | 120-83-2 | 1 | μg/L | <1.0 | 5 μg/L | 72.9 | 53.0 | 105 |
| P075(SIM): 2.6-Dichlorophenol | 87-65-0 | 1 | μg/L | <1.0 | 5 μg/L | 69.2 | 57.0 | 105 |
| P075(SIM): 4-Chloro-3-methylphenol | 59-50-7 | 1 | μg/L | <1.0 | 5 μg/L | 71.1 | 53.0 | 99.0 |
| P075(SIM): 2.4.6-Trichlorophenol | 88-06-2 | 1 | μg/L | <1.0 | 5 μg/L | 67.4 | 50.0 | 106 |
| P075(SIM): 2.4.5-Trichlorophenol | 95-95-4 | 1 | μg/L | <1.0 | 5 μg/L | 69.7 | 51.0 | 105 |
| P075(SIM): Pentachlorophenol | 87-86-5 | 2 | μg/L | <2.0 | 10 μg/L | 43.4 | 10.0 | 95.0 |
| P075(SIM)B: Polynuclear Aromatic Hydrocarbo | ns (QCLot: 4866073) | | | | | | | |
| P075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 72.1 | 50.0 | 94.0 |
| P075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 84.9 | 63.6 | 114 |
| P075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 75.1 | 62.2 | 113 |
| P075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 72.2 | 63.9 | 115 |
| P075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 77.5 | 62.6 | 116 |
| P075(SIM): Anthracene | 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 89.5 | 64.3 | 116 |
| P075(SIM): Fluoranthene | 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 71.1 | 63.6 | 118 |
| P075(SIM): Pyrene | 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 79.4 | 63.1 | 118 |
| P075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 66.4 | 64.1 | 117 |
| P075(SIM): Chrysene | 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 71.5 | 62.5 | 116 |
| P075(SIM): Benzo(b+j)fluoranthene | 205-99-2 | 1 | μg/L | <1.0 | 5 μg/L | 70.4 | 61.7 | 119 |
| | 205-82-3 | | | | | | | |
| P075(SIM): Benzo(k)fluoranthene | 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 79.6 | 63.0 | 115 |
| P075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 69.2 | 63.3 | 117 |
| P075(SIM): Indeno(1.2.3.cd)pyrene | 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 66.2 | 59.9 | 118 |
| P075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 66.5 | 61.2 | 117 |
| P075(SIM): Benzo(g.h.i)perylene | 191-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 68.7 | 59.1 | 118 |
| P080/071: Total Petroleum Hydrocarbons (QCI | _ot: 4866074) | | | | | | | |
| P071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 57.2 | 53.7 | 97.0 |
| P071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 81.3 | 63.3 | 107 |
| P071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 87.3 | 58.3 | 120 |
| P080/071: Total Petroleum Hydrocarbons (QCI | ot: 4868872) | | | | | | | |
| P080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 95.2 | 75.0 | 127 |
| P080/071: Total Recoverable Hydrocarbons - N | EDM 2013 Fractions (OCL) | | , F-5 | - | FU | | | |
| P071: >C10 - C16 Fraction | EPW 2013 Fractions (QCL) | 100 | μg/L | <100 | 500 μg/L | 62.9 | 53.9 | 95.5 |
| P071: >C10 - C16 Fraction P071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 87.0 | 57.8 | 110 |
| | | 100 | μg/L | <100 | 700 μg/L 300 μg/L | 86.2 | 50.5 | 115 |
| P071: >C34 - C40 Fraction P080/071: Total Recoverable Hydrocarbons - N | | | ру/с | 100 | 300 μg/L | 00.2 | 30.3 | 113 |

Page : 6 of 7 Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|--|-----------------------|--------------------|---------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP080/071: Total Recoverable Hydrocarbons - NEPN | 1 2013 Fractions (QCI | Lot: 4868872) - co | ntinued | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 99.3 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 4868872) | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 97.8 | 70.0 | 122 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 87.9 | 69.0 | 123 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 86.4 | 70.0 | 120 |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 83.2 | 69.0 | 121 |
| | 106-42-3 | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 90.0 | 72.0 | 122 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 96.0 | 70.0 | 120 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | | Ma | atrix Spike (MS) Report | | |
|----------------------|--|--------------------------------------|------------|---------------|-------------------------|--------------|-----------|
| | | | | Spike | SpikeRecovery(%) | Acceptable L | imits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG005(ED093)F: D | issolved Metals by ICP-AES (QCLot: 4869828) | | | | | | |
| ES2304342-001 | SW1 | EG005F: Manganese | 7439-96-5 | 1 mg/L | 108 | 70.0 | 130 |
| EG020F: Dissolve | d Metals by ICP-MS (QCLot: 4869827) | | | | | | |
| ES2304488-002 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 91.9 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 93.6 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 89.0 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 91.2 | 70.0 | 130 |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 87.3 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 90.6 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 91.3 | 70.0 | 130 |
| EG035F: Dissolve | d Mercury by FIMS (QCLot: 4869826) | | | | | | |
| ES2304342-001 | SW1 | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 89.4 | 70.0 | 130 |
| EK059G: Nitrite p | us Nitrate as N (NOx) by Discrete Analyser (QCLot: 486 | 9687) | | | | | |
| ES2304232-003 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 103 | 70.0 | 130 |
| EK061G: Total Kje | Idahl Nitrogen By Discrete Analyser (QCLot: 4869684) | | | | | | |
| ES2304233-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 5 mg/L | 82.4 | 70.0 | 130 |
| EK067G: Total Pho | osphorus as P by Discrete Analyser (QCLot: 4869683) | | | | | | |
| ES2304233-001 | Anonymous | EK067G: Total Phosphorus as P | | 10 mg/L | 92.5 | 70.0 | 130 |
| EP080/071: Total F | Petroleum Hydrocarbons (QCLot: 4868872) | | | | | | |
| ES2304473-001 | Anonymous | | | | | | |

Page : 7 of 7 Work Order : ES2304342

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Matrix Spike (MS) Report | | | | |
|----------------------|---|----------------------------|------------|--------------------------|------------------|--------------|-----------|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable I | imits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | |
| EP080/071: Total F | Petroleum Hydrocarbons (QCLot: 4868872) - continued | | | | | | | |
| ES2304473-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 108 | 70.0 | 130 | |
| EP080/071: Total F | Recoverable Hydrocarbons - NEPM 2013 Fractions (QCL | ot: 4868872) | | | | | | |
| ES2304473-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 112 | 70.0 | 130 | |
| EP080: BTEXN (Q | CLot: 4868872) | | | | | | | |
| ES2304473-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 85.3 | 70.0 | 130 | |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 93.5 | 70.0 | 130 | |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 100 | 70.0 | 130 | |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 98.8 | 70.0 | 130 | |
| | | | 106-42-3 | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 102 | 70.0 | 130 | |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 99.4 | 70.0 | 130 | |



CLIENT: SPANGJA

CHAIN OF CUSTODY

ALS Laboratory: please tick >

| | _HCRARSHEE = T.Carlion Shael Rimon > .0 1812 en .07 4772 (0000 E.M. Steren Townsonthi Cleriotest dom _HACK CHECORS 618.21 (Saight Blad Time, Nath Vyodon of et NEW 2010) on .02 (4255-3155 E. wellongongiócksglober com | yan och 1847 vilote kajastal som 34 om och 1916 Möllön globet som | WSW 28 | |
|-----|--|--|--------|-----|
| - 1 | FOR LABORATORY USE ONLY (Circle) | Jircle) | | |
| | Custody Seat Intact? | Yes | No | NA |
| | Free ice / frozen ice bricks present upon receipt? | Yes | No | N/A |
| 7 | Random Sample Temperature on Receipt: | | °C | |
| 7 | Other comment: | | | |
| 四 | ELINQUISHED BY: | RECEIVED BY: | | |
| A | дтеліме: | DATE/TIME: | 2000 | |

OF: DATE/TIME RECEIVED BY: COC SEQUENCE NUMBER (Circle)

COC 3 4 5 6

E 0 2 3 4 5 6 N (4)01x

Email Invoice to (will default to PM if no other addresses are listed): Supplieraccounts (a) Sequence .com. Email Reports to (will default to PM if no other addresses are listed): bec.chappe(c) servere, com. cun

2

14/8/23

2,45pm

CONTAINER INFORMATION

ANALYSIS REQUIRED including SUITES (NB. Suite Codes must be listed to attract suite price)

Additional Information

COMMENTS/SPECIAL HANDLING/STORAGE OR DISPOSAL

ALS USE ONLY

SAMPLE DETAILS MATRIX: Solid(S) Water(W)

LAB ID

SAMPLE ID

DATE / TIME

MATRIX

TYPE & PRESERVATIVE (refer to codes below)

BOTTLES W-26

W-2 (8M)

W-14 A CPAH/phenol)

NT-11

NT-8

W-18

S

MW3 MWZ

N

300

1

18/23

5

5

 $\times \times \times \times$

4

ナスと

300

0 S

500

SIN

XX

S

力無

TRIP Blank

Telephone: + 61-2-8784 8555

TRIP SPIKE

0

QC402

Q(50Z

4

2 00

00302

4

QC102 SW2 PROJECT MANAGER: Be C

Chapple rechowise

SAMPLER MOBILE: 0429 727968

RELINQUISHED BY

HWWh

CONTACT PH:

COUNTRY OF ORIGIN

(Standard TAT may be longer for some tests e.g., Ultra Trace Organics)

Non Standard or urgent TAT (List due date)

coc:

TURNAROUND REQUIREMENTS:

X Standard TAT (List due date)

EDD FORMAT (or default)

ORDER NUMBER:

SAMPLER:

Hanley

COC Emailed to ALS? (YES / NO)

PROJECT: WE RECTIVECT WETNETH POWN PROJECT NO.: 2010Z ALS QUOTE NO.:

PURCHASE ORDER NO.

Total dissolved
solids (TDS)
+ Suspended
Folids
(TDS)

Sydne) **Environmental Division**

Work Order Reference

 $\times \times \times \times$

VOA Vlai HCl Preserved; VB = VOA Vial Sodium Bisulphate Preserved: VS = VOA Vial Sulfuric Preserved: AV = Airfreight Unpre ed; S = Sodium Hydroxidis Preserved Plastic; AG = Amber Glass Unpreserved; AP - Airfreight Unpreserved Plastic
Sulfuric Preserved Amber Glass; H = HCI preserved Plastic; HS = HCI preserved Speciation bottle; SP = Sulfuric Preserved Plastic; F = Formaldehyde Preser
LI = LugoIs lodine Preserved Bottles; STT = Sterile Sodium Thiosulfate Preserved Bottles. ved Glass;

TOTAL



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2327328

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : BEC CHAPPLE Contact : Khaleda Ataei

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

 Telephone
 : --- Telephone
 : + 61 2 8784 8555

 Facsimile
 : --- Facsimile
 : +61-2-8784 8500

Project : 20102 REDIRECT WETHERILL PARK Page : 1 of 3

 Order number
 : --- Quote number
 : EM2020SENVER0016 (EN/103/21)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

 Site
 : ---

Sampler : Hayley Yellowlees

Dates

Date

Mode of Delivery : Client Drop Off Security Seal : Not Available

No. of coolers/boxes : 1 Temperature : 10.8'C,12.2'C,13.9'C - Ice

present

Receipt Detail : No. of samples received / analysed : 11 / 10

General Comments

Delivery Details

• This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 14-Aug-2023 Issue Date

Page

2 of 3 ES2327328 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

| process necessa tasks. Packages as the determina tasks, that are inclu- lf no sampling default 00:00 on | ry for the execution may contain addition of moisture uded in the package. Itime is provided, the date of sampling date with | the sampling time will g. If no sampling date II be assumed by the ckets without a time | Standard Level | WATER - EA025H Suspended Solids - Standard Level | WATER - NT-08 Total Nitrogen + NO2 + NO3 + NH3 + Total P | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - W-02 8 Metals | WATER - W-14A PAH/Phenols (SIM) | WATER - W-26 TRH/BTEXN/PAH/8 Metals |
|--|--|---|--|---|--|--|--------------------------|------------------------------------|-------------------------------------|
| | - | MW2 | + | | ✓ | | | | ∀ |
| ES2327328-002 ES2327328-003 | 14-Aug-2023 00:00 14-Aug-2023 00:00 | MW3 | +- | | ∀ | | | | ∀ |
| | ū | MW4 | - | | ∀ | | | | ∀ |
| ES2327328-004 | 14-Aug-2023 00:00 | | - | | ∀ | | | | ∀ |
| ES2327328-005 | 14-Aug-2023 00:00 | MW6 | | | V | | | - | V |
| ES2327328-006 | 14-Aug-2023 00:00 | SW1 | √ | ✓ | | ✓ | √ | ✓ | |
| ES2327328-007 | 14-Aug-2023 00:00 | SW2 | ✓ | ✓ | | ✓ | ✓ | ✓ | |
| ES2327328-008 | 14-Aug-2023 00:00 | QC102 | | | | | | | ✓ |
| Matrix: WATER Laboratory sample | Sampling date / time | Sample ID | (On Hold) WATER No analysis requested | WATER - EA005P pH (Auto Titrator) | WATER - EP080 BTEXN | WATER - W-18 TRH(C6 - C9)/BTEXN | | | |
| ES2327328-006 | 14-Aug-2023 00:00 | SW1 | | ✓ | | | | | |
| ES2327328-007 | 14-Aug-2023 00:00 | SW2 | | ✓ | | | | | |
| ES2327328-009 | 14-Aug-2023 00:00 | QC302 | 1 | | | | | | |
| ES2327328-010 | 31-Jul-2023 00:00 | QC402 TRIP SPIKE | | | ✓ | | | | |
| ES2327328-011 | 01-Aug-2023 00:00 | QC502 TRIP BLANK | | | | 1 | 1 | | |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Issue Date : 14-Aug-2023

Page

3 of 3 ES2327328 Amendment 0 Work Order Client : SENVERSA PTY LTD



Requested Deliverables

BEC CHAPPLE

- *AU Certificate of Analysis - NATA (COA) Email bec.chapple@senversa.com.au - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) Email bec.chapple@senversa.com.au - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) Email bec.chapple@senversa.com.au - A4 - AU Sample Receipt Notification - Environmental HT (SRN) Email bec.chapple@senversa.com.au - A4 - AU Tax Invoice (INV) Email bec.chapple@senversa.com.au - Chain of Custody (CoC) (COC) Email bec.chapple@senversa.com.au - EDI Format - ESDAT (ESDAT) Email bec.chapple@senversa.com.au

SUPPLIER ACCOUNTS

- A4 - AU Tax Invoice (INV) Email supplieraccounts@senversa.com.a



CERTIFICATE OF ANALYSIS

Page **Work Order** : ES2327328 : 1 of 10

Amendment : 2

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : BEC CHAPPLE Contact : Khaleda Ataei

Address Address : 277-289 Woodpark Road Smithfield NSW Australia 2164 : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone Telephone : + 61 2 8784 8555 : 20102 REDIRECT WETHERILL PARK **Date Samples Received** Project : 14-Aug-2023 17:45

Order number C-O-C number

Sampler : Hayley Yellowlees

Site

Quote number : EN/103/21

No. of samples received : 11 No. of samples analysed : 11 **Date Analysis Commenced** : 14-Aug-2023 : 23-Aug-2023 12:16 Accreditation No. 825

> Accredited for compliance with ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full

Issue Date

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with **Quality Review and Sample Receipt Notification.**

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Position Signatories Accreditation Category

Alex Rossi Organic Chemist Sydney Organics, Smithfield, NSW Inorganics Coordinator Sydney Inorganics, Smithfield, NSW Wisam Marassa

Page : 2 of 10

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project · 20102 REDIRECT WETHERILL PARK



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- EG020: LORs have been raised for some samples due to matrix interference (High sample salinity)
- Amendment (23/08/2023): This report has been amended and re-released to allow the reporting of additional analytical data, specifically method EG020F for samples 001-009.
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.

: 3 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK

| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|--|---------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 14-Aug-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2327328-001 | ES2327328-002 | ES2327328-003 | ES2327328-004 | ES2327328-005 |
| | | | | Result | Result | Result | Result | Result |
| EG020F: Dissolved Metals by ICP-M | S | 13 | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.008 | 0.004 | <0.010 | 0.007 | 0.002 |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0010 | <0.0001 | <0.0001 |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | <0.010 | <0.001 | <0.001 |
| Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | <0.010 | <0.001 | <0.001 |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.036 | 0.005 | 0.207 | 0.020 | 0.002 |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | <0.010 | <0.001 | <0.001 |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.045 | 0.009 | 0.122 | <0.005 | 0.006 |
| Manganese | 7439-96-5 | 0.001 | mg/L | 2.26 | 1.00 | 6.39 | 6.04 | 0.225 |
| Iron | 7439-89-6 | 0.05 | mg/L | 2.01 | 0.58 | 5.64 | 2.91 | 0.20 |
| EG035F: Dissolved Mercury by FIMS | S | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| EK055G: Ammonia as N by Discrete | Analyser | | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.49 | 0.52 | 0.29 | 0.32 | 0.09 |
| EK057G: Nitrite as N by Discrete Ar | | | | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| EK058G: Nitrate as N by Discrete A | | - | Ü | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | 0.02 | <0.01 | <0.01 | <0.01 | 0.18 |
| | | | 9/_ | | 0.01 | 0.01 | 0.0 . | VII.0 |
| EK059G: Nitrite plus Nitrate as N (N Nitrite + Nitrate as N | OX) by Discrete Ana | 0.01 | mg/L | 0.02 | <0.01 | <0.01 | <0.01 | 0.18 |
| | | 0.01 | IIIg/L | 0.02 | 10.01 | 40.01 | 10.01 | 0.10 |
| EK061G: Total Kjeldahl Nitrogen By Total Kjeldahl Nitrogen as N | | 0.1 | ma/l | 0.6 | 0.6 | 0.4 | 0.5 | 0.4 |
| | | | mg/L | 0.6 | 0.6 | 0.4 | 0.5 | 0.4 |
| EK062G: Total Nitrogen as N (TKN + | NOx) by Discrete Ar | | (I | | | | | • |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 0.6 | 0.6 | 0.4 | 0.5 | 0.6 |
| EK067G: Total Phosphorus as P by | Discrete Analyser | | | | | | | 1 |
| Total Phosphorus as P | | 0.01 | mg/L | 0.02 | 0.04 | 0.02 | 0.01 | 0.14 |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |

: 4 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|--|---------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| , | | Samplii | ng date / time | 14-Aug-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2327328-001 | ES2327328-002 | ES2327328-003 | ES2327328-004 | ES2327328-005 |
| | | | | Result | Result | Result | Result | Result |
| EP075(SIM)B: Polynuclear Aromatic F | lydrocarbons - Cont | inued | | | | | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| ^ Sum of polycyclic aromatic hydrocarbor | ns | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| EP080/071: Total Petroleum Hydrocar | bons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | <50 |
| C15 - C28 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | <50 |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | <50 | <50 | <50 | <50 |
| EP080/071: Total Recoverable Hydroc | arbons - NEPM 201 | 3 Fraction | าร | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |
| ^ C6 - C10 Fraction minus BTEX | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |
| (F1) | | | | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| ^ >C10 - C16 Fraction minus Naphthalene | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| (F2) | | | | | | | | |
| EP080: BTEXN Benzene | 74.40.0 | 1 | ug/l | <1 | <1 | <1 | <1 | <1 |
| Toluene | 71-43-2 108-88-3 | 2 | μg/L μg/L | <2 | <2 | <2 | <2 | <2 |
| Ethylbenzene | 100-41-4 | 2 | μg/L μg/L | <2 | <2 | <2 | <2 | <2 |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L μg/L | <2 | <2 | <2 | <2 | <2 |
| ortho-Xylene | 95-47-6 | 2 | μg/L μg/L | <2 | <2 | <2 | <2 | <2 |
| ^ Total Xylenes | 90-47-0 | 2 | μg/L μg/L | <2 | <2 | <2 | <2 | <2 |

: 5 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK

| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|--|------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 14-Aug-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2327328-001 | ES2327328-002 | ES2327328-003 | ES2327328-004 | ES2327328-005 |
| | | | | Result | Result | Result | Result | Result |
| EP080: BTEXN - Continued | | | | | | | | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | <1 | <1 |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | <5 | <5 |
| EP075(SIM)S: Phenolic Compound Surroga | ates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 22.9 | 22.8 | 26.0 | 20.8 | 21.5 |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 50.6 | 51.0 | 57.1 | 47.4 | 48.6 |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 42.6 | 40.1 | 48.2 | 42.4 | 38.5 |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 63.7 | 62.6 | 69.8 | 58.0 | 60.5 |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 78.5 | 75.9 | 82.2 | 72.3 | 72.0 |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 85.5 | 84.1 | 89.2 | 82.0 | 79.2 |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 121 | 117 | 110 | 116 | 97.2 |
| Toluene-D8 | 2037-26-5 | 2 | % | 110 | 113 | 116 | 112 | 99.9 |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 124 | 125 | 123 | 120 | 107 |

: 6 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK

| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | SW2 | QC102 | QC302 | QC402 |
|---|---------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|---------------------------------|
| (Matrix: WATER) | | Samnli | ng date / time | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | TRIP SPIKE 31-Jul-2023 00:00 |
| O man a mark | 0.4.0.4// | LOR | Unit | ES2327328-006 | ES2327328-007 | ES2327328-008 | ES2327328-009 | ES2327328-010 |
| Compound | CAS Number | LOR | Onn | | | | | |
| FACCED all by DC Tituates | | | | Result | Result | Result | Result | Result |
| EA005P: pH by PC Titrator pH Value | | 0.01 | pH Unit | 8.03 | 7.75 | | | |
| | | 0.01 | prionit | 0.03 | 7.75 | | | |
| EA015: Total Dissolved Solids dried Total Dissolved Solids @180°C | | 10 | mg/L | 316 | 105 | | l | |
| | | 10 | IIIg/L | 310 | 105 | | | |
| EA025: Total Suspended Solids drie | | _ | | | | | | |
| Suspended Solids (SS) | | 5 | mg/L | 238 | 39 | | | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.001 | <0.001 | <0.010 | <0.001 | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0010 | <0.0001 | |
| Chromium | 7440-47-3 | 0.001 | mg/L | 0.001 | <0.001 | <0.010 | <0.001 | |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.004 | 0.001 | <0.010 | <0.001 | |
| Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.205 | <0.001 | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | <0.010 | <0.001 | |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.005 | 0.038 | 0.074 | <0.005 | |
| Manganese | 7439-96-5 | 0.001 | mg/L | 0.016 | 0.007 | 6.57 | 0.038 | |
| Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | 6.04 | <0.05 | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0001 | <0.0001 | |
| EK059G: Nitrite plus Nitrate as N (N | Ox) by Discrete Ana | lyser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.68 | 0.62 | | | |
| EK061G: Total Kjeldahl Nitrogen By | Discrete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.7 | 0.7 | | | |
| EK062G: Total Nitrogen as N (TKN + | NOx) by Discrete Ar | alvser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 2.4 | 1.3 | | | |
| EK067G: Total Phosphorus as P by I | Discrete Analyser | 7 | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.35 | 0.09 | | | |
| EP075(SIM)A: Phenolic Compounds | 13 14 15 | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |

: 7 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | SW2 | QC102 | QC302 | QC402 TRIP SPIKE |
|--|---------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|---------------------|
| | | Sampli | ng date / time | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 31-Jul-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2327328-006 | ES2327328-007 | ES2327328-008 | ES2327328-009 | ES2327328-010 |
| | | | | Result | Result | Result | Result | Result |
| EP075(SIM)A: Phenolic Compound | s - Continued | | | | | | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| EP075(SIM)B: Polynuclear Aromatic | c Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | |
| ^ Sum of polycyclic aromatic hydrocarl | oons | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | |
| EP080/071: Total Petroleum Hydrod | carbons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | <20 | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| C15 - C28 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | <50 | <50 | <50 | |
| EP080/071: Total Recoverable Hydr | ocarbons - NEPM 201 | 3 Fraction | ns | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | <20 | |
| ^ C6 - C10 Fraction minus BTEX | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | <20 | |
| (F1) | | | | | | | | |

: 8 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK



| Sub-Matrix: WATER (Matrix: WATER) | | | SW1 | SW1 SW2 | | QC302 | QC402 TRIP SPIKE | |
|---|----------------------|------------|----------------|-------------------|-------------------|-------------------|---------------------|-------------------|
| | Sampling date / time | | | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 14-Aug-2023 00:00 | 31-Jul-2023 00:00 |
| Compound | CAS Number | LOR | Unit | ES2327328-006 | ES2327328-007 | ES2327328-008 | ES2327328-009 | ES2327328-010 |
| | | | | Result | Result | Result | Result | Result |
| EP080/071: Total Recoverable Hydroc | arbons - NEPM 201 | 3 Fraction | ns - Continued | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| ^ >C10 - C16 Fraction minus Naphthalene | | 100 | μg/L | <100 | <100 | <100 | <100 | |
| (F2) | | | | | | | | |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | <1 | 16 |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | <2 | 16 |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | <2 | 17 |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | <2 <2 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 <2 | | 19 |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | <2 | 37 |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | <1 | 86 |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | <5 | 17 |
| EP075(SIM)S: Phenolic Compound Su | rrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 23.9 | 23.1 | 22.4 | 19.8 | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 52.5 | 51.4 | 47.7 | 49.2 | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 50.5 | 54.1 | 42.3 | 40.1 | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 67.4 | 67.4 | 57.2 | 64.8 | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 86.2 | 85.2 | 71.5 | 71.2 | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 87.7 | 86.7 | 76.6 | 78.4 | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 99.7 | 111 | 114 | 112 | 98.0 |
| Toluene-D8 | 2037-26-5 | 2 | % | 105 | 110 | 108 | 110 | 116 |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 115 | 115 | 119 | 122 | 123 |

: 9 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

Project 20102 REDIRECT WETHERILL PARK



| Sub-Matrix: WATER | Sample ID | | | QC502 | | | |
|---|---|----|-------------------|------------|------|------|--|
| (Matrix: WATER) | | | | TRIP BLANK | | | |
| Sampling date / time | | | 01-Aug-2023 00:00 | | | | |
| Compound | CAS Number LOR Unit | | ES2327328-011 | | | | |
| | | | | Result | | | |
| EP080/071: Total Petroleum Hydroca | EP080/071: Total Petroleum Hydrocarbons | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions | | | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | | | |
| ^ C6 - C10 Fraction minus BTEX | C6_C10-BTEX | 20 | μg/L | <20 | | | |
| (F1) | | | | | | | |
| EP080: BTEXN | EP080: BTEXN | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | | | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | | | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | | | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | | | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | | | |
| ^ Total Xylenes | | 2 | μg/L | <2 | | | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | | | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 96.6 | | | |
| Toluene-D8 | 2037-26-5 | 2 | % | 115 | | | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 121 | | | |

: 10 of 10 : ES2327328 Amendment 2 Work Order : SENVERSA PTY LTD Client

20102 REDIRECT WETHERILL PARK Project

Surrogate Control Limits

| Sub-Matrix: WATER | Recovery Limits (%) | | | | |
|---|---------------------|-----|------|--|--|
| Compound | CAS Number | Low | High | | |
| EP075(SIM)S: Phenolic Compound Surrogates | ; | | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 | | |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 | | |
| EP075(SIM)T: PAH Surrogates | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 | | |
| Anthracene-d10 | 1719-06-8 | 27 | 113 | | |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 | | |
| Toluene-D8 | 2037-26-5 | 75 | 131 | | |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 | | |





QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2327328** Page : 1 of 9

Amendment : 2

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

 Contact
 : BEC CHAPPLE
 Telephone
 : + 61 2 8784 8555

 Project
 : 20102 REDIRECT WETHERILL PARK
 Date Samples Received
 : 14-Aug-2023

Site :--- Issue Date : 23-Aug-2023

Sampler : Hayley Yellowlees No. of samples received : 11
Order number : ---- No. of samples analysed : 11

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- NO Matrix Spike outliers occur.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | Count | | Rate (%) | | Quality Control Specification |
|---|-------|---------|----------|----------|--------------------------------|
| Method | QC | Regular | Actual | Expected | |
| Laboraton, Dunlington (DLID) | U | | | | |
| Laboratory Duplicates (DUP) PAH/Phenols (GC/MS - SIM) | 0 | 16 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 18 | 0.00 | | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | |
| PAH/Phenols (GC/MS - SIM) | 0 | 16 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | 0 | 18 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Matrix: WATER | | | | | Lvaluation | . ~ - Holding time | breach; ▼ = within | ir noluling time |
|---|--------------------------------|-------------|--------------------------|--------------------|------------|--------------------|--------------------|------------------|
| Method | | Sample Date | Extraction / Preparation | | | Analysis | | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EA005P: pH by PC Titrator | | | | | | | | |
| Clear Plastic Bottle - Natural (EA005-P) SW1, | SW2 | 14-Aug-2023 | | | | 14-Aug-2023 | 14-Aug-2023 | √ |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA015H) SW1, | SW2 | 14-Aug-2023 | | | | 17-Aug-2023 | 21-Aug-2023 | √ |
| EA025: Total Suspended Solids dried at 104 ± 2°C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA025H) SW1, | SW2 | 14-Aug-2023 | | | | 17-Aug-2023 | 21-Aug-2023 | ✓ |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) MW1, MW3, MW6, SW2, QC302 | MW2, MW4, SW1, QC102, | 14-Aug-2023 | **** | | | 16-Aug-2023 | 10-Feb-2024 | ✓ |

Page : 3 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Matrix: WATER | | | | | Evaluation | n: × = Holding time | breach ; ✓ = Withi | n holding time. |
|---|--------------------------------|-------------|----------------|------------------------|------------|---------------------|--------------------|-----------------|
| Method | | Sample Date | E) | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) MW1, MW3, MW6, SW2, | MW2, MW4, SW1, QC102, | 14-Aug-2023 | | | | 17-Aug-2023 | 11-Sep-2023 | ✓ |
| QC302 | QC 102, | | | | | | | |
| EK055G: Ammonia as N by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK055G) MW1, MW3, MW6 | MW2, MW4, | 14-Aug-2023 | | | | 17-Aug-2023 | 11-Sep-2023 | ✓ |
| EK057G: Nitrite as N by Discrete Analyser | | <u> </u> | | | | | | |
| Clear Plastic Bottle - Natural (EK057G) MW1, MW3, MW6 | MW2, MW4, | 14-Aug-2023 | | | | 16-Aug-2023 | 16-Aug-2023 | ✓ |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete An | alyser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) MW1, MW3, MW6, SW2 | MW2, MW4, SW1, | 14-Aug-2023 | | | | 17-Aug-2023 | 11-Sep-2023 | ✓ |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) MW1, MW3, MW6, SW2 | MW2, MW4, SW1, | 14-Aug-2023 | 16-Aug-2023 | 11-Sep-2023 | 1 | 17-Aug-2023 | 11-Sep-2023 | ✓ |
| EK067G: Total Phosphorus as P by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK067G) MW1, MW3, MW6, SW2 | MW2, MW4, SW1, | 14-Aug-2023 | 16-Aug-2023 | 11-Sep-2023 | ✓ | 17-Aug-2023 | 11-Sep-2023 | ✓ |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) SW1, | SW2 | 14-Aug-2023 | 16-Aug-2023 | 21-Aug-2023 | 1 | 17-Aug-2023 | 25-Sep-2023 | ✓ |

Page : 4 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sample Date Extraction / Preparation Date entracted Due for extraction Evaluation Date enalysed Due for analysis Evaluation | | | | | | | | | |
|---|--|------------------------|-------------|----------------|------------------------|------------|---------------------|--------------------|----------------|
| Container (Client Semple (Eff.) Entry (SIMI) Polynuclear Aromatic Hydrocarbons | Matrix: WATER | | | | | Evaluation | n: × = Holding time | breach ; ✓ = Withi | n holding time |
| ### PROPERTY SIMILE Polynuclear Aromatic Hydrocarbons #################################### | Method | | Sample Date | E) | traction / Preparation | | | Analysis | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) MW1, MW3, MW4, MW6, SW2 SW1. CC302 14-Aug-2023 16-Aug-2023 14-Aug-2023 16-Aug-2023 11-Aug-2023 18-Aug-2023 | Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| MW1, MW2, MW4, MW6, SW1, MW4, MW6, SW1, MW4, MW6, SW1, MW6, SW1, MW6, SW1, MW6, SW1, MW6, SW2, MW6, | EP075(SIM)B: Polynuclear Aromatic Hydrocar | rbons | | | | | | | |
| MW1, MW2, MW4, MW6, SW1, MW4, MW6, SW1, MW6, SW1, MW6, SW1, MW6, SW1, MW6, SW2, MW6, | Amber Glass Bottle - Unpreserved (EP075(SIM | VI)) | | | | | | | |
| MW6, SW1, | MW1, | MW2, | 14-Aug-2023 | 16-Aug-2023 | 21-Aug-2023 | 1 | 17-Aug-2023 | 25-Sep-2023 | 1 |
| MW6, SW2 | MW3, | MW4, | | | | | | | |
| SW2 | MW6, | | | | | | | | |
| Amber Color Colo | SW2 | | | | | | | | |
| CC102, QC302 14-Aug-2023 16-Aug-2023 21-Aug-2023 21-Aug-2023 25-Sep-2023 | | VI)) | | | | | | | |
| Amber (Glass Bottle - Unpreserved (EP071) MW1, MW2, MW3, MW4, SW1, SW2, QC 102, QC 302 CC 302 CC 302 CC 102, CC 302 CC 102 - TRIP BLANK MW2, MW3, MW4, MW4, MW4, MW4, MW4, MW4, MW4, MW4 | | | 14-Aug-2023 | 16-Aug-2023 | 21-Aug-2023 | ✓ | 18-Aug-2023 | 25-Sep-2023 | ✓ |
| Amber (Glass Bottle - Unpreserved (EP071) MW1, MW2, MW3, MW4, SW1, SW2, QC 102, QC 302 CC 302 CC 302 CC 102, CC 302 CC 102 - TRIP BLANK MW2, MW3, MW4, MW4, MW4, MW4, MW4, MW4, MW4, MW4 | EP080/071: Total Petroleum Hydrocarbons | | | | | | | | |
| MW3, MW4, SW1, SW2, QC102, QC302 Amber VOC Vial - Sulfuric Acid (EP080) | Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| MW6, SW1, QC102, QC102, QC102, QC102, QC202 SW1, SW2, QC102, QC302 SW1, SW2, QC102, QC302 SW1, SW2, QC102, QC302 SW1, SW1, SW2, QC102, QC302 SW2, SW1, SW2, QC102, QC302 SW2, SW1, SW2, QC102, QC302 SW1, SW1, SW2, QC102, QC302 SW1, SW1, SW2, QC102, QC302 SW2, SW1, SW2, QC102, QC302 SW2, SW1, SW2, QC102, QC302 SW2, SW1, SW2, QC102, QC302 SW3, SW1, SW2, SW1, SW2, QC102, QC302 SW3, SW1, SW1, SW2, SW1, SW2, SW1, SW1, SW2, SW1, SW2, SW1, SW1, SW2, SW1, SW1, SW1, SW1, SW1, SW1, SW1, SW1 | MW1, | MW2, | 14-Aug-2023 | 16-Aug-2023 | 21-Aug-2023 | 1 | 17-Aug-2023 | 25-Sep-2023 | √ |
| SW2, QC102, QC502 - TRIP BLANK 01-Aug-2023 | MW3, | MW4, | | | | | | | |
| SW2, QC102, QC502 - TRIP BLANK 01-Aug-2023 | MW6, | SW1, | | | | | | | |
| QC302 | The state of the s | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK MW2, MW3, MW4, MW6, SW1, SW2, QC102, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2015 Fractions Amber VOC Vial - Sulfuric Acid (EP080) MW1, MW3, MW2, MW2, MW3, MW4, MW6, SW1, SW2, QC102, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2015 Fractions Amber Glass Bottle - Unpreserved (EP071) MW1, MW2, MW3, MW4, MW6, SW1, SW2, QC102, QC302 T1-Aug-2023 16-Aug-2023 21-Aug-2023 21-Aug-2023 25-Sep-2023 Amber VOC Vial - Sulfuric Acid (EP080) QC302 Amber VOC Vial - Sulfuric Acid (EP080) MW2, Amber OVC Vial - Sulfuric Acid (EP080) MW2, Amber VOC Vial - Sulfuric Acid (EP080) MW2, MW3, MW4, MW6, SW1, MW2, MW8, MW4, MW6, SW1, MW2, MW8, MW8, MW4, MW6, SW1, MW8, MW8 | I | 40.10- | | | | | | | |
| QC502 - TRIP BLANK | | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | 01-Aug-2023 | 14-Aug-2023 | 15-Aug-2023 | 1 | 14-Aug-2023 | 15-Aug-2023 | 1 |
| MW1, MW2, MW4, MW4, SW1, SW2, QC102, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) MW3, MW4, MW6, SW1, SW2, QC102, QC302 MW9, MW3, MW4, MW6, SW1, SW2, QC102, QC302 Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK Amber Glass Bottle - Unpreserved (EP080) MW1, MW2, MW3, MW4, MW4, MW6, SW1, SW1, MW2, MR9, MW2, MR9, MR9, MR9, MR9, MR9, MR9, MR9, MR9 | | | | | | | | | |
| MW6, SW1, QC102, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) | The state of the s | MW2, | 14-Aug-2023 | 17-Aug-2023 | 28-Aug-2023 | 1 | 17-Aug-2023 | 28-Aug-2023 | ✓ |
| MW6, SW1, QC102, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) | MW3. | MW4. | | | | | | | · |
| SW2, QC302 EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) MW1, MW6, SW1, QC302 Amber VOC Vial - Sulfuric Acid (EP080) MW2, MW3, MW4, MW6, SW1 - Sulfuric Acid (EP080) QC502 - TRIP BLANK Amber VOC Vial - Sulfuric Acid (EP080) MW1, MW2, MW3, MW4, MW4, MW4, MW4, MW4, MW4, MW6, SW1 - SW1, MW4, MW6, SW1, MW4, SW1, MW4, SW1, MW4, SW1, MW4, SW1, MW6, SW1, SW1, WW4, SW1, MW6, SW1, WW4, SW1, MW6, SW1, WM6, S | · · · · · · · · · · · · · · · · · · · | , | | | | | | | |
| CG302 FP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions | · · · · · · · · · · · · · · · · · · · | • | | | | | | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) | · · · · · · · · · · · · · · · · · · · | ασ.σΞ, | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | - NEDM 2013 Fractions | | | | | | | |
| MW1, MW2, MW4, MW6, SW1, SW2, QC102, QC302 Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK Amber VOC Vial - Sulfuric Acid (EP080) MW2, MW2, MW4, MW6, SW1, SW2, QC502 - TRIP BLANK Amber VOC Vial - Sulfuric Acid (EP080) MW2, MW4, MW4, MW6, SW1, MW4, SW1, MW6, SW1, | - | FILE IN 2010 Fractions | | | | | | | |
| MW3, MW6, SW1, SW2, QC102, QC302 Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK MW2, MW2, MW2, MW4, MW4, MW4, MW6, SW1, MW3, MW4, SW1, SW1, MW4, SW1, SW1, | | MW2. | 14-Aug-2023 | 16-Aug-2023 | 21-Aug-2023 | / | 17-Aug-2023 | 25-Sep-2023 | 1 |
| MW6, SW1, SW2, QC102, QC302 Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK 01-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 ✓ Amber VOC Vial - Sulfuric Acid (EP080) MW2, 14-Aug-2023 17-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ MW3, MW4, MW4, SW1, SW1, NW2, NW3, NW4, | · · · · · · · · · · · · · · · · · · · | , | | | | _ | | · | • |
| SW2, QC302 QC102, COUNTY OF AUGUSTS COUNTY OF AUG-2023 15-Aug-2023 17-Aug-2023 17-Aug-2023 28-Aug-2023 √ 17-Aug-2023 28-Aug-2023 √ MW3, MW6, SW1, SW1 | · · · · · · · · · · · · · · · · · · · | | | | | | | | |
| QC302 Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK O1-Aug-2023 Amber VOC Vial - Sulfuric Acid (EP080) MW1, MW3, MW4, MW6, SW1, O1-Aug-2023 MW2, MW4, MW6, SW1, O1-Aug-2023 MW2, MW4, MW6, O1-Aug-2023 O1-Au | l ' | • | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK 14-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 ✓ Amber VOC Vial - Sulfuric Acid (EP080) MW2, 14-Aug-2023 17-Aug-2023 28-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ MW3, MW4, SW1, SW1, <td< td=""><td>l control of the cont</td><td>QO 102,</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<> | l control of the cont | QO 102, | | | | | | | |
| QC502 - TRIP BLANK 01-Aug-2023 14-Aug-2023 15-Aug-2023 15-Aug-2023 15-Aug-2023 ✓ Amber VOC Vial - Sulfuric Acid (EP080) MW2, 14-Aug-2023 17-Aug-2023 28-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ MW3, MW4, SW1, SW1, SW1, | | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) MW1, MW2, 11-Aug-2023 17-Aug-2023 28-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ MW3, MW4, SW1, SW1, 17-Aug-2023 28-Aug-2023 ✓ 17-Aug-2023 28-Aug-2023 ✓ | | | 01-Aug-2023 | 14-Aug-2023 | 15-Aug-2023 | 1 | 14-Aug-2023 | 15-Aug-2023 | 1 |
| MW1, MW2, 14-Aug-2023 17-Aug-2023 | | | 1 13 | | J. | _ | | <u>u</u> | _ |
| MW3, MW4, MW6, SW1, | · · · | MW2, | 14-Aug-2023 | 17-Aug-2023 | 28-Aug-2023 | 1 | 17-Aug-2023 | 28-Aug-2023 | 1 |
| MW6, SW1, | The state of the s | | | | | | | | |
| | · · | , | | | | | | | |
| O | The state of the s | | | | | | | | |
| QC302 | · · · · · · · · · · · · · · · · · · · | QO 102, | | | | | | | |

Page : 5 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Matrix: WATER Evaluation: ★ = Holding time breach; ✓ = \ | | | | | | | | |
|---|--------|----------------|--------------------|------------------------|---------------|------------------|-------------|---|
| Method | | Sample Date | Ex | traction / Preparation | | Analysis | | |
| Container / Client Sample ID(s) | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EP080: BTEXN | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) QC502 - TRIP BLANK | | 01-Aug-2023 | 14-Aug-2023 | 15-Aug-2023 | 1 | 14-Aug-2023 | 15-Aug-2023 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| MW1, | MW2, | 14-Aug-2023 | 17-Aug-2023 | 28-Aug-2023 | ✓ | 17-Aug-2023 | 28-Aug-2023 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | SW1, | | | | | | | |
| SW2, | QC102, | | | | | | | |
| QC302 | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| QC402 - TRIP SPIKE | | 31-Jul-2023 | 14-Aug-2023 | 14-Aug-2023 | ✓ | 14-Aug-2023 | 14-Aug-2023 | ✓ |

Page : 6 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: WATER

Evaluation: × = Quality Control frequency not within specification: √ = Quality Control frequency within specification.

| Matrix: WATER | | | | Evaluatio | n: × = Quality Co | ontrol frequency | not within specification ; ✓ = Quality Control frequency within specification |
|---|------------|----|---------|-----------|-------------------|------------------|---|
| Quality Control Sample Type | | С | ount | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 2 | 17 | 11.76 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 4 | 39 | 10.26 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 2 | 16 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 16 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| pH by Auto Titrator | EA005-P | 2 | 18 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 18 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 3 | 25 | 12.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | 1 | 1 | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 39 | 5.13 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 2 | 16 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| pH by Auto Titrator | EA005-P | 2 | 18 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 5 | 40 | 12.50 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 5 | 40 | 12.50 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 19 | 15.79 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 3 | 19 | 15.79 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 2 | 18 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 25 | 8.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | 1 | 1 | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 39 | 5.13 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 19 | 5.26 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 16 | 6.25 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 2 | 16 | 12.50 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 2 | 40 | 5.00 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 2 | 40 | 5.00 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |

Page : 7 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Matrix: WATER Evaluation: ▼ = Quality Control frequency not within specification; ✓ = Quality Control frequency within specification. | | | | | | | | | |
|--|------------|----|---------|--------|----------|------------|--------------------------------|--|--|
| Quality Control Sample Type | | Co | unt | | Rate (%) | | Quality Control Specification | | |
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | | | |
| Method Blanks (MB) - Continued | | | | | | | | | |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| TRH - Semivolatile Fraction | EP071 | 2 | 18 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| TRH Volatiles/BTEX | EP080 | 2 | 25 | 8.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Matrix Spikes (MS) | | | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 17 | 5.88 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 39 | 5.13 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 16 | 0.00 | 5.00 | x | NEPM 2013 B3 & ALS QC Standard | | |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |
| TRH - Semivolatile Fraction | EP071 | 0 | 18 | 0.00 | 5.00 | x | NEPM 2013 B3 & ALS QC Standard | | |
| TRH Volatiles/BTEX | EP080 | 2 | 25 | 8.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard | | |

Page : 8 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|---|----------|--------|---|
| pH by Auto Titrator | EA005-P | WATER | In house: Referenced to APHA 4500 H+ B. This procedure determines pH of water samples by automated ISE. This method is compliant with NEPM Schedule B(3) |
| Total Dissolved Solids (High Level) | EA015H | WATER | In house: Referenced to APHA 2540C. A gravimetric procedure that determines the amount of `filterable` residue in an aqueous sample. A well-mixed sample is filtered through a glass fibre filter (1.2um). The filtrate is evaporated to dryness and dried to constant weight at 180+/-5C. This method is compliant with NEPM Schedule B(3) |
| Suspended Solids (High Level) | EA025H | WATER | In house: Referenced to APHA 2540D. A gravimetric procedure employed to determine the amount of `non-filterable` residue in a aqueous sample. The prescribed GFC (1.2um) filter is rinsed with deionised water, oven dried and weighed prior to analysis. A well-mixed sample is filtered through a glass fibre filter (1.2um). The residue on the filter paper is dried at 104+/-2C. This method is compliant with NEPM Schedule B(3) |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Ammonia as N by Discrete analyser | EK055G | WATER | In house: Referenced to APHA 4500-NH3 G Ammonia is determined by direct colorimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrite as N by Discrete Analyser | EK057G | WATER | In house: Referenced to APHA 4500-NO2- B. Nitrite is determined by direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrate as N by Discrete Analyser | EK058G | WATER | In house: Referenced to APHA 4500-NO3- F. Nitrate is reduced to nitrite by way of a chemical reduction followed by quantification by Discrete Analyser. Nitrite is determined seperately by direct colourimetry and result for Nitrate calculated as the difference between the two results. This method is compliant with NEPM Schedule B(3) |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |

Page : 9 of 9

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Analytical Methods | Method | Matrix | Method Descriptions |
|---|-------------|--------|---|
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| Preparation Methods | Method | Matrix | Method Descriptions |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| Separatory Funnel Extraction of Liquids | ORG14 | WATER | In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated and concentrated for analysis. This method is compliant with NEPM Schedule B(3). ALS default excludes sediment which may be resident in the container. |
| Volatiles Water Preparation | ORG16-W | WATER | A 5 mL aliquot or 5 mL of a diluted sample is added to a 40 mL VOC vial for purging. |



QUALITY CONTROL REPORT

: ES2327328 Work Order Page : 1 of 11

: 2 Amendment

Client Laboratory : Environmental Division Sydney : SENVERSA PTY LTD

Contact : BEC CHAPPLE Contact : Khaleda Ataei

Address Address : 277-289 Woodpark Road Smithfield NSW Australia 2164 : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone Telephone : ----: + 61 2 8784 8555

Project Date Samples Received : 20102 REDIRECT WETHERILL PARK : 14-Aug-2023 Order number **Date Analysis Commenced** : 14-Aug-2023

Sampler : Hayley Yellowlees

Site

Quote number : EN/103/21

No. of samples received : 11 No. of samples analysed : 11

Accreditation No. 825 Accredited for compliance with

ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

Issue Date

· 23-Aug-2023

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

C-O-C number

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Alex Rossi Organic Chemist Sydney Organics, Smithfield, NSW Wisam Marassa Inorganics Coordinator Sydney Inorganics, Smithfield, NSW Page : 2 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory L | Duplicate (DUP) Report | | |
|----------------------|--------------------------------|---------------------------------------|------------|--------|---------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EA005P: pH by PC 1 | Fitrator (QC Lot: 5233621) | | | | | | | | |
| ES2327282-001 | Anonymous | EA005-P: pH Value | | 0.01 | pH Unit | 7.64 | 7.72 | 1.0 | 0% - 20% |
| ES2327333-005 | Anonymous | EA005-P: pH Value | | 0.01 | pH Unit | 7.64 | 7.67 | 0.4 | 0% - 20% |
| EA015: Total Dissol | ved Solids dried at 180 ± 5 °C | C (QC Lot: 5239101) | | | | | | | |
| ES2327006-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 160 | 157 | 1.9 | 0% - 50% |
| ES2327035-005 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 47200000 μg/L | 44600 | 5.5 | 0% - 20% |
| ES2327110-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 1680 | 1580 | 5.9 | 0% - 20% |
| EW2303543-002 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 530 | 537 | 1.3 | 0% - 20% |
| EA025: Total Suspe | nded Solids dried at 104 ± 2° | °C (QC Lot: 5239102) | | | | | | | |
| ES2327006-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | <5 | 0.0 | No Limit |
| ES2327035-005 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 30000 μg/L | 43 | 34.8 | No Limit |
| ES2327110-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 356 | 310 | 14.0 | 0% - 20% |
| EW2303543-002 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | <5 | 0.0 | No Limit |
| EG020F: Dissolved | Metals by ICP-MS (QC Lot: | 5236663) | | | | | | | |
| ES2327041-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | 0.001 | 0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 0.002 | 0.002 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | 0.022 | 0.022 | 0.0 | 0% - 20% |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 0.088 | 0.088 | 0.0 | 0% - 50% |
| | | EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | 0.18 | 0.18 | 0.0 | No Limit |
| ES2327081-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |

Page : 3 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|----------------------|--|------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG020F: Dissolved | Metals by ICP-MS (Q | C Lot: 5236663) - continued | | | | | | | |
| ES2327081-001 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.002 | 0.002 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | 0.002 | 0.002 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | 0.104 | 0.099 | 4.9 | 0% - 20% |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.006 | 0.005 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| | | EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EG020F: Dissolved | Metals by ICP-MS (Q | C Lot: 5236666) | | | | | | | |
| EW2303610-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | 0.060 | 0.060 | 1.7 | 0% - 20% |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| | | EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | 0.07 | 0.07 | 0.0 | No Limit |
| EW2303629-005 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | 0.050 | 0.051 | 2.2 | 0% - 20% |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| | | EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | 0.13 | 0.22 | 47.9 | No Limit |
| EG035F: Dissolved | Mercury by FIMS (Q | C Lot: 5236665) | | | | | | | |
| ES2327080-002 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| ES2327328-002 | MW2 | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| EK055G: Ammonia | as N by Discrete Ana | lyser (QC Lot: 5238030) | | | | | | | |
| ES2327281-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.02 | 0.02 | 0.0 | No Limit |
| ES2327328-003 | MW3 | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.29 | 0.29 | 0.0 | 0% - 20% |
| EK057G: Nitrite as | N by Discrete Analys | er (QC Lot: 5237562) | | | | | | | |
| ES2327328-003 | MW3 | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| ES2327281-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK059G: Nitrite plu | , | by Discrete Analyser (QC Lot: 5238031) | | | | | | | |
| ES2327281-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.09 | 0.09 | 0.0 | No Limit |
| ES2327328-003 | MW3 | EK059G: Nitrite + Nitrate as N EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| L02021020-000 | IVIVVO | ENUDSIG. Millille + Millialle as M | | 0.01 | mg/L | NO.01 | ~0.01 | 0.0 | INO LITTIL |

Page : 4 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | | | Laboratory L | Duplicate (DUP) Report | | |
|----------------------|-----------------------|---|--------------------|------|--------------|-----------------|------------------------|---------|----------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EK061G: Total Kjeld | ahl Nitrogen By Discr | rete Analyser (QC Lot: 5238027) | | | | | | | |
| ES2327281-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.9 | 0.9 | 0.0 | No Limit |
| ES2327328-002 | MW2 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.6 | 0.6 | 0.0 | No Limit |
| EK067G: Total Phos | phorus as P by Discr | ete Analyser (QC Lot: 5238026) | | | | | | | |
| ES2327281-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.04 | 0.04 | 0.0 | No Limit |
| ES2327328-002 | MW2 | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.04 | 0.04 | 0.0 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbon | s (QC Lot: 5233625) | | | | | | | |
| ES2327291-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbon | s (QC Lot: 5236018) | | | | | | | |
| ES2327093-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2327167-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Red | coverable Hydrocarb | ons - NEPM 2013 Fractions (QC Lot: 5233625) | | | | | | | |
| ES2327291-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Red | coverable Hydrocarb | ons - NEPM 2013 Fractions (QC Lot: 5236018) | | | | | | | |
| ES2327093-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2327167-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5233625) | 1 11 11 11 11 11 11 11 11 11 | | | | | | | |
| ES2327291-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| EP080: BTEXN (QC I | Lot: 5236018) | | | | | | | | |
| ES2327093-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | 0 | | -0 | -0 | 0.0 | NI - Limit |
| | | EP080: ortho-Xylene | 95-47-6 91-20-3 | 5 | μg/L | <2 <5 | <2 <5 | 0.0 | No Limit |
| ES2327167-001 | Anonymous | EP080: Naphthalene | 71-43-2 | 1 | μg/L μg/L | <5 <1 | <5 <1 | 0.0 | No Limit No Limit |
| L02021 101-001 | Anonymous | EP080: Benzene EP080: Toluene | 108-88-3 | 2 | μg/L μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Toluene EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | Li 000. Hieta- α para-λyierie | 106-42-3 | - | ₩9, - | | | 0.0 | THO EITH |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| 1 | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 5 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|------------|--------|---------|-------------------|---------------|------------------------------|------------|--------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EA005P: pH by PC Titrator (QCLot: 5233621) | | | | | | | | |
| EA005-P: pH Value | | | pH Unit | | 4 pH Unit | 99.8 | 98.8 | 101 |
| | | | | | 7 pH Unit | 99.8 | 99.2 | 101 |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C (QCLot: | 5239101) | | | | | | | |
| EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | <10 | 2000 mg/L | 101 | 87.0 | 109 |
| | | | | <10 | 293 mg/L | 102 | 75.2 | 126 |
| | | | | <10 | 2380 mg/L | 103 | 83.0 | 124 |
| EA025: Total Suspended Solids dried at 104 ± 2°C (QCLot: | 5239102) | | | | | | | |
| EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | 150 mg/L | 102 | 83.0 | 129 |
| | | | | <5 | 1000 mg/L | 98.0 | 82.0 | 110 |
| | | | | <5 | 931 mg/L | 102 | 83.0 | 118 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5236663) | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.1 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 98.0 | 84.0 | 110 |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.0 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 98.5 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.8 | 83.0 | 111 |
| EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 101 | 82.0 | 110 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 94.1 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 103 | 81.0 | 117 |
| EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 99.4 | 82.0 | 112 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5236666) | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 100 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 95.8 | 84.0 | 110 |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.9 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 101 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.8 | 83.0 | 111 |
| EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 99.1 | 82.0 | 110 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.0 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 106 | 81.0 | 117 |
| EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | | 82.0 | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5236665) | 7-100-00-0 | 0.00 | IIIg/L | ٧٥.٥٥ | 0.5 mg/L | 97.7 | 02.0 | 112 |

Page : 6 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | Method Blank (MB) | | Laboratory Control Spike (LC | oratory Control Spike (LCS) Report | | |
|--|---------|------|-------------------|---------------|------------------------------|------------------------------------|------------|--|
| | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| Method: Compound CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5236665) - continued | | | | | | | | |
| EG035F: Mercury 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 92.8 | 83.0 | 105 | |
| EK055G: Ammonia as N by Discrete Analyser (QCLot: 5238030) | | | | | | | | |
| EK055G: Ammonia as N 7664-41-7 | 0.01 | mg/L | <0.01 | 1 mg/L | 102 | 90.0 | 114 | |
| EK057G: Nitrite as N by Discrete Analyser (QCLot: 5237562) | | | | | | | | |
| EK057G: Nitrite as N 14797-65-0 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 104 | 82.0 | 114 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analyser (QCLot: 8 | 238031) | | | | | | | |
| EK059G: Nitrite + Nitrate as N | 0.01 | mg/L | <0.01 | 0.5 mg/L | 103 | 91.0 | 113 | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser (QCLot: 5238027 | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | 0.1 | mg/L | <0.1 | 10 mg/L | 89.1 | 69.0 | 101 | |
| | | | <0.1 | 1 mg/L | 89.2 | 70.0 | 118 | |
| | | | <0.1 | 5 mg/L | 91.0 | 70.0 | 130 | |
| EK067G: Total Phosphorus as P by Discrete Analyser (QCLot: 5238026) | | | | | | | | |
| EK067G: Total Phosphorus as P | 0.01 | mg/L | <0.01 | 4.42 mg/L | 87.1 | 71.3 | 126 | |
| | | | <0.01 | 0.442 mg/L | 87.0 | 71.3 | 126 | |
| | | | <0.01 | 1 mg/L | 97.4 | 70.0 | 130 | |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5235620) | | | | | | | | |
| EP075(SIM): Phenol 108-95-2 | 1 | μg/L | <1.0 | 5 μg/L | 33.4 | 24.5 | 61.9 | |
| EP075(SIM): 2-Chlorophenol 95-57-8 | 1 | μg/L | <1.0 | 5 μg/L | 68.8 | 52.0 | 90.0 | |
| EP075(SIM): 2-Methylphenol 95-48-7 | 1 | μg/L | <1.0 | 5 μg/L | 77.5 | 51.0 | 91.0 | |
| EP075(SIM): 3- & 4-Methylphenol 1319-77-3 | 2 | μg/L | <2.0 | 10 μg/L | 63.5 | 44.0 | 88.0 | |
| EP075(SIM): 2-Nitrophenol 88-75-5 | 1 | μg/L | <1.0 | 5 μg/L | 75.4 | 48.0 | 100 | |
| EP075(SIM): 2.4-Dimethylphenol | 1 | μg/L | <1.0 | 5 μg/L | 71.7 | 49.0 | 99.0 | |
| EP075(SIM): 2.4-Dichlorophenol 120-83-2 | 1 | μg/L | <1.0 | 5 μg/L | 68.2 | 53.0 | 105 | |
| EP075(SIM): 2.6-Dichlorophenol 87-65-0 | 1 | μg/L | <1.0 | 5 μg/L | 69.9 | 57.0 | 105 | |
| EP075(SIM): 4-Chloro-3-methylphenol 59-50-7 | 1 | μg/L | <1.0 | 5 μg/L | 71.2 | 53.0 | 99.0 | |
| EP075(SIM): 2.4.6-Trichlorophenol 88-06-2 | 1 | μg/L | <1.0 | 5 μg/L | 71.8 | 50.0 | 106 | |
| EP075(SIM): 2.4.5-Trichlorophenol 95-95-4 | 1 | μg/L | <1.0 | 5 μg/L | 76.0 | 51.0 | 105 | |
| EP075(SIM): Pentachlorophenol 87-86-5 | 2 | μg/L | <2.0 | 10 μg/L | 33.4 | 10.0 | 95.0 | |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5235874) | | | | | | | | |
| EP075(SIM): Phenol 108-95-2 | 1 | μg/L | <1.0 | 5 μg/L | 35.5 | 24.5 | 61.9 | |
| EP075(SIM): 2-Chlorophenol 95-57-8 | 1 | μg/L | <1.0 | 5 μg/L | 72.2 | 52.0 | 90.0 | |
| EP075(SIM): 2-Methylphenol 95-48-7 | 1 | μg/L | <1.0 | 5 μg/L | 66.5 | 51.0 | 91.0 | |
| EP075(SIM): 3- & 4-Methylphenol 1319-77-3 | 2 | μg/L | <2.0 | 10 μg/L | 59.0 | 44.0 | 88.0 | |
| EP075(SIM): 2-Nitrophenol 88-75-5 | 1 | μg/L | <1.0 | 5 μg/L | 66.8 | 48.0 | 100 | |

Page : 7 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|----------------------|-----|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5235874) - | | | | | | | | |
| EP075(SIM): 2.4-Dimethylphenol | 105-67-9 | 1 | μg/L | <1.0 | 5 μg/L | 72.6 | 49.0 | 99.0 |
| EP075(SIM): 2.4-Dichlorophenol | 120-83-2 | 1 | μg/L | <1.0 | 5 μg/L | 66.7 | 53.0 | 105 |
| EP075(SIM): 2.6-Dichlorophenol | 87-65-0 | 1 | μg/L | <1.0 | 5 μg/L | 73.2 | 57.0 | 105 |
| EP075(SIM): 4-Chloro-3-methylphenol | 59-50-7 | 1 | μg/L | <1.0 | 5 μg/L | 73.1 | 53.0 | 99.0 |
| EP075(SIM): 2.4.6-Trichlorophenol | 88-06-2 | 1 | μg/L | <1.0 | 5 μg/L | 67.1 | 50.0 | 106 |
| EP075(SIM): 2.4.5-Trichlorophenol | 95-95-4 | 1 | μg/L | <1.0 | 5 μg/L | 77.8 | 51.0 | 105 |
| EP075(SIM): Pentachlorophenol | 87-86-5 | 2 | μg/L | <2.0 | 10 μg/L | 35.7 | 10.0 | 95.0 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCL | ot: 5235620) | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 66.7 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 72.0 | 63.6 | 114 |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 72.0 | 62.2 | 113 |
| EP075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 73.8 | 63.9 | 115 |
| EP075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 70.4 | 62.6 | 116 |
| EP075(SIM): Anthracene | 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 93.1 | 64.3 | 116 |
| EP075(SIM): Fluoranthene | 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 76.7 | 63.6 | 118 |
| EP075(SIM): Pyrene | 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 78.3 | 63.1 | 118 |
| EP075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 78.2 | 64.1 | 117 |
| EP075(SIM): Chrysene | 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 77.0 | 62.5 | 116 |
| EP075(SIM): Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1 | μg/L | <1.0 | 5 μg/L | 71.7 | 61.7 | 119 |
| EP075(SIM): Benzo(k)fluoranthene | 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 85.5 | 63.0 | 115 |
| EP075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 78.8 | 63.3 | 117 |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 68.6 | 59.9 | 118 |
| EP075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 70.2 | 61.2 | 117 |
| EP075(SIM): Benzo(g.h.i)perylene | 191-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 69.0 | 59.1 | 118 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCL | ot: 5235874) | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 70.9 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 76.6 | 63.6 | 114 |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 80.4 | 62.2 | 113 |
| EP075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 79.3 | 63.9 | 115 |
| EP075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 75.7 | 62.6 | 116 |
| EP075(SIM): Anthracene | 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 91.4 | 64.3 | 116 |
| EP075(SIM): Fluoranthene | 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 81.0 | 63.6 | 118 |
| EP075(SIM): Pyrene | 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 83.4 | 63.1 | 118 |

Page : 8 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | S) Report | rt Acceptable Limits (%) | | |
|---|----------------------|--------------|------|-------------------|---------------------------------------|--------------------|------------|-----------------------------|--|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | · · · / | | |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | | |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCL | | | | | | | | | | |
| EP075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 73.5 | 64.1 | 117 | | |
| EP075(SIM): Chrysene | 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 87.1 | 62.5 | 116 | | |
| EP075(SIM): Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1 | μg/L | <1.0 | 5 μg/L | 69.2 | 61.7 | 119 | | |
| EP075(SIM): Benzo(k)fluoranthene | 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 76.0 | 63.0 | 115 | | |
| EP075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 83.0 | 63.3 | 117 | | |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 68.6 | 59.9 | 118 | | |
| EP075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 69.8 | 61.2 | 117 | | |
| EP075(SIM): Benzo(g.h.i)perylene | 191-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 71.0 | 59.1 | 118 | | |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 52336 | 625) | | | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 101 | 75.0 | 127 | | |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 52356 | 621) | | | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 66.4 | 53.7 | 97.0 | | |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 80.9 | 63.3 | 107 | | |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 91.1 | 58.3 | 120 | | |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 52358 | 873) | | | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 83.4 | 53.7 | 97.0 | | |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 88.5 | 63.3 | 107 | | |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 79.5 | 58.3 | 120 | | |
| P080/071: Total Petroleum Hydrocarbons (QCLot: 5236 | 018) | | | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 93.9 | 75.0 | 127 | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | 3 Fractions (QCL | ot: 5233625) | | | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 98.0 | 75.0 | 127 | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | 3 Fractions (QCL | ot: 5235621) | | | | | | | | |
| EP071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 79.7 | 53.9 | 95.5 | | |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 80.9 | 57.8 | 110 | | |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 93.4 | 50.5 | 115 | | |
| P080/071: Total Recoverable Hydrocarbons - NEPM 2013 | 3 Fractions (QCL | ot: 5235873) | | | | | | | | |
| P071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 66.9 | 53.9 | 95.5 | | |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 89.5 | 57.8 | 110 | | |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 89.3 | 50.5 | 115 | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | 3 Fractions (QCL | ot: 5236018) | | | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 99.5 | 75.0 | 127 | | |

Page : 9 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD

Project : 20102 REDIRECT WETHERILL PARK



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|------------|-----|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP080: BTEXN (QCLot: 5233625) - continued | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 91.7 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 92.1 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 89.1 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 99.7 | 73.0 | 122 |
| | 106-42-3 | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 102 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 84.2 | 75.5 | 124 |
| EP080: BTEXN (QCLot: 5236018) | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 99.0 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 95.6 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 100 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 102 | 73.0 | 122 |
| | 106-42-3 | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 104 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 102 | 75.5 | 124 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | | Ma | atrix Spike (MS) Report | | |
|---------------------|-----------------------------------|---------------------|------------|---------------|-------------------------|------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable | Limits (%) |
| aboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG020F: Dissolved | Metals by ICP-MS (QCLot: 5236663) | | | | | | |
| ES2327041-002 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 106 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 98.2 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 98.4 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 105 | 70.0 | 130 |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 95.7 | 70.0 | 130 |
| | | EG020A-F: Manganese | 7439-96-5 | 1 mg/L | 93.8 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 97.4 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 98.0 | 70.0 | 130 |
| G020F: Dissolved | Metals by ICP-MS (QCLot: 5236666) | | | | | | |
| ES2327328-005 | MW6 | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 109 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 126 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 118 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 108 | 70.0 | 130 |

Page : 10 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | M | atrix Spike (MS) Report | • | |
|----------------------|--|--------------------------------------|--------------------|---------------|-------------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG020F: Dissolve | d Metals by ICP-MS (QCLot: 5236666) - continue | d | | | | | |
| ES2327328-005 | MW6 | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 115 | 70.0 | 130 |
| | | EG020A-F: Manganese | 7439-96-5 | 1 mg/L | 125 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 129 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 103 | 70.0 | 130 |
| EG035F: Dissolve | d Mercury by FIMS (QCLot: 5236665) | | | | | | |
| ES2327080-001 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 89.1 | 70.0 | 130 |
| EK055G: Ammonia | a as N by Discrete Analyser (QCLot: 5238030) | | | | | | |
| ES2327281-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 1 mg/L | 119 | 70.0 | 130 |
| EK057G: Nitrite as | s N by Discrete Analyser (QCLot: 5237562) | | | | | | |
| ES2327281-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.5 mg/L | 112 | 70.0 | 130 |
| EK059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCI | | | | | | |
| ES2327281-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 110 | 70.0 | 130 |
| | eldahl Nitrogen By Discrete Analyser (QCLot: 523 | | | 3 | | | |
| ES2327281-002 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 5 mg/L | 88.5 | 70.0 | 130 |
| | osphorus as P by Discrete Analyser (QCLot: 523) | | | o mg/L | 00.0 | 70.0 | 100 |
| ES2327281-002 | | | | 1 ma/l | 92.5 | 70.0 | 130 |
| | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 92.5 | 70.0 | 130 |
| | Petroleum Hydrocarbons (QCLot: 5233625) | | | | | | |
| ES2327291-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 127 | 70.0 | 130 |
| EP080/071: Total F | Petroleum Hydrocarbons (QCLot: 5236018) | | | | | | |
| ES2327093-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 88.8 | 70.0 | 130 |
| EP080/071: Total F | Recoverable Hydrocarbons - NEPM 2013 Fractions | s (QCLot: 5233625) | | | | | |
| ES2327291-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 128 | 70.0 | 130 |
| EP080/071: Total F | Recoverable Hydrocarbons - NEPM 2013 Fractions | s (QCLot: 5236018) | | | | | |
| ES2327093-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 90.7 | 70.0 | 130 |
| EP080: BTEXN (Q | (CLot: 5233625) | | | | | | |
| ES2327291-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 106 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 112 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 123 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 128 | 70.0 | 130 |
| | | | 106-42-3 | 05. " | 400 | 70.0 | 100 |
| | | EP080: ortho-Xylene | 95-47-6 91-20-3 | 25 μg/L | 126 94.9 | 70.0 70.0 | 130 130 |
| | 201 / 2000/0 | EP080: Naphthalene | 91-20-3 | 25 μg/L | 94.9 | 70.0 | 130 |
| EP080: BTEXN (Q | | | | | | | |
| ES2327093-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 95.5 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 91.4 | 70.0 | 130 |

Page : 11 of 11

Work Order : ES2327328 Amendment 2
Client : SENVERSA PTY LTD



| Sub-Matrix: WATER | | | | Ma | Matrix Spike (MS) Report | | | | |
|----------------------|----------------------------|----------------------------|------------|---------------|--------------------------|--------------|-----------|--|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable l | imits (%) | | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | | |
| EP080: BTEXN (QC | CLot: 5236018) - continued | | | | | | | | |
| ES2327093-001 | Anonymous | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 100.0 | 70.0 | 130 | | |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 101 | 70.0 | 130 | | |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 102 | 70.0 | 130 | | |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 86.1 | 70.0 | 130 | | |

Carrier / Reference #: Carrier / Reference #: Carrier / Reference #:

Name/Signature: Name/Signature: lame/Signature:

Date/Time: Date/Time:

Date: Time:

Name/Signature: Name/Signature:

Time: Date: Time: Date:

Chain of Custody Documentation

senversa

| Senversa Pty Ltd | Senversa Pty Ltd | | | Laboratory: | | L | | | (| Analysis Required | | | ſ |
|-------------------------------|---|---|--------------------------|--------------------------------|-------------------------------|---|--------|--------------|-----------------|--|----------------------------|---|---------|
| ABN 89 132 231 380 | 2 231 380 · * | | | Address: Contact: Phone: | | · · · · · | HHy ! | | (x3 | | | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. | : trace |
| Job Number: | \$ 20102 | | | D. Coophorn | | ×SI | | | | | | s | |
| Project Name: | 2 | ill Park | NA | Quote No: | EN/103/21 | 15/ | | DT UT | - | | | | - |
| Sampled By: |) | 10.2 | | _ | Standord | | | |] = | | | | |
| Project Manager: | | sals a | 4 | _ | - | ~ | | | 15 | | | 4 | |
| Email Report To: | | 20 318 COS | The war to save the come | | C42021847 | راء | | 20 | Ç00 | | | of. | |
| | | Sample Information | tion | | Container Information | | | | ر | | a | | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | al Bottles | | 7070 | 3 | | ПОГ | | - |
| - | 50 3 | 3 | 712124 | | | ٥ | × | T | _ | | 1 | | T |
| ~ 0 | SW 2 | 3 | 712124 | | | ٩ | X | × | /× | | | | T |
| 7 | BC403 TR | 3 | 212124 | | | - | | 1 | × | | | | I |
| 5 | 51 50570 | 3 | riric | | | - | | - | 0 | | | | T |
| | | | | | | | | I | | | + | | T |
| | | | | | | | | | | | | | T |
| | | * | | | | | | | | r | 1 | | |
| | * | | | | | | | - | | Fnvir | Environmental Division | noioivi | T |
| | | | | | | | | - | 1 | Svdn | ev ev | I I I I I I I I I I I I I I I I I I I | T |
| | | | | | | 1 | + | | | Wo | Work Order Reference | ence | T |
| | | | | | | | _ | | | <u>Н</u> | S2403 | 942 | |
| | ** | | | | | | | | | | | | T |
| | | | | | | | | | + | | 後くきりまし | | T |
| | | | | | | | | L | | | | | T |
| | | | | | 1 | | | - | | | | | T |
| D | | | | | | | | | | | | 4 | T |
| | | | | | | Dec 19 | | | | Telephon | Telephone: +61-2-8784 8555 | | Τ |
| | | | | | | | | | | | | | |
| | | | | | | | | | | | | | Γ |
| | | | | | | | | | | | | | * |
| Total | | | | | | | | | | | | | |
| Sampler: I a specification | Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: | ing procedures in ection of these sa | accordance with Ser | nversa standard proce | | Sampler Name: R | Parote | 7 | Signature: Read | 257 | Date: 2/ | 721216 | Г |
| Relinquished By: | ad By: | | | | Method of Shipmont (if any or | icable). | | 11 | | The second secon | | | 7 |
| Name/Signature: | Ro | he - | | Date: 71217 | Carrier / Reference #* | icable). | A Z | Received by: | inco C | S. S | | 2/2/2. | T |
| OF: | | W | | L. 1 | Date/Time: | | Of: | of: | 200 | | Date: | 100 | T |

| Time: | Date/Time: | Date/Tim

Completed by: Checked by:



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2403942

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : sandy.phan@alsglobal.com

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555 Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 2

 Order number
 : --- Quote number
 : EB2023SENVER0001 (EN/000)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Site : ---Sampler : Rowan Faint

Dates

Date Samples Received : 07-Feb-2024 15:46 Issue Date : 08-Feb-2024

Client Requested Due : 15-Feb-2024 Scheduled Reporting Date : 15-Feb-2024

Date

Delivery Details

Mode of Delivery : Client Drop Off Security Seal : Not Available

No. of coolers/boxes : 1 Temperature : 11.3'C, 14.4'C, 16.2'C - Ice

present

Receipt Detail : No. of samples received / analysed : 4 / 4

General Comments

• This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 08-Feb-2024 Issue Date

Page

: 2 of 2 : ES2403942 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

| process necessal tasks. Packages as the determin tasks, that are included in the sampling default 00:00 on | may for the execution may contain ad ation of moisture uded in the package. time is provided, the date of sampling date wi | be part of a laboratory ion of client requested ditional analyses, such content and preparation the sampling time will ag. If no sampling date ill be assumed by the ackets without a time Sample ID | WATER - EA015H Total Dissolved Solids - Standard Level | WATER - EA025H Suspended Solids - Standard Level | WATER - EG005F Dissolved Metals by ICPAES | WATER - EP080 BTEXN | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - W-18 TRH(C6 - C9)/BTEXN | WATER - W-27 TRH/BTEXN/PAH/Phenols/8 Metals |
|--|--|--|--|--|---|------------------------|---|------------------------------------|--|
| ES2403942-001 | 07-Feb-2024 00:00 | SW 1 | ✓ | ✓ | ✓ | | ✓ | | ✓ |
| ES2403942-002 | 07-Feb-2024 00:00 | SW 2 | ✓ | ✓ | 1 | | ✓ | | ✓ |
| ES2403942-003 | 05-Feb-2024 00:00 | QC403 | | | | | | ✓ | |
| ES2403942-004 | 05-Feb-2024 00:00 | QC503 | | | | 1 | | | |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

BEC CHAPPLE

| 520 017.11 22 | | |
|--|-------|---------------------------------|
| - *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| EMMA WALSH | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Tax Invoice (INV) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| Rowan Faint | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | rowan.faint@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | rowan.faint@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | rowan.faint@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | rowan.faint@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | rowan.faint@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | rowan.faint@senversa.com.au |
| SUPPLIER ACCOUNTS | | |
| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
| | | u |
| | | |



C-O-C number

CERTIFICATE OF ANALYSIS

Work Order : ES2403942

Client : SENVERSA PTY LTD

Contact : EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number : ----

Sampler : Rowan Faint

Site : ---

Quote number : EN/000

No. of samples received : 4
No. of samples analysed : 4

Page : 1 of 7

Laboratory : Environmental Division Sydney

Contact : Sandy Phan

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61-2-8784 8555

Date Samples Received : 07-Feb-2024 15:46

Date Analysis Commenced : 08-Feb-2024

Issue Date : 15-Feb-2024 14:03



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-----------------|-----------------------------|------------------------------------|
| Ankit Joshi | Senior Chemist - Inorganics | Sydney Inorganics, Smithfield, NSW |
| Edwandy Fadjar | Organic Coordinator | Sydney Organics, Smithfield, NSW |
| Sanjeshni Jyoti | Senior Chemist Volatiles | Sydney Organics, Smithfield, NSW |

Page : 2 of 7

Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a.h)anthracene (1.0), Benzo(g.h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.

Page : 3 of 7
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | Sample ID | SW 1 | SW 2 | QC403 | QC503 | |
|---------------------------------------|-----------------------------|---------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| (Matrix: WATER) | | Sampli | ng date / time | 07-Feb-2024 00:00 | 07-Feb-2024 00:00 | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2403942-001 | ES2403942-002 | ES2403942-003 | ES2403942-004 | |
| | <i>5,</i> 15, 14, 11, 15, 1 | | | Result | Result | Result | Result | |
| EA015: Total Dissolved Solids dried a | at 180 ± 5 °C | | | | | | | |
| Total Dissolved Solids @180°C | | 10 | mg/L | 374 | 394 | | | |
| EA025: Total Suspended Solids dried | l at 104 ± 2°C | | | | | | | |
| Suspended Solids (SS) | | 5 | mg/L | 91 | 8 | | | |
| EG005(ED093)F: Dissolved Metals by | ICP-AES | | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | | | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 0.02 | <0.01 | | | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.001 | 0.003 | | | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.004 | 0.004 | | | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.014 | | | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| EK059G: Nitrite plus Nitrate as N (NC | 0x) by Discrete Ana | lyser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.41 | 0.28 | | | |
| EK061G: Total Kjeldahl Nitrogen By D | Discrete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.7 | 0.9 | | | |
| EK062G: Total Nitrogen as N (TKN + I | NOx) by Discrete A | nalyser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 2.1 | 1.2 | | | |
| EK067G: Total Phosphorus as P by D | iscrete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.10 | 0.03 | | | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |

Page : 4 of 7
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW 1 | SW 2 | QC403 | QC503 | |
|---------------------------------------|-------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 07-Feb-2024 00:00 | 07-Feb-2024 00:00 | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2403942-001 | ES2403942-002 | ES2403942-003 | ES2403942-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| EP075(SIM)B: Polynuclear Aromatic | c Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| ^ Sum of polycyclic aromatic hydrocar | rbons | 0.5 | μg/L | <0.5 | <0.5 | | | |

Page : 5 of 7
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW 1 | SW 2 | QC403 | QC503 | |
|--|---------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 07-Feb-2024 00:00 | 07-Feb-2024 00:00 | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2403942-001 | ES2403942-002 | ES2403942-003 | ES2403942-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons - Cont | inued | | | <u></u> | | | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | | | |
| EP080/071: Total Petroleum Hydroca | rbons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | 490 | <50 | | | |
| C15 - C28 Fraction | | 100 | μg/L | 560 | <100 | | | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | | | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | 1050 | <50 | | | |
| EP080/071: Total Recoverable Hydro | carbons - NEPM 201 | 3 Fraction | ns | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | | |
| ^ C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | | |
| >C10 - C16 Fraction | | 100 | μg/L | 630 | <100 | | | |
| >C16 - C34 Fraction | | 100 | μg/L | 460 | <100 | | | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | | | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | 1090 | <100 | | | |
| ^ >C10 - C16 Fraction minus Naphthalen (F2) | e | 100 | μg/L | 630 | <100 | | | |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | 20 | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | 18 | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | 17 | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | 16 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | 16 | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | 32 | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | 87 | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | 16 | |
| EP075(SIM)S: Phenolic Compound S | urrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 25.8 | 26.8 | | | |

Page : 6 of 7
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW 1 | SW 2 | QC403 | QC503 | |
|-----------------------------------|------------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 07-Feb-2024 00:00 | 07-Feb-2024 00:00 | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2403942-001 | ES2403942-002 | ES2403942-003 | ES2403942-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)S: Phenolic Compound | Surrogates - Continued | 1 | | | | | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 51.8 | 54.8 | | | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 58.8 | 72.6 | | | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 76.4 | 74.8 | | | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 92.4 | 75.5 | | | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 92.6 | 89.0 | | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 122 | 134 | 115 | 101 | |
| Toluene-D8 | 2037-26-5 | 2 | % | 119 | 129 | 114 | 109 | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 122 | 135 | 104 | 97.7 | |

Page : 7 of 7
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | | | |
|--------------------------------------|------------|---------------------|------|--|--|
| Compound | CAS Number | Low | High | | |
| EP075(SIM)S: Phenolic Compound Surro | ogates | | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 | | |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 | | |
| EP075(SIM)T: PAH Surrogates | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 | | |
| Anthracene-d10 | 1719-06-8 | 27 | 113 | | |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 | | |
| Toluene-D8 | 2037-26-5 | 75 | 131 | | |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 | | |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2403942** Page : 1 of 8

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

 Contact
 : EMMA WALSH
 Telephone
 : +61-2-8784 8555

 Project
 : S20102 Wetherill Park WME
 Date Samples Received
 : 07-Feb-2024

 Site
 : --- Issue Date
 : 15-Feb-2024

Sampler : Rowan Faint No. of samples received : 4
Order number : ---- No. of samples analysed : 4

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- Matrix Spike outliers exist please see following pages for full details.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

• NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 8 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Outliers: Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: WATER

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|---|----------------------|------------------|------------------------|------------|------------|--------|----------------------------------|
| Matrix Spike (MS) Recoveries | | | | | | | |
| EG020F: Dissolved Metals by ICP-MS | ES2403761003 | Anonymous | Zinc | 7440-66-6 | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete A | ES2403912001 | Anonymous | Nitrite + Nitrate as N | | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |
| EK067G: Total Phosphorus as P by Discrete Analyser | ES2403868001 | Anonymous | Total Phosphorus as P | | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |

Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | | Co | ount | Rate | e (%) | Quality Control Specification |
|-----------------------------|------------|----|---------|--------|----------|--------------------------------|
| Analytical Methods | Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 4 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | 1 | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 4 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: **x** = Holding time breach ; ✓ = Within holding time.

| Watth. WATER | | | | | Lvaluation | . W - Holding time | breach, with | ir riolaling time. |
|---|------|-------------|--------------------------|--------------------|------------|--------------------|------------------|--------------------|
| Method | hod | | Extraction / Preparation | | | Analysis | | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA015H) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 13-Feb-2024 | 14-Feb-2024 | ✓ |
| EA025: Total Suspended Solids dried at 104 ± 2°C | | | | | | | | |
| Clear Plastic Bottle - Natural (EA025H) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 13-Feb-2024 | 14-Feb-2024 | ✓ |

Page : 3 of 8 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Matrix: WATER | | | | | Lvaldation | . • - Holding time | breach, V = With | ir noluling time |
|--|----------------|--------------|----------------|------------------------|------------|--------------------|------------------|------------------|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG005(ED093)F: Dissolved Metals by ICP-AES | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG005F) | | T T | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 09-Feb-2024 | 05-Aug-2024 | ✓ |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 08-Feb-2024 | 05-Aug-2024 | ✓ |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 12-Feb-2024 | 06-Mar-2024 | ✓ |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete A | nalyser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) | | | | | | | 00 M = 0004 | |
| SW 1, | SW 2 | 07-Feb-2024 | | | | 12-Feb-2024 | 06-Mar-2024 | ✓ |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | SW 2 | 07-Feb-2024 | 12-Feb-2024 | 06-Mar-2024 | , | 12-Feb-2024 | 06-Mar-2024 | |
| SW 1, | | 07-Feb-2024 | 12-Peb-2024 | 00-IVIAI-2024 | ✓ | 12-Feb-2024 | 00-Mai-2024 | ✓ |
| EK067G: Total Phosphorus as P by Discrete Analyser | | 1 | | ı | | | | <u> </u> |
| Clear Plastic Bottle - Sulfuric Acid (EK067G) SW 1. | SW 2 | 07-Feb-2024 | 12-Feb-2024 | 06-Mar-2024 | 1 | 12-Feb-2024 | 06-Mar-2024 | 1 |
| | SW Z | 07-1 05-2024 | 12-1 05-2024 | 00 Mai 2021 | | 12-1 05-2024 | 00 Mai 2021 | V |
| EP075(SIM)A: Phenolic Compounds Amber Glass Bottle - Unpreserved (EP075(SIM)) | | <u> </u> | <u> </u> | | | <u> </u> | | |
| SW 1, | SW 2 | 07-Feb-2024 | 08-Feb-2024 | 14-Feb-2024 | 1 | 13-Feb-2024 | 19-Mar-2024 | 1 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons | | | | | _ | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | 08-Feb-2024 | 14-Feb-2024 | ✓ | 13-Feb-2024 | 19-Mar-2024 | 1 |
| EP080/071: Total Petroleum Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | 08-Feb-2024 | 14-Feb-2024 | ✓ | 12-Feb-2024 | 19-Mar-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| QC403 | | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | √ | 14-Feb-2024 | 19-Feb-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) SW 1, | SW 2 | 07-Feb-2024 | 09-Feb-2024 | 21-Feb-2024 | 1 | 10-Feb-2024 | 21-Feb-2024 | 1 |
| | | 01-1 CD-2024 | 00-1 00-2024 | 21100-2024 | <u> </u> | 10-1 05-2024 | 21105-2024 | V |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2 | 2013 Fractions | 1 | <u> </u> | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) SW 1, | SW 2 | 07-Feb-2024 | 08-Feb-2024 | 14-Feb-2024 | 1 | 12-Feb-2024 | 19-Mar-2024 | 1 |
| Amber VOC Vial - Sulfuric Acid (EP080) | 52 | | | | | | | V |
| QC403 | | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | ✓ | 14-Feb-2024 | 19-Feb-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | 09-Feb-2024 | 21-Feb-2024 | ✓ | 10-Feb-2024 | 21-Feb-2024 | ✓ |

Page : 4 of 8 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
|--|-------|-------------|----------------|------------------------|------------|---------------|------------------|------------|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP080: BTEXN | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) |) | | | | | | | |
| QC403, | QC503 | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | ✓ | 14-Feb-2024 | 19-Feb-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) |) | | | | | | | |
| SW 1, | SW 2 | 07-Feb-2024 | 09-Feb-2024 | 21-Feb-2024 | ✓ | 10-Feb-2024 | 21-Feb-2024 | ✓ |

Page : 5 of 8
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

he expected rate. A listing of breaches is provided in the Summary of Outliers.

| Matrix: WATER | | | | Evaluatio | n: × = Quality Co | entrol frequency | not within specification; ✓ = Quality Control frequency within specification |
|---|------------|----|---------|-----------|-------------------|------------------|--|
| Quality Control Sample Type | | Co | ount | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Regular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 9 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 2 | 4 | 50.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 18 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 10.00 | 3c | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 4 | 39 | 10.26 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 18 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 18 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 4 | 0.00 | 10.00 | se | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 4 | 39 | 10.26 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 9 | 11.11 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 4 | 25.00 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 18 | 5.56 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 4 | 25.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 5 | 40 | 12.50 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 5 | 39 | 12.82 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 18 | 16.67 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 3 | 18 | 16.67 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 4 | 25.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 39 | 5.13 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 9 | 11.11 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 4 | 25.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 18 | 5.56 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 4 | 25.00 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 2 | 40 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 2 | 39 | 5.13 | 5.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 18 | 5.56 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 18 | 5.56 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 4 | 25.00 | 5.00 | √ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 39 | 5.13 | 5.00 | 1 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |

Page : 6 of 8 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Evaluation: × = Quality Control frequency not within specification; ✓ = Quality Control frequency within specification. Matrix: WATER Quality Control Sample Type Count Rate (%) Quality Control Specification Analytical Methods Method QC Evaluation Regular Actual Expected Matrix Spikes (MS) - Continued Dissolved Mercury by FIMS 9 5.00 NEPM 2013 B3 & ALS QC Standard EG035F 1 11.11 1 Dissolved Metals by ICP-AES 1 4 NEPM 2013 B3 & ALS QC Standard 25.00 5.00 EG005F 1 9 Dissolved Metals by ICP-MS - Suite A 1 11.11 5.00 NEPM 2013 B3 & ALS QC Standard EG020A-F ✓ Nitrite and Nitrate as N (NOx) by Discrete Analyser 1 18 5.56 NEPM 2013 B3 & ALS QC Standard EK059G 5.00 1 PAH/Phenols (GC/MS - SIM) EP075(SIM) 0 4 0.00 5.00 NEPM 2013 B3 & ALS QC Standard × Total Kjeldahl Nitrogen as N By Discrete Analyser 1 18 NEPM 2013 B3 & ALS QC Standard 5.56 5.00 EK061G ✓ Total Phosphorus as P By Discrete Analyser 1 18 5.56 5.00 NEPM 2013 B3 & ALS QC Standard EK067G 1 TRH - Semivolatile Fraction 0 4 0.00 5.00 NEPM 2013 B3 & ALS QC Standard EP071 × TRH Volatiles/BTEX 2 39 EP080 5.13 5.00 1 NEPM 2013 B3 & ALS QC Standard

Page : 7 of 8
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|--|----------|--------|---|
| Total Dissolved Solids (High Level) | EA015H | WATER | In house: Referenced to APHA 2540C. A gravimetric procedure that determines the amount of 'filterable' residue in an aqueous sample. A well-mixed sample is filtered through a glass fibre filter (1.2um). The filtrate is evaporated to dryness and dried to constant weight at 180+/-5C. This method is compliant with NEPM Schedule B(3) |
| Suspended Solids (High Level) | EA025H | WATER | In house: Referenced to APHA 2540D. A gravimetric procedure employed to determine the amount of `non-filterable` residue in a aqueous sample. The prescribed GFC (1.2um) filter is rinsed with deionised water, oven dried and weighed prior to analysis. A well-mixed sample is filtered through a glass fibre filter (1.2um). The residue on the filter paper is dried at 104+/-2C. This method is compliant with NEPM Schedule B(3) |
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered samples, emitting a characteristic spectrum which is compared against matrix matched standards. This method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |

Page : 8 of 8 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Analytical Methods | Method | Matrix | Method Descriptions |
|---|-------------|--------|--|
| PAH/Phenois (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| Preparation Methods | Method | Matrix | Method Descriptions |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| Separatory Funnel Extraction of Liquids | ORG14 | WATER | In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated and concentrated for analysis. This method is compliant with NEPM Schedule B(3). ALS default excludes sediment which may be resident in the container. |
| | | | |



QUALITY CONTROL REPORT

Work Order : **ES2403942** Page : 1 of 9

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555

Project : S20102 Wetherill Park WME Date Samples Received : 07-Feb-2024
Order number : ---- Date Analysis Commenced : 08-Feb-2024

C-O-C number ---- Issue Date · 15-Feb-2024

Sampler : Rowan Faint

Site : ----

No. of samples analysed : 4

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall

Accreditation No. 825

Accredited for compliance with

not be reproduced, except in full.

This Quality Control Report contains the following information:

: EN/000

: 4

Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits

Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits

Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

Quote number

No. of samples received

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-----------------|-----------------------------|------------------------------------|
| Ankit Joshi | Senior Chemist - Inorganics | Sydney Inorganics, Smithfield, NSW |
| Edwandy Fadjar | Organic Coordinator | Sydney Organics, Smithfield, NSW |
| Sanjeshni Jyoti | Senior Chemist Volatiles | Sydney Organics, Smithfield, NSW |

Page : 2 of 9 Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit: Result between 10 and 20 times LOR: 0% - 50%: Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory L | Duplicate (DUP) Report | | |
|----------------------|-----------------------------|---------------------------------------|------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | solved Metals by ICP-AES | G (QC Lot: 5590163) | | | | | | | |
| ES2403595-002 | Anonymous | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 47.0 | 47.8 | 1.6 | 0% - 20% |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 175 | 178 | 2.0 | 0% - 20% |
| ES2403942-002 | SW 2 | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EA015: Total Dissolv | ved Solids dried at 180 ± 5 | °C (QC Lot: 5598190) | | | | | | | |
| ES2403942-001 | SW 1 | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 374 | 367 | 1.9 | 0% - 20% |
| ES2404000-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 182 | 212 | 14.8 | 0% - 20% |
| ES2404300-002 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 36 | 47 | 27.2 | No Limit |
| EW2400609-004 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 146 | 145 | 0.0 | 0% - 50% |
| EA025: Total Susper | nded Solids dried at 104 ± | 2°C (QC Lot: 5598191) | | | | | | | |
| ES2403942-001 | SW 1 | EA025H: Suspended Solids (SS) | | 5 | mg/L | 91 | 77 | 16.6 | 0% - 50% |
| ES2404000-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 140 | 130 | 7.6 | 0% - 20% |
| ES2404300-002 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 10 | 9 | 12.7 | No Limit |
| EW2400609-004 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | <5 | 0.0 | No Limit |
| EG020F: Dissolved I | Metals by ICP-MS (QC Lot | t: 5590161) | | | | | | | |
| ES2403595-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | 0.0026 | 0.0026 | 0.0 | 0% - 20% |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.001 | 0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 0.025 | 0.024 | 0.0 | 0% - 20% |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | 0.005 | 0.005 | 0.0 | No Limit |

Page : 3 of 9
Work Order : ES2403942



| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|---------------------------|---|------------|--------------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG020F: Dissolved | Metals by ICP-MS (QC L | ot: 5590161) - continued | | | | | | | |
| ES2403595-001 | Anonymous | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 1.45 | 1.43 | 1.6 | 0% - 20% |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 2.04 | 2.01 | 1.2 | 0% - 20% |
| EG035F: Dissolved | Mercury by FIMS (QC Lo | ot: 5590159) | | | | | | | |
| ES2403761-002 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | 0.0070 | 0.0074 | 5.3 | 0% - 20% |
| EK059G: Nitrite plus | s Nitrate as N (NOx) by | Discrete Analyser (QC Lot: 5594704) | | | | | | | |
| ES2403912-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 6.88 | 6.79 | 1.3 | 0% - 20% |
| ES2403955-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.04 | 0.03 | 0.0 | No Limit |
| EK061G: Total Kjeld | lahl Nitrogen By Discrete | e Analyser (QC Lot: 5594699) | | | | | | | |
| ES2403852-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (1.0)* | mg/L | 27.9 | 26.8 | 4.2 | 0% - 20% |
| ES2403919-008 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 2.9 | 2.4 | 18.8 | 0% - 20% |
| EK067G: Total Phos | phorus as P by Discrete | Analyser (QC Lot: 5594700) | | | | | | | |
| ES2403852-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 (0.10)* | mg/L | 8.31 | 8.35 | 0.5 | 0% - 20% |
| ES2403919-008 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.04 | 0.03 | 0.0 | No Limit |
| EP080/071: Total Pe | troleum Hydrocarbons (| (QC Lot: 5593731) | | | | | | | |
| ES2403857-007 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <0.02 mg/L | <20 | 0.0 | No Limit |
| ES2403857-046 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <0.02 mg/L | <20 | 0.0 | No Limit |
| EP080/071: Total Pe | troleum Hydrocarbons (| (QC Lot: 5596095) | | | | | | | |
| ES2403888-002 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2403888-008 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Re | coverable Hydrocarbons | s - NEPM 2013 Fractions (QC Lot: 5593731) | | | | | | | |
| ES2403857-007 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <0.02 mg/L | <20 | 0.0 | No Limit |
| ES2403857-046 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <0.02 mg/L | <20 | 0.0 | No Limit |
| EP080/071: Total Re | coverable Hydrocarbons | s - NEPM 2013 Fractions (QC Lot: 5596095) | | | | | | | |
| ES2403888-002 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2403888-008 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5593731) | | | | | | | | |
| ES2403857-007 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <0.001 mg/L | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <0.005 mg/L | <5 | 0.0 | No Limit |
| ES2403857-046 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <0.001 mg/L | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |

Page : 4 of 9
Work Order : ES2403942



| Sub-Matrix: WATER | | | | | | Laboratory L | Ouplicate (DUP) Report | | |
|----------------------|---------------------|----------------------------|------------|-----|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP080: BTEXN (QC | Lot: 5593731) - con | tinued | | | | | | | |
| ES2403857-046 | Anonymous | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <0.002 mg/L | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <0.005 mg/L | <5 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5596095) | | | | | | | | |
| ES2403888-002 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| ES2403888-008 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 5 of 9
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|--|----------------------|--------|------|-------------------|---------------|------------------------------|------------|--------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCL | ot: 5590163) | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 106 | 82.0 | 114 |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 106 | 81.0 | 113 |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C (Q | CLot: 5598190) | | | | | | | |
| EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | <10 | 2000 mg/L | 102 | 87.0 | 109 |
| | | | | <10 | 293 mg/L | 117 | 75.2 | 126 |
| | | | | <10 | 2470 mg/L | 103 | 83.0 | 124 |
| EA025: Total Suspended Solids dried at 104 ± 2°C(0 | QCLot: 5598191) | | | | | | | |
| EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | 150 mg/L | 94.7 | 83.0 | 129 |
| | | | | <5 | 1000 mg/L | 94.4 | 82.0 | 110 |
| | | | | <5 | 841 mg/L | 106 | 83.0 | 118 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 55901 | 61) | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 99.8 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 97.6 | 84.0 | 110 |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 97.2 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 104 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.5 | 83.0 | 111 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.3 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 86.7 | 81.0 | 117 |
| EG035F: Dissolved Mercury by FIMS (QCLot: 55901 | 59) | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 93.7 | 83.0 | 105 |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete | Analyser (QCLot: 559 | (4704) | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 107 | 91.0 | 113 |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analys | er (OCL of: 5594699) | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 93.3 | 69.0 | 101 |
| , , | | | | <0.1 | 1 mg/L | 97.1 | 70.0 | 118 |
| | | | | <0.1 | 5 mg/L | 105 | 70.0 | 130 |
| EK067G: Total Phosphorus as P by Discrete Analyse | r (QCLot: 5594700) | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 87.9 | 71.3 | 126 |
| · | | | | <0.01 | 0.442 mg/L | 89.7 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 102 | 70.0 | 130 |
| EP075(SIM)A: Phenolic Compounds (QCLot: 558882 | 5) | | | | | | | |

Page : 6 of 9
Work Order : ES2403942



| Sub-Matrix: WATER | | | | Method Blank (MB) | | S) Report | | | |
|--|----------------|-----|------|-------------------|--|-----------|------|------|--|
| | | | | Report | Spike Spike Recovery (%) Acceptable Limits Concentration LCS Low | | | | |
| Method: Compound CAS N | umber | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5588825) - continue | | | | | | | | | |
| Zi oro(ciiti). I nonoi | -95-2 | 1 | μg/L | <1.0 | 5 μg/L | 32.5 | 24.5 | 61.9 | |
| EP075(SIM): 2-Chlorophenol | -57-8 | 1 | μg/L | <1.0 | 5 μg/L | 72.2 | 52.0 | 90.0 | |
| EP075(SIM): 2-Methylphenol 95 | -48-7 | 1 | μg/L | <1.0 | 5 μg/L | 68.8 | 51.0 | 91.0 | |
| EP075(SIM): 3- & 4-Methylphenol | -77-3 | 2 | μg/L | <2.0 | 10 μg/L | 58.8 | 44.0 | 88.0 | |
| EP075(SIM): 2-Nitrophenol | -75-5 | 1 | μg/L | <1.0 | 5 μg/L | 71.8 | 48.0 | 100 | |
| EP075(SIM): 2.4-Dimethylphenol | -67-9 | 1 | μg/L | <1.0 | 5 μg/L | 86.5 | 49.0 | 99.0 | |
| EP075(SIM): 2.4-Dichlorophenol | -83-2 | 1 | μg/L | <1.0 | 5 μg/L | 71.7 | 53.0 | 105 | |
| EP075(SIM): 2.6-Dichlorophenol | -65-0 | 1 | μg/L | <1.0 | 5 μg/L | 74.7 | 57.0 | 105 | |
| EP075(SIM): 4-Chloro-3-methylphenol | -50-7 | 1 | μg/L | <1.0 | 5 μg/L | 66.4 | 53.0 | 99.0 | |
| EP075(SIM): 2.4.6-Trichlorophenol | -06-2 | 1 | μg/L | <1.0 | 5 μg/L | 80.4 | 50.0 | 106 | |
| EP075(SIM): 2.4.5-Trichlorophenol | -95-4 | 1 | μg/L | <1.0 | 5 μg/L | 70.8 | 51.0 | 105 | |
| EP075(SIM): Pentachlorophenol 87 | -86-5 | 2 | μg/L | <2.0 | 10 μg/L | 37.9 | 10.0 | 95.0 | |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLot: 55888 | 25) | | | | | | | | |
| EP075(SIM): Naphthalene 91 | -20-3 | 1 | μg/L | <1.0 | 5 μg/L | 68.8 | 50.0 | 94.0 | |
| EP075(SIM): Acenaphthylene 208 | -96-8 | 1 | μg/L | <1.0 | 5 μg/L | 70.7 | 63.6 | 114 | |
| EP075(SIM): Acenaphthene | -32-9 | 1 | μg/L | <1.0 | 5 μg/L | 72.6 | 62.2 | 113 | |
| EP075(SIM): Fluorene 86 | -73-7 | 1 | μg/L | <1.0 | 5 μg/L | 76.6 | 63.9 | 115 | |
| EP075(SIM): Phenanthrene 85 | -01-8 | 1 | μg/L | <1.0 | 5 μg/L | 86.1 | 62.6 | 116 | |
| EP075(SIM): Anthracene 120 | -12-7 | 1 | μg/L | <1.0 | 5 μg/L | 81.4 | 64.3 | 116 | |
| EP075(SIM): Fluoranthene 206 | -44-0 | 1 | μg/L | <1.0 | 5 μg/L | 98.4 | 63.6 | 118 | |
| EP075(SIM): Pyrene 129 | -00-0 | 1 | μg/L | <1.0 | 5 μg/L | 99.0 | 63.1 | 118 | |
| EP075(SIM): Benz(a)anthracene 56 | -55-3 | 1 | μg/L | <1.0 | 5 μg/L | 100 | 64.1 | 117 | |
| EP075(SIM): Chrysene 218 | -01-9 | 1 | μg/L | <1.0 | 5 μg/L | 98.6 | 62.5 | 116 | |
| , | -99-2 -82-3 | 1 | μg/L | <1.0 | 5 μg/L | 94.5 | 61.7 | 119 | |
| EP075(SIM): Benzo(k)fluoranthene 207 | -08-9 | 1 | μg/L | <1.0 | 5 μg/L | 85.9 | 63.0 | 115 | |
| EP075(SIM): Benzo(a)pyrene 50 | -32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 96.1 | 63.3 | 117 | |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | -39-5 | 1 | μg/L | <1.0 | 5 μg/L | 96.5 | 59.9 | 118 | |
| EP075(SIM): Dibenz(a.h)anthracene 53 | -70-3 | 1 | μg/L | <1.0 | 5 μg/L | 96.5 | 61.2 | 117 | |
| EP075(SIM): Benzo(g.h.i)perylene | -24-2 | 1 | μg/L | <1.0 | 5 μg/L | 94.8 | 59.1 | 118 | |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5588826) | | | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 73.9 | 53.7 | 97.0 | |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 85.6 | 63.3 | 107 | |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 98.5 | 58.3 | 120 | |

Page : 7 of 9
Work Order : ES2403942

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|----------------------|---------------|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5 | 593731) | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 86.0 | 75.0 | 127 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5 | 596095) | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 104 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM | 2013 Fractions (QC | Lot: 5588826) | | | | | | |
| EP071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 64.5 | 53.9 | 95.5 |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 78.6 | 57.8 | 110 |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 74.1 | 50.5 | 115 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM | 2013 Fractions (QC | Lot: 5593731) | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 79.2 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM | 2013 Fractions (QC | Lot: 5596095) | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 95.4 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 5593731) | 11 11 11 | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 113 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 95.6 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 96.5 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | 10 μg/L | 100 | 73.0 | 122 |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 99.8 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 93.2 | 75.5 | 124 |
| EP080: BTEXN (QCLot: 5596095) | 11 11 | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 116 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 111 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 105 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | 10 μg/L | 88.8 | 73.0 | 122 |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 102 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 113 | 75.5 | 124 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | | Ма | trix Spike (MS) Repor | t | |
|----------------------|-----------|------------------|------------|---------------|-----------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable l | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |

Page : 8 of 9
Work Order : ES2403942



| Sub-Matrix: WATER | | | | Ma | atrix Spike (MS) Report | t | |
|----------------------|--|--------------------------------------|------------|---------------|-------------------------|--------------|-----------|
| | | | | Spike | SpikeRecovery(%) | Acceptable L | imits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG005(ED093)F: D | issolved Metals by ICP-AES (QCLot: 5590163) | | | | | | |
| ES2403761-001 | Anonymous | EG005F: Manganese | 7439-96-5 | 1 mg/L | 108 | 70.0 | 130 |
| EG020F: Dissolved | Metals by ICP-MS (QCLot: 5590161) | | | | | | |
| ES2403761-003 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 116 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 104 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 102 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 122 | 70.0 | 130 |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 81.1 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 113 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | # Not | 70.0 | 130 |
| EC025E: Discolves | Morough by EIMS (OCL et: EE004E0) | | | | Determined | | |
| ES2403595-003 | Mercury by FIMS (QCLot: 5590159) Anonymous | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 92.0 | 70.0 | 130 |
| | | 2000 moreary | 1439-91-0 | 0.01 Hig/L | 92.0 | 70.0 | 130 |
| | us Nitrate as N (NOx) by Discrete Analyser(QCLot: 55 | | | | | | |
| ES2403912-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | # Not Determined | 70.0 | 130 |
| EK061G: Total Kjel | dahl Nitrogen By Discrete Analyser (QCLot: 5594699) | | | | | | |
| ES2403860-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 50 mg/L | 105 | 70.0 | 130 |
| EK067G: Total Pho | osphorus as P by Discrete Analyser (QCLot: 5594700) | | | | | | |
| ES2403868-001 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | # Not | 70.0 | 130 |
| | | | | | Determined | | |
| EP080/071: Total P | etroleum Hydrocarbons (QCLot: 5593731) | | | | | | |
| ES2403857-007 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 94.9 | 70.0 | 130 |
| EP080/071: Total P | etroleum Hydrocarbons (QCLot: 5596095) | | | | | | |
| ES2403888-002 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 77.8 | 70.0 | 130 |
| EP080/071: Total R | ecoverable Hydrocarbons - NEPM 2013 Fractions (QC | Lot: 5593731) | | | | | |
| ES2403857-007 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 90.1 | 70.0 | 130 |
| EP080/071: Total R | ecoverable Hydrocarbons - NEPM 2013 Fractions (QC | Lot: 5596095) | | | | | |
| ES2403888-002 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 71.4 | 70.0 | 130 |
| EP080: BTEXN (Q | CLot: 5593731) | | | | | | |
| ES2403857-007 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 111 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 93.1 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 93.9 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 97.0 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | El coci ciale rijisho | 95-47-6 | 25 μg/L | 95.6 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 96.1 | 70.0 | 130 |

Page : 9 of 9 Work Order : ES2403942



| Sub-Matrix: WATER | | | | M | atrix Spike (MS) Repor | t | |
|----------------------|-----------------|----------------------------|------------|---------------|------------------------|------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP080: BTEXN (C | (CLot: 5596095) | | | | | | |
| ES2403888-002 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 85.5 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 78.3 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 78.2 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 77.9 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 72.8 | 70.0 | 130 |
| | | FP080: Naphthalene | 91-20-3 | 25 μg/L | 77.8 | 70.0 | 130 |

COC_RF_GW sampling.xlsx

Subconforward Lab / Split WO

Chain of Custody อิคุรมากอกtation

Senversa

4119

Comments: e.g. Highly contaminated sample; hazardous materials present, trace Work Order Reference ES2404239 Environmental Division Sydney Please forward to Eurofins Telephone: +61-2-8784 8555 LORs etc. Date: Praw EG005F (FE AND MN) Attached By PO / Internal Signature: Connote / Courier: 8-TN NO NO: Rowan Faint W-26 (TRH/BTEX/PAH/8 METALS) W-18 (TRH/BTEXN) Sampler Name: Total Bottles 44 9 9 9 9 9 9 0408038593, 0404011544 Container Information of 1 Standard 7 Days P, VS x2, N, UA, VSA Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project Type / Code EN/103/21 VOA VOA Sample Receipt ALS NSW Turn Around Time: Purchase Order Phone/Mobile: Time Laboratory: Address: Contact: Quote No: Phone: Page: 9/02/2024 9/02/2024 9/02/2024 9/02/2024 9/02/2024 9/02/2024 9/02/2024 9/02/2024 9/02/2024 rowan.faint@senversa.com.au Date Wetherill Park WME Emma Walsh Rowan Faint specifications were used during the collection of these samples: Sample Information Matrix * 3 3 3 3 3 3 3 3 > Sample ID QC103 QC203 QC404 QC504 QC303 MW2 MW3 MW6 MW1 ABN 89 132 231 380 Senversa Pty Ltd Email Report To: Project Manager Project Name: Sampled By: Job Number: Lab ID 9 55 W Total

| Polinguished Rv. | | | [Method of Shipment (if applicable): | Keceived by: | |
|--|--------------|--|---|--|--|
| | 7 C | Doto: 0/0/07 | Carrier / Deference # | Name/Signature: ACC / 1/2 | Date: 4/2/24 |
| Name/Signature: | Rowall Faint | Date: 3/2/2+ | 51 | | 0000 |
|) UE | | Time: 3:30 PM | Date/Time: | | Time: (6.50 |
| · · | | THE RESERVE THE PERSON NAMED IN COLUMN TWO IS NOT THE PERSON NAMED IN TRANSPORT OF THE PERSON NAMED IN THE P | CE COMPANION DE CONTRACTOR DE | | |
| Nome/Signature | | Date: | Carrier / Reference #: | Name/Signature: | Date: |
| Tallio Ognanie. | | | | | i |
| Of | | Time: | Date/Time: | Of: | Time: |
| NAME AND ADDRESS OF THE OWNER, OF TAXABLE PARTY OF TAXABL | | THE RESERVE THE PROPERTY OF TH | | | |
| Namo/Signafilia. | | Date: | Carrier / Reference #: | Name/Signature: | Date: |
| Tall of Olymonic. | | | | | i |
| j. | | Time: | Date/Time: | Of: | Time: |
| | | CONTRACTOR DESCRIPTION OF STREET, STRE | | the state of the s | The state of the s |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; N = Sulphuric Preserved (HO) Preserved; VS = Sulphuric Preserved Share Slate (HO) Preserved (HO) Preserved (HO) Preserved Plastic; HO) Preserved (HO) Preserved Plastic; N = Sulphuric Preserved Marker (HO) Preserved (HO) Preserved Marker (HO) Preserved Marker (HO) Preserved Marker (HO) Preserved (HO) Preserved Marker (HO) Preserved Mark

Completed by:



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2404239

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : sandy.phan@alsglobal.com

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555 Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 3

 Order number
 : --- Quote number
 : EB2023SENVER0001 (EN/000)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Sampler : Rowan Faint

Dates

Date Samples Received : 09-Feb-2024 16:30 Issue Date : 10-Feb-2024 Client Requested Due : 15-Feb-2024 Scheduled Reporting Date : 15-Feb-2024

Date

Delivery Details

Mode of Delivery : Client Drop Off Security Seal : Not Available

No. of coolers/boxes : 2 Temperature : 12.0'C, 13.6'C, 9.7'C - Ice

present

Receipt Detail : Large Esky No. of samples received / analysed : 8 / 8

General Comments

This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- Sample QC203 to be forwarded to Eurofins for analysis.
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 10-Feb-2024 Issue Date

Page

: 2 of 3 : ES2404239 Amendment 0 Work Order : SENVERSA PTY LTD Client



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Any sample identifications that cannot be displayed entirely in the analysis summary table will be listed below.

ES2404239-002 : [05-Feb-2024] : QC504 - Trip Spike 13

Summary of Sample(s) and Requested Analysis

| process necessar tasks. Packages as the determinates, that are included for no sampling default 00:00 on | ary for the execution and contain and contain and contain of moisture uded in the package. Itime is provided, the date of sampling date with the contained the contained contained the contained con | be part of a laboratory on of client requested ditional analyses, such content and preparation the sampling time will g. If no sampling date ll be assumed by the ckets without a time | EG005F Metals by ICPAES | EP080 | NT-08 ogen + NO2 + NO3 + NH3 + Total P | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - W-18 TRH(C6 - C9)/BTEXN | NATER - W-26 IRH/BTEXN/PAH/8 Metals |
|--|--|--|----------------------------|------------------|---|--|------------------------------------|--|
| Laboratory sample | Sampling date / | Sample ID | WATER - Dissolved | WATER - BTEXN | WATER - NT-08 Total Nitrogen + | WATER - NT-11 Total Nitrogen a | WATER - W-18 TRH(C6 - C9)/B | WATER - W-26 TRH/BTEXN/PA |
| ES2404239-001 | 05-Feb-2024 00:00 | QC404 Trip Blank | | | | | 1 | |
| ES2404239-002 | 05-Feb-2024 00:00 | QC504 Trip Spike 13 | | ✓ | | | | |
| ES2404239-003 | 09-Feb-2024 00:00 | QC303 | | | | 1 | | ✓ |
| ES2404239-004 | 09-Feb-2024 00:00 | MW1 | ✓ | | ✓ | | | ✓ |
| ES2404239-005 | 09-Feb-2024 00:00 | MW2 | 1 | | 1 | | | ✓ |
| ES2404239-006 | 09-Feb-2024 00:00 | MW3 | ✓ | | ✓ | | | ✓ |
| ES2404239-007 | 09-Feb-2024 00:00 | MW6 | 1 | | ✓ | | | ✓ |
| ES2404239-008 | 09-Feb-2024 00:00 | QC103 | 1 | | | ✓ | | ✓ |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

: 10-Feb-2024 Issue Date

Page

3 of 3 ES2404239 Amendment 0 Work Order Client : SENVERSA PTY LTD



Requested Deliverables

| BE | | | |
|----|--|--|--|
| | | | |

SUPPLIER ACCOUNTS - A4 - AU Tax Invoice (INV)

| BEC CHAPPLE | | |
|---|-------|-----------------------------|
| - *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
| *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| EMMA WALSH | | |
| *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
| *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Tax Invoice (INV) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| Rowan Faint | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | rowan.faint@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | rowan.faint@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | rowan.faint@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | rowan.faint@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | rowan.faint@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | rowan.faint@senversa.com.au |
| | | |

Email

supplieraccounts@senversa.com.a



CERTIFICATE OF ANALYSIS

Work Order : ES2404239

Client : SENVERSA PTY LTD

Contact : EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number : ----

Sampler : Rowan Faint

Site : ---

C-O-C number

Quote number : EN/000

No. of samples received : 8
No. of samples analysed : 8

Page : 1 of 10

Laboratory : Environmental Division Sydney

Contact : Sandy Phan

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61-2-8784 8555

Date Samples Received : 09-Feb-2024 16:30

Date Analysis Commenced : 10-Feb-2024

Issue Date : 15-Feb-2024 18:29

Sydney Organics, Smithfield, NSW



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

Edwandy Fadjar

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW

Organic Coordinator

Page : 2 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a.h)anthracene (1.0), Benzo(g.h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- EG020: LORs have been raised for some samples due to matrix interference (High sample salinity)
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.

Page : 3 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC404 Trip Blank | QC504 Trip Spike 13 | QC303 | MW1 | MW2 |
|---|-------------------|--------|----------------|---------------------|------------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2404239-001 | ES2404239-002 | ES2404239-003 | ES2404239-004 | ES2404239-005 |
| | | | | Result | Result | Result | Result | Result |
| EG005(ED093)F: Dissolved Metals by IC | | 0.05 | | | | | | 2.51 |
| Iron | 7439-89-6 | 0.05 | mg/L | | | | 3.96 | 0.54 |
| Manganese | 7439-96-5 | 0.01 | mg/L | | | | 0.59 | 1.76 |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | | | <0.001 | 0.012 | 0.004 |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | | | <0.0001 | <0.0001 | <0.0001 |
| Chromium | 7440-47-3 | 0.001 | mg/L | | | <0.001 | <0.001 | <0.001 |
| Copper | 7440-50-8 | 0.001 | mg/L | | | <0.001 | <0.001 | <0.001 |
| Lead | 7439-92-1 | 0.001 | mg/L | | | <0.001 | <0.001 | <0.001 |
| Nickel | 7440-02-0 | 0.001 | mg/L | | | <0.001 | 0.015 | 0.006 |
| Zinc | 7440-66-6 | 0.005 | mg/L | | | <0.005 | 0.016 | 0.009 |
| EG035F: Dissolved Mercury by FIMS | | 4 | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | | | <0.0001 | <0.0001 | <0.0001 |
| EK055G: Ammonia as N by Discrete Ana | alyser | | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | | | | 0.48 | 0.44 |
| EK057G: Nitrite as N by Discrete Analys | ser | | | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | | | | <0.01 | <0.01 |
| EK058G: Nitrate as N by Discrete Analy | ser | | | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | | | | <0.01 | <0.01 |
| EK059G: Nitrite plus Nitrate as N (NOx) | by Discrete Ana | lyser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | | | <0.01 | <0.01 | <0.01 |
| EK061G: Total Kjeldahl Nitrogen By Disc | crete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | | | <0.1 | 0.6 | 0.8 |
| EK062G: Total Nitrogen as N (TKN + NO | x) by Discrete Ar | alyser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | | | <0.1 | 0.6 | 0.8 |
| EK067G: Total Phosphorus as P by Disc | crete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | | | 0.02 | <0.01 | 0.05 |
| EP075(SIM)B: Polynuclear Aromatic Hyd | drocarbons | | | | | | | |

Page : 4 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC404 Trip Blank | QC504 Trip Spike 13 | QC303 | MW1 | MW2 |
|--|-------------------|------------|----------------|---------------------|------------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2404239-001 | ES2404239-002 | ES2404239-003 | ES2404239-004 | ES2404239-005 |
| | | | | Result | Result | Result | Result | Result |
| EP075(SIM)B: Polynuclear Aromatic Hy | drocarbons - Cont | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Fluorene | 86-73-7 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Anthracene | 120-12-7 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Pyrene | 129-00-0 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Chrysene | 218-01-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | | | <1.0 | <1.0 | <1.0 |
| ^ Sum of polycyclic aromatic hydrocarbon | s | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | | | <0.5 | <0.5 | <0.5 |
| EP080/071: Total Petroleum Hydrocarb | ons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | | <20 | <20 | <20 |
| C10 - C14 Fraction | | 50 | μg/L | | | <50 | <50 | <50 |
| C15 - C28 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| C29 - C36 Fraction | | 50 | μg/L | | | <50 | <50 | <50 |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | | | <50 | <50 | <50 |
| EP080/071: Total Recoverable Hydroca | rbons - NEPM 201 | 3 Fraction | | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | | <20 | <20 | <20 |

Page : 5 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC404 Trip Blank | QC504 Trip Spike 13 | QC303 | MW1 | MW2 |
|---|-------------------|--------|----------------|---------------------|------------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2404239-001 | ES2404239-002 | ES2404239-003 | ES2404239-004 | ES2404239-005 |
| | | | | Result | Result | Result | Result | Result |
| EP080/071: Total Recoverable Hydrod | | | | | | | | |
| ^ C6 - C10 Fraction minus BTEX | C6_C10-BTEX | 20 | μg/L | <20 | | <20 | <20 | <20 |
| (F1) >C10 - C16 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| >C16 - C34 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| >C34 - C40 Fraction | | 100 | μg/L | | | <100 | <100 | <100 |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | | | <100 | <100 | <100 |
| >C10 - C16 Fraction minus Naphthalene (F2) | e | 100 | μg/L | | | <100 | <100 | <100 |
| EP080: BTEXN | | - G | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | 15 | <1 | <1 | <1 |
| Toluene | 108-88-3 | 2 | μg/L | <2 | 16 | <2 | <2 | <2 |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 16 | <2 | <2 | <2 |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | 17 | <2 | <2 | <2 |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 17 | <2 | <2 | <2 |
| ^ Total Xylenes | | 2 | μg/L | <2 | 34 | <2 | <2 | <2 |
| ^ Sum of BTEX | | 1 | μg/L | <1 | 81 | <1 | <1 | <1 |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | 18 | <5 | <5 | <5 |
| EP075(SIM)S: Phenolic Compound St | urrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | | | 32.7 | 28.9 | 28.0 |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | | | 63.8 | 61.2 | 56.1 |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | | | 79.8 | 85.0 | 79.5 |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | | | 70.6 | 67.4 | 67.7 |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | | | 64.0 | 65.1 | 59.5 |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | | | 85.8 | 87.7 | 76.2 |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 78.3 | 89.0 | 90.2 | 90.0 | 87.5 |
| Toluene-D8 | 2037-26-5 | 2 | % | 86.6 | 96.6 | 95.2 | 97.5 | 96.8 |

Page : 6 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | Sample ID | | QC404 | QC504 | QC303 | MW1 | MW2 | |
|----------------------------------|-----------|----------------|-------------------|-------------------------------------|---------------|-------------------|-------------------|---------------|--|
| (Matrix: WATER) | | | | Trip Blank | Trip Spike 13 | | | | |
| Sampling date / tin | | ng date / time | 05-Feb-2024 00:00 | 05-Feb-2024 00:00 09-Feb-2024 00:00 | | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | | |
| Compound CAS Number LO | | LOR | Unit | ES2404239-001 | ES2404239-002 | ES2404239-003 | ES2404239-004 | ES2404239-005 | |
| | | | | Result | Result | Result | Result | Result | |
| EP080S: TPH(V)/BTEX Surrogates - | Continued | | | | | | | | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 112 | 125 | 126 | 127 | 125 | |

Page : 7 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| (Matrix: WATER) | | | Sample ID | MW3 | MW6 | QC103 | |
|---------------------------------------|----------------------|--------|----------------|-------------------|-------------------|-------------------|------|
| | | Sampli | ng date / time | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2404239-006 | ES2404239-007 | ES2404239-008 | |
| | | | | Result | Result | Result | |
| EG005(ED093)F: Dissolved Metals by | ICP-AES | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 8.01 | <0.05 | 8.08 | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 7.00 | 0.06 | 7.08 | |
| EG020F: Dissolved Metals by ICP-MS | 3 | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | <0.010 | 0.002 | <0.010 | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0010 | <0.0001 | <0.0010 | |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.010 | <0.001 | <0.010 | |
| Copper | 7440-50-8 | 0.001 | mg/L | <0.010 | <0.001 | <0.010 | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.010 | <0.001 | <0.010 | |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.191 | 0.001 | 0.197 | |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.247 | <0.005 | 0.253 | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0001 | |
| EK055G: Ammonia as N by Discrete | Analyser | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.29 | <0.01 | | |
| EK057G: Nitrite as N by Discrete Ana | alyser | | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | | |
| EK058G: Nitrate as N by Discrete An | alyser | | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | 0.03 | 1.93 | | |
| EK059G: Nitrite plus Nitrate as N (NC | Ox) by Discrete Anal | lyser | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.03 | 1.93 | 0.02 | |
| EK061G: Total Kjeldahl Nitrogen By I | Discrete Analyser | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.9 | 1.0 | 0.9 | |
| EK062G: Total Nitrogen as N (TKN + | NOx) by Discrete An | alyser | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 0.9 | 2.9 | 0.9 | |
| EK067G: Total Phosphorus as P by D | Discrete Analyser | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.08 | 0.03 | 0.09 | |
| EP075(SIM)B: Polynuclear Aromatic l | Hydrocarbons | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | |

Page : 8 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW6 | QC103 | | |
|---|-------------------|------------|----------------|-------------------|-------------------|-------------------|-------------|--|
| (massi tortal) | | Samplii | ng date / time | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2404239-006 | ES2404239-007 | ES2404239-008 | | |
| | | | | Result | Result | Result | | |
| EP075(SIM)B: Polynuclear Aromatic Hyd | | | | | | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | | |
| Sum of polycyclic aromatic hydrocarbons | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | | |
| EP080/071: Total Petroleum Hydrocarbo | ons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | <50 | | |
| C15 - C28 Fraction | | 100 | μg/L | <100 | <100 | <100 | | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | <50 | | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | <50 | <50 | | |
| EP080/071: Total Recoverable Hydrocar | rbons - NEPM 201 | 3 Fraction | ıs | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | | |
| ^ C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | | |

Page : 9 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW3 | MW6 | QC103 | |
|--|-------------------|--------|----------------|-------------------|-------------------|-------------------|------|
| | | Sampli | ng date / time | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | 09-Feb-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2404239-006 | ES2404239-007 | ES2404239-008 | |
| | | | | Result | Result | Result | |
| EP080/071: Total Recoverable Hydroc | arbons - NEPM 201 | | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | <100 | |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | <100 | <100 | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | <100 | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | <100 | <100 | |
| ^ >C10 - C16 Fraction minus Naphthalene (F2) | | 100 | μg/L | <100 | <100 | <100 | |
| EP080: BTEXN | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | |
| EP075(SIM)S: Phenolic Compound Su | urrogates | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 29.1 | 25.9 | 24.8 | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 58.1 | 60.0 | 50.4 | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 75.2 | 90.8 | 65.4 | |
| EP075(SIM)T: PAH Surrogates | | 12 | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 65.2 | 73.3 | 57.3 | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 76.4 | 65.3 | 64.2 | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 74.0 | 84.9 | 66.0 | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 80.2 | 89.0 | 90.8 | |
| Toluene-D8 | 2037-26-5 | 2 | % | 89.2 | 102 | 94.3 | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 114 | 127 | 127 | |

Page : 10 of 10 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery | Limits (%) |
|---|------------|----------|------------|
| Compound | CAS Number | Low | High |
| EP075(SIM)S: Phenolic Compound Surrogates | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 |
| EP075(SIM)T: PAH Surrogates | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 |
| Anthracene-d10 | 1719-06-8 | 27 | 113 |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 |
| EP080S: TPH(V)/BTEX Surrogates | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 |
| Toluene-D8 | 2037-26-5 | 75 | 131 |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2404239** Page : 1 of 8

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

 Contact
 : EMMA WALSH
 Telephone
 : +61-2-8784 8555

 Project
 : S20102 Wetherill Park WME
 Date Samples Received
 : 09-Feb-2024

 Site
 : --- Issue Date
 : 15-Feb-2024

Sampler : Rowan Faint No. of samples received : 8
Order number : ---- No. of samples analysed : 8

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- Matrix Spike outliers exist please see following pages for full details.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

• NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME

ALS

Outliers: Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: WATER

| | Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|---|---|----------------------|------------------|--------------|------------|------------|--------|----------------------------------|
| M | atrix Spike (MS) Recoveries | | | | | | | |
| | EK055G: Ammonia as N by Discrete Analyser | ES2403164001 | Anonymous | Ammonia as N | 7664-41-7 | Not | | MS recovery not determined, |
| | | | | | | Determined | | background level greater than or |
| | | | | | | | | equal to 4x spike level. |

Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Matrix: WATER | | | | | | |
|-----------------------------|------------|----|---------|--------|----------|--------------------------------|
| Quality Control Sample Type | | | Count | | e (%) | Quality Control Specification |
| Analytical Methods | Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 9 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 9 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 9 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 9 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive <u>or</u> Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: **x** = Holding time breach; ✓ = Within holding time.

| Method | Sample Date | Ex | traction / Preparation | | Analysis | | | |
|---|-------------|-------------|------------------------|--------------------|------------|---------------|------------------|------------|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG005(ED093)F: Dissolved Metals by ICP-AES | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG005F) | | | | | | | | |
| MW1, | MW2, | 09-Feb-2024 | | | | 13-Feb-2024 | 07-Aug-2024 | ✓ |
| MW3, | MW6, | | | | | | | |
| QC103 | | | | | | | | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | | | | 12-Feb-2024 | 07-Aug-2024 | ✓ |
| MW2, | MW3, | | | | | | | |
| MW6, | QC103 | | | | | | | |

Page : 3 of 8
Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Method | Method | | | traction / Preparation | | Analysis | | | |
|---|--------------|-------------|----------------|------------------------|------------|---------------|------------------|------------|--|
| Container / Client Sample ID(s) | | Sample Date | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (I | EG035F) | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | | | | 13-Feb-2024 | 08-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EK055G: Ammonia as N by Discrete Analy | | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK0550 | | | | | | | | | |
| MW1, | MW2, | 09-Feb-2024 | | | | 14-Feb-2024 | 08-Mar-2024 | ✓ | |
| MW3, | MW6 | | | | | | | | |
| EK057G: Nitrite as N by Discrete Analyser | | | | | | | | | |
| Clear Plastic Bottle - Natural (EK057G) | | | | | | | | | |
| MW1, | MW2, | 09-Feb-2024 | | | | 10-Feb-2024 | 11-Feb-2024 | ✓ | |
| MW3, | MW6 | | | | | | | | |
| EK059G: Nitrite plus Nitrate as N (NOx) b | | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK0590 | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | | | | 14-Feb-2024 | 08-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen By Discre | ete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK0610 | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 14-Feb-2024 | 08-Mar-2024 | ✓ | 14-Feb-2024 | 08-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EK067G: Total Phosphorus as P by Discre | | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK0670 | | | | 00.14 | | | 00.14 | | |
| QC303, | MW1, | 09-Feb-2024 | 14-Feb-2024 | 08-Mar-2024 | ✓ | 14-Feb-2024 | 08-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EP075(SIM)B: Polynuclear Aromatic Hydro | | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075) | • ** | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 12-Feb-2024 | 16-Feb-2024 | ✓ | 15-Feb-2024 | 23-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |

Page : 4 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Watth: WATER | | Evaluation: - Floriding time breach; Wi | | | | | | | |
|--|-----------------------|---|----------------|------------------------|------------|---------------|------------------|------------|--|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EP080/071: Total Petroleum Hydrocarbons | | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 12-Feb-2024 | 16-Feb-2024 | ✓ | 14-Feb-2024 | 23-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC404 - Trip Blank | | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | ✓ | 13-Feb-2024 | 19-Feb-2024 | ✓ | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 13-Feb-2024 | 23-Feb-2024 | ✓ | 13-Feb-2024 | 23-Feb-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EP080/071: Total Recoverable Hydrocarbons - NE | PM 2013 Fractions | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 12-Feb-2024 | 16-Feb-2024 | ✓ | 14-Feb-2024 | 23-Mar-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC404 - Trip Blank | | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | ✓ | 13-Feb-2024 | 19-Feb-2024 | ✓ | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 13-Feb-2024 | 23-Feb-2024 | ✓ | 13-Feb-2024 | 23-Feb-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |
| EP080: BTEXN | | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC404 - Trip Blank, | QC504 - Trip Spike 13 | 05-Feb-2024 | 13-Feb-2024 | 19-Feb-2024 | ✓ | 13-Feb-2024 | 19-Feb-2024 | ✓ | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | | |
| QC303, | MW1, | 09-Feb-2024 | 13-Feb-2024 | 23-Feb-2024 | ✓ | 13-Feb-2024 | 23-Feb-2024 | ✓ | |
| MW2, | MW3, | | | | | | | | |
| MW6, | QC103 | | | | | | | | |

Page : 5 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

ne expected rate. A listing of breaches is provided in the Summary of Outliers.

| Matrix: WATER Evaluation: × = Quality Control free | | | | | | ntrol frequency r | not within specification; ✓ = Quality Control frequency within specification. |
|---|------------|----|---------|--------|----------|-------------------|---|
| Quality Control Sample Type | | Co | ount | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 5 | 20.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 19 | 10.53 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 9 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 4 | 37 | 10.81 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 9 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 20 | 15.00 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 6 | 37 | 16.22 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 19 | 5.26 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 37 | 5.41 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |

Page : 6 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: * = Quality Control frequency not within specification; * = Quality Control frequency within specification. Quality Control Sample Type Count Rate (%) Quality Control Specification Analytical Methods Method QC Evaluation Regular Actual Expected Matrix Spikes (MS) - Continued Ammonia as N by Discrete analyser 20 5.00 NEPM 2013 B3 & ALS QC Standard EK055G 1 5.00 1 Dissolved Mercury by FIMS 1 20 NEPM 2013 B3 & ALS QC Standard 5.00 5.00 EG035F 1 5 Dissolved Metals by ICP-AES 1 20.00 5.00 NEPM 2013 B3 & ALS QC Standard EG005F ✓ Dissolved Metals by ICP-MS - Suite A 1 20 NEPM 2013 B3 & ALS QC Standard EG020A-F 5.00 5.00 1 Nitrite and Nitrate as N (NOx) by Discrete Analyser EK059G 1 19 5.26 5.00 1 NEPM 2013 B3 & ALS QC Standard Nitrite as N by Discrete Analyser 1 20 NEPM 2013 B3 & ALS QC Standard 5.00 5.00 EK057G ✓ PAH/Phenols (GC/MS - SIM) 0 9 0.00 5.00 NEPM 2013 B3 & ALS QC Standard EP075(SIM) × Total Kjeldahl Nitrogen as N By Discrete Analyser 1 20 5.00 NEPM 2013 B3 & ALS QC Standard EK061G 5.00 1 Total Phosphorus as P By Discrete Analyser 2 37 5.41 5.00 1 NEPM 2013 B3 & ALS QC Standard EK067G TRH - Semivolatile Fraction 0 9 0.00 NEPM 2013 B3 & ALS QC Standard EP071 5.00 × TRH Volatiles/BTEX EP080 1 20 5.00 5.00 NEPM 2013 B3 & ALS QC Standard

Page : 7 of 8
Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|---|----------|--------|---|
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered samples, emitting a characteristic spectrum which is compared against matrix matched standards. This method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Ammonia as N by Discrete analyser | EK055G | WATER | In house: Referenced to APHA 4500-NH3 G Ammonia is determined by direct colorimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrite as N by Discrete Analyser | EK057G | WATER | In house: Referenced to APHA 4500-NO2- B. Nitrite is determined by direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrate as N by Discrete Analyser | EK058G | WATER | In house: Referenced to APHA 4500-NO3- F. Nitrate is reduced to nitrite by way of a chemical reduction followed by quantification by Discrete Analyser. Nitrite is determined seperately by direct colourimetry and result for Nitrate calculated as the difference between the two results. This method is compliant with NEPM Schedule B(3) |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |

Page : 8 of 8 Work Order : ES2404239



| Analytical Methods | Method | Matrix | Method Descriptions |
|--------------------------------------|-----------------------|------------------------|--|
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode |
| | | | and quantification is by comparison against an established 5 point calibration curve. This method is compliant |
| | | | with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary |
| | | | GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a |
| | | | sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This |
| | | | method is compliant with the QC requirements of NEPM Schedule B(3) |
| | | | |
| Preparation Methods | Method | Matrix | Method Descriptions |
| Preparation Methods TKN/TP Digestion | Method EK061/EK067 | <i>Matrix</i> WATER | Method Descriptions In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| | | 11 | · |
| | | 11 | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated |



QUALITY CONTROL REPORT

Work Order : **ES2404239** Page : 1 of 8

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555

Project : \$20102 Wetherill Park WME Date Samples Received : 09-Feb-2024
Order number Date Analysis Commenced : 10-Feb-2024

C-O-C number ---- Issue Date 15-Feb-2024

Sampler : Rowan Faint

Site : ---Quote number : EN/000

No. of samples received : 8

No. of samples analysed : 8

Organic Coordinator

Accreditation No. 825
Accredited for compliance with
ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

Sydney Organics, Smithfield, NSW

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

Edwandy Fadjar

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW

Page : 2 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit: Result between 10 and 20 times LOR: 0% - 50%: Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | Laboratory Duplicate (DUP) Report | | | | |
|--|----------------------|-------------------------|------------|--------------|-----------------------------------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | solved Metals by ICF | P-AES (QC Lot: 5595964) | | | | | | | |
| ES2404239-004 | MW1 | EG005F: Manganese | 7439-96-5 | 0.01 (0.10)* | mg/L | 0.59 | 0.60 | 2.5 | No Limit |
| | | EG005F: Iron | 7439-89-6 | 0.05 (0.10)* | mg/L | 3.96 | 4.09 | 3.4 | 0% - 20% |
| EG020F: Dissolved Metals by ICP-MS (QC Lot: 5595960) | | | | | | | | | |
| ES2404140-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 0.022 | 0.021 | 7.0 | No Limit |
| ES2403778-007 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.001 | 0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| EG035F: Dissolved I | Mercury by FIMS (Q | C Lot: 5595963) | | | | | | | |
| ES2403998-007 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| ES2404239-003 | QC303 | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |

Page : 3 of 8
Work Order : ES2404239



| Sub-Matrix: WATER | | | | Laboratory Duplicate (DUP) Report | | | | | |
|----------------------|------------------------------|--------------------------------------|---------------------|-----------------------------------|--------------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EK055G: Ammonia a | s N by Discrete Analyser(C | (C Lot: 5599208) | | | | | | | |
| ES2403164-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 (1.00)* | mg/L | 947 | 992 | 4.7 | 0% - 20% |
| ES2404239-004 | MW1 | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.48 | 0.47 | 0.0 | 0% - 20% |
| EK057G: Nitrite as N | N by Discrete Analyser (QC | Lot: 5594300) | | | | | | | |
| ES2404227-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| ES2404102-002 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK059G: Nitrite plus | Nitrate as N (NOx) by Disc | rete Analyser (QC Lot: 5599209) | | | | | | | |
| ES2404030-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.01 | <0.01 | 0.0 | No Limit |
| ES2404239-004 | MW1 | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK061G: Total Kjeld | ahl Nitrogen By Discrete Ana | alyser (QC Lot: 5599213) | | | | | | | |
| ES2404024-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (0.5)* | mg/L | 37.1 | 38.0 | 2.5 | 0% - 20% |
| ES2404239-004 | MW1 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.6 | 0.6 | 0.0 | No Limit |
| EK067G: Total Phos | phorus as P by Discrete Ana | llyser (QC Lot: 5599212) | | | | | | | |
| ES2404157-007 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 1.07 | 1.04 | 2.6 | 0% - 20% |
| ES2404024-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 (0.05)* | mg/L | 6.01 | 5.89 | 2.1 | 0% - 20% |
| EK067G: Total Phos | phorus as P by Discrete Ana | llyser (QC Lot: 5599214) | | | | | | | |
| ES2404239-004 | MW1 | EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 0.02 | 74.2 | No Limit |
| ES2404300-007 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbons (QC | Lot: 5596119) | | | | | | | |
| ES2403922-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2404239-004 | MW1 | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Re | coverable Hydrocarbons - N | EPM 2013 Fractions (QC Lot: 5596119) | | | | | | | |
| ES2403922-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2404239-004 | MW1 | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5596119) | | | | | | | | |
| ES2403922-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| ES2404239-004 | MW1 | EP080: Benzene | 71-43-2 | 1 | μg/L " | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: ortho-Xylene | 106-42-3 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | LF 000. Ortilo-Aylette | 35 47-0 | | μg/ L | | | 0.0 | 140 Lillin |

Page : 4 of 8 Work Order : ES2404239



| Sub-Matrix: WATER | | | | | Laboratory Duplicate (DUP) Report | | | | | |
|----------------------|--------------------------|--------------------|------------|-----|-----------------------------------|-----------------|------------------|---------|--------------------|--|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) | |
| EP080: BTEXN (QC L | ot: 5596119) - continued | | | | | | | | | |
| ES2404239-004 | MW1 | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit | |

Page : 5 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Method: Compound | | | | | | | Control Spike (LCS) Report | | |
|---|------------------|--------|------|---------|---------------|--------------------|----------------------------|------------|--|
| Method: Compound | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 5 | | | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 109 | 82.0 | 114 | |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 104 | 81.0 | 113 | |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5595960) | | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 108 | 85.0 | 114 | |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 107 | 84.0 | 110 | |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 106 | 85.0 | 111 | |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.6 | 81.0 | 111 | |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 103 | 83.0 | 111 | |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 106 | 82.0 | 112 | |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 110 | 81.0 | 117 | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5595963) | | | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 89.9 | 83.0 | 105 | |
| EK055G: Ammonia as N by Discrete Analyser (QCLot: 5 | 599208) | | | | | | | | |
| EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 97.0 | 90.0 | 114 | |
| EK057G: Nitrite as N by Discrete Analyser (QCLot: 5594 | 300) | | | | | | | | |
| EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 96.5 | 82.0 | 114 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Ana | lvser (QCLot: 55 | 99209) | | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 104 | 91.0 | 113 | |
| EK061G: Total Kieldahl Nitrogen By Discrete Analyser (0 | OCL of: 5599213) | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 89.7 | 69.0 | 101 | |
| , , | | | | <0.1 | 1 mg/L | 97.3 | 70.0 | 118 | |
| | | | | <0.1 | 5 mg/L | 106 | 70.0 | 130 | |
| EK067G: Total Phosphorus as P by Discrete Analyser (C | CLot: 5599212) | | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 92.7 | 71.3 | 126 | |
| · | | | | <0.01 | 0.442 mg/L | 96.4 | 71.3 | 126 | |
| | | | | <0.01 | 1 mg/L | 113 | 70.0 | 130 | |
| EK067G: Total Phosphorus as P by Discrete Analyser (C | QCLot: 5599214) | | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 87.9 | 71.3 | 126 | |
| | | | | <0.01 | 0.442 mg/L | 94.0 | 71.3 | 126 | |
| | | | | <0.01 | 1 mg/L | 112 | 70.0 | 130 | |

Page : 6 of 8
Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|---------------|------|-------------------|---------------|------------------------------|------------|------------|
| | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLot: 5594837) | | | | | | | |
| EP075(SIM): Naphthalene 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 67.2 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 81.5 | 63.6 | 114 |
| EP075(SIM): Acenaphthene 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 66.6 | 62.2 | 113 |
| EP075(SIM): Fluorene 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 78.4 | 63.9 | 115 |
| EP075(SIM): Phenanthrene 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 74.8 | 62.6 | 116 |
| EP075(SIM): Anthracene 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 73.2 | 64.3 | 116 |
| EP075(SIM): Fluoranthene 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 79.8 | 63.6 | 118 |
| EP075(SIM): Pyrene 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 82.5 | 63.1 | 118 |
| EP075(SIM): Benz(a)anthracene 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 76.2 | 64.1 | 117 |
| EP075(SIM): Chrysene 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 77.6 | 62.5 | 116 |
| EP075(SIM): Benzo(b+j)fluoranthene 205-99-2 205-82-3 | 1 | μg/L | <1.0 | 5 μg/L | 83.1 | 61.7 | 119 |
| EP075(SIM): Benzo(k)fluoranthene 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 68.3 | 63.0 | 115 |
| EP075(SIM): Benzo(a)pyrene 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 75.4 | 63.3 | 117 |
| EP075(SIM): Indeno(1.2.3.cd)pyrene 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 75.7 | 59.9 | 118 |
| EP075(SIM): Dibenz(a.h)anthracene 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 75.7 | 61.2 | 117 |
| EP075(SIM): Benzo(g.h.i)perylene | 1 | μg/L | <1.0 | 5 μg/L | 70.8 | 59.1 | 118 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5594836) | | | | | | | |
| EP071: C10 - C14 Fraction | 50 | μg/L | <50 | 400 μg/L | 63.5 | 53.7 | 97.0 |
| EP071: C15 - C28 Fraction | 100 | μg/L | <100 | 600 μg/L | 79.4 | 63.3 | 107 |
| EP071: C29 - C36 Fraction | 50 | μg/L | <50 | 400 μg/L | 79.8 | 58.3 | 120 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5596119) | | | | | | | |
| EP080: C6 - C9 Fraction | 20 | μg/L | <20 | 260 μg/L | 103 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC | Lot: 5594836) | | | | | | |
| EP071: >C10 - C16 Fraction | 100 | μg/L | <100 | 500 μg/L | 61.0 | 53.9 | 95.5 |
| EP071: >C16 - C34 Fraction | 100 | μg/L | <100 | 700 μg/L | 77.3 | 57.8 | 110 |
| EP071: >C34 - C40 Fraction | 100 | μg/L | <100 | 300 μg/L | 78.8 | 50.5 | 115 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC | <u> </u> | | | | | | |
| EP080: C6 - C10 Fraction C6_C10 | 20 | μg/L | <20 | 310 μg/L | 95.5 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 5596119) | | | | | | | |
| EP080: Benzene 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 101 | 68.3 | 119 |
| EP080: Toluene 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 111 | 73.5 | 120 |
| EP080: Ethylbenzene 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 107 | 73.8 | 122 |
| EP080: meta- & para-Xylene 108-38-3 106-42-3 | 2 | μg/L | <2 | 10 μg/L | 115 | 73.0 | 122 |

Page : 7 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|------------|-----|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP080: BTEXN (QCLot: 5596119) - continued | | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 108 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 97.2 | 75.5 | 124 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | Matrix Spike (MS) Report | | | | | | | |
|--|---|--------------------------------------|--------------------------|---------------|------------------|--------------|-----------|--|--|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable L | imits (%) | | | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | | | |
| EG005(ED093)F: D | Dissolved Metals by ICP-AES (QCLot: 5595964) | | | | | | | | | |
| ES2404239-005 | MW2 | EG005F: Manganese | 7439-96-5 | 1 mg/L | 100 | 70.0 | 130 | | | |
| EG020F: Dissolve | d Metals by ICP-MS (QCLot: 5595960) | | | | | | | | | |
| ES2404001-001 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 2 mg/L | 87.8 | 70.0 | 130 | | | |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.5 mg/L | 86.0 | 70.0 | 130 | | | |
| | | EG020A-F: Chromium | 7440-47-3 | 2 mg/L | 89.0 | 70.0 | 130 | | | |
| | | EG020A-F: Copper | 7440-50-8 | 2 mg/L | 81.5 | 70.0 | 130 | | | |
| | | EG020A-F: Lead | 7439-92-1 | 2 mg/L | 87.5 | 70.0 | 130 | | | |
| | | EG020A-F: Nickel | 7440-02-0 | 2 mg/L | 87.7 | 70.0 | 130 | | | |
| | | EG020A-F: Zinc | 7440-66-6 | 2 mg/L | 83.6 | 70.0 | 130 | | | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5595963) | | | | | | | | | | |
| ES2403888-013 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 88.7 | 70.0 | 130 | | | |
| EK055G: Ammoni | a as N by Discrete Analyser (QCLot: 5599208) | | | | | | | | | |
| ES2403164-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.5 mg/L | # Not | 70.0 | 130 | | | |
| | | | | | Determined | | | | | |
| EK057G: Nitrite a | s N by Discrete Analyser (QCLot: 5594300) | | | | | | | | | |
| ES2404102-002 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.5 mg/L | 101 | 70.0 | 130 | | | |
| EK059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCLot: 559 | 99209) | | | | | | | | |
| ES2404030-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 99.7 | 70.0 | 130 | | | |
| EK061G: Total Kje | eldahl Nitrogen By Discrete Analyser (QCLot: 5599213) | | | | | | | | | |
| ES2404030-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 5 mg/L | 105 | 70.0 | 130 | | | |
| EK067G: Total Ph | osphorus as P by Discrete Analyser (QCLot: 5599212) | | | | | | | | | |
| ES2404030-001 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 115 | 70.0 | 130 | | | |
| EK067G: Total Ph | osphorus as P by Discrete Analyser (QCLot: 5599214) | | | | | | | | | |
| ES2404300-008 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 115 | 70.0 | 130 | | | |

Page : 8 of 8 Work Order : ES2404239

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Matrix Spike (MS) Report | | | | |
|----------------------|---|----------------------------|------------|--------------------------|------------------|--------------|------------|--|
| | | | | Spike | SpikeRecovery(%) | Acceptable l | Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | |
| EP080/071: Total | Petroleum Hydrocarbons (QCLot: 5596119) | | | | | | | |
| ES2403922-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 83.5 | 70.0 | 130 | |
| EP080/071: Total | Recoverable Hydrocarbons - NEPM 2013 Fractions (QCL | ot: 5596119) | | | | | | |
| ES2403922-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 79.0 | 70.0 | 130 | |
| EP080: BTEXN (| QCLot: 5596119) | | | | | | | |
| ES2403922-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 94.1 | 70.0 | 130 | |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 96.3 | 70.0 | 130 | |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 99.4 | 70.0 | 130 | |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 106 | 70.0 | 130 | |
| | | | 106-42-3 | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 98.2 | 70.0 | 130 | |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 91.6 | 70.0 | 130 | |

| Documentation | |
|---------------|--|
| of Custody | |
| Chain | |

| Seriversa | | | | | Chain of Custody Documentation | stody i | Docum | chiano | = | | | | | |
|---|--|---|--------------------------------------|--|--------------------------------|----------|-----------------|------------|----------|----------|-------------------|-------------|---|---------------------------|
| Senversa Pty Ltd www.senversa.com.au ABN 89 132 231 380 | | | Laboratory: Address: Contact: Phone: | ALS NSW Sample Receipt | | | | | | Analysi | Analysis Required | | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. | minated present; trace |
| Job Number: | | S20102 | Purchase Order: | | | (\$7\ | | | | J | E - H | - | | ş |
| | | Wetherill Park WME | Quote No: | EN/103/21 | | | | | | 2 | | | | |
| Sampled By: | | Rowan Faint | Turn Around Time: | Standard 7 Days | S | 1 8/H/ | | | | () | r: + | | | . 7 |
| Project Manager: | <u> </u> | Emma Walsh | Page: | 1 of | | _ | | | | IM O | | | | *** |
| | rowan.fain Bec.Chappli | rowan.faint@senversa.com.au Bec.Chapple@senversa.com.au; | Phone/Mobile: | 0408038593, 0404011544 | | (3T8/HЯ) | | \ | NT | нA ЭЭ) Э | 1 | 1 | | · . |
| | Sample Information | ation | | Container Information | | | 314 | 8 | pue | 1900 | | רם | | |
| Lab ID Sa | Sample ID Matrix * | Date | Time | Type / Code | al Bottles | | | -TN | - GI | ECC | | ЮН | | |
| | MW4 W | 14/02/2024 | | P, VS x2, N, UA, VSA | 9 | × | | × | | × | | | | |
| | | | | • | | 44 | • | | | | | | 7 | |
| | 11-11 | | | | | | | | | | | - | | |
| | | | | | | 30 | | | | | | | | |
| | | | | | | | | | | | , | - | | |
| | × | | | | | | | | | | EN | ironme | Environmental Division | 2 |
| | 14 101 | | | | | | | | | | Syc | Iney | | |
| | | | | | , jay- | | | | | | > - | Vork Ord | er Reference | |
| | 9 2 2 | | | | | | | | | | | スクロ | 404/22 | |
| - 1 84 9 1 | | | | | The second second | | | | | | _ | | | |
| | | | | | | | | | | | | 3 | | |
| | | | | And the state of t | 1808 | | | | | | | | | |
| | | | | | | | | | | | | E | | |
| 100 | | | | | | | | | | | | <u> </u> | | |
| | | | | | 2 | | | | | | Telept | hone: + 61. | Telephone: + 61-2-8784 8555 | |
| | | | | | 10 m | | | | | | _ | | | |
| | | | | | | | | | | | _ | _ | | |
| Total | 4. 11 | | | | 9 | | | | | | | | | |
| Sampler: Tattest that pospecifications were use | Sampler: Tattest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: | in accordance with Samples: | enversa standard proced | | Sampler Name: | Rowa | Rowan Faint | Signature: | ure: | 2Fall | nt | Date: | | 14/02/2024 |
| Relinquished By: | | | | Method of Shipment (if applicable): | :able): | | Received by | ıy: | | | | | | |
| Name/Signatūre: | Rowan Faint | | Date: 14/2/24 | Carrier / Reference #: | | | Name/Signature: | | 505 10th | 3 | | | Date: 14/2/24 | |
| Of | 700 | | | Date/Time: | | | Of: | AUS. | 2 | | | | 15 | |
| Name/Signature: | | | Ų | Carrier / Reference #: | | | Name/Signature: | ature: | | | | | Date: | |
| Of. | | | | Date/Time: | | | Of: | | | | | | Time: | |
| Name/Signature: | | | Date: | Carrier / Reference #: | | | Name/Signature: | ature: | | | | | Date: | |
| 101 | | | | Data/Time. | | | 75 | | | | | | | • |

Completed by:

Water Container Codes: P = Unpreserved Plastic; N = Nith Carid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOHyCadmium Hydroxide Preserved; S = Sodium Hydroxide Preserved Plastic; SH = Suphuric Preserved Plastic; VS = VOA Vial Sulphuric Preserved Plastic; NS = Suphuric Preserved Amber Glass; H = HCI Preserved Plastic; HS = HCI Preserved Speciation Bottle; SP = Suphuric Preserved Bottle; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved while plastic bottle; SW = sulfurcadd preserved wide mouth glass jar

Date/Time:



SAMPLE RECEIPT NOTIFICATION (SRN)

: ES2404752 Work Order

: SENVERSA PTY LTD Client Laboratory : Environmental Division Sydney

Contact : Rowan Faint Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

2000 SYDNEY NSW 2000

E-mail E-mail : rowan.faint@senversa.com.au : sandy.phan@alsglobal.com

Telephone Telephone : +61-2-8784 8555 Facsimile Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page · 1 of 2

Order number Quote number : EB2023SENVER0001 (EN/000) C-O-C number QC Level : NEPM 2013 B3 & ALS QC Standard

Site Sampler

: Rowan Faint

Dates

Date Samples Received : 14-Feb-2024 15:46 Issue Date : 14-Feb-2024 Scheduled Reporting Date : 20-Feb-2024 Client Requested Due 20-Feb-2024

Date

Delivery Details

Mode of Delivery Security Seal : Client Drop Off : Not Available

No of coolers/hoxes : 1 Temperature : 14.0, 12.1, 11.0'C - Ice

present

Receipt Detail No. of samples received / analysed : 1/1

General Comments

This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 14-Feb-2024 Issue Date

Page

2 of 2 ES2404752 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

BEC CHAPPLE

| - *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
|--|-------|---------------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| Rowan Faint | | |
| *AU Certificate of Analysis - NATA (COA) | Email | rowan.faint@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | rowan.faint@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | rowan.faint@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | rowan.faint@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | rowan.faint@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | rowan.faint@senversa.com.au |
| SUPPLIER ACCOUNTS | | |
| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
| | | u |



CERTIFICATE OF ANALYSIS

Work Order : E\$2404752

Client : SENVERSA PTY LTD

Contact : Rowan Faint

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : ---

Project : S20102 Wetherill Park WME

Order number : ----

C-O-C number : ----

Sampler : Rowan Faint

Site : ----

Quote number : EN/000

No. of samples received : 1
No. of samples analysed : 1

Page : 1 of 6

Laboratory : Environmental Division Sydney

Contact : Sandy Phan

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61-2-8784 8555

Date Samples Received : 14-Feb-2024 15:46

Date Analysis Commenced : 14-Feb-2024

Issue Date : 20-Feb-2024 12:37



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

SignatoriesPositionAccreditation CategoryAnkit JoshiSenior Chemist - InorganicsSydney Inorganics, Smithfield, NSWEdwandy FadjarOrganic CoordinatorSydney Organics, Smithfield, NSW

Page : 2 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- EP080: The result for sample ES2404752-001 was confirmed by re-analysis.

Page : 3 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW4 | | | |
|--------------------------------------|----------------------|--------|----------------|-------------------|------|-------------|--|
| (Wattix, WATER) | | Sampli | ng date / time | 14-Feb-2024 00:00 | | | |
| Compound | CAS Number | LOR | Unit | ES2404752-001 | | | |
| | | | | Result | | | |
| EG005(ED093)F: Dissolved Metals b | y ICP-AES | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 2.04 | | | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 5.03 | | | |
| EG020F: Dissolved Metals by ICP-N | IS | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.008 | | | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | | | |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | | | |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.001 | | | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | | | |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.017 | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.006 | | | |
| EG035F: Dissolved Mercury by FIM | S | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | | | |
| EK055G: Ammonia as N by Discrete | Analyser | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.30 | | | |
| EK057G: Nitrite as N by Discrete A | nalyser | | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | | | |
| EK058G: Nitrate as N by Discrete A | nalyser | | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | <0.01 | | | |
| EK059G: Nitrite plus Nitrate as N (N | IOx) by Discrete Ana | lyser | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | | | |
| EK061G: Total Kjeldahl Nitrogen By | Discrete Analyser | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.2 | | | |
| EK062G: Total Nitrogen as N (TKN - | NOx) by Discrete An | alyser | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 1.2 | | | |
| EK067G: Total Phosphorus as P by | Discrete Analyser | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.07 | | | |
| EP075(SIM)B: Polynuclear Aromatic | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | | | |

Page : 4 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW4 | | |
|---------------------------------------|---------------------|-----------|----------------|-------------------|------|------|
| (manus 1971 Ery | | Sampli | ng date / time | 14-Feb-2024 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2404752-001 | | |
| | | | | Result | | |
| EP075(SIM)B: Polynuclear Aromatic F | lydrocarbons - Cont | inued | | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | | |
| Sum of polycyclic aromatic hydrocarbo | ns | 0.5 | μg/L | <0.5 | | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | | |
| EP080/071: Total Petroleum Hydrocar | bons | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | | |
| C15 - C28 Fraction | | 100 | μg/L | <100 | | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | <50 | | |
| EP080/071: Total Recoverable Hydrod | arbons - NEPM 201 | 3 Fractio | ns | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | | |
| ^ C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | | |

Page : 5 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | Sample ID | MW4 | | |
|---|-------------------|--------|----------------|-------------------|------|------|
| (Matrix: WATER) | | | Gampio 12 | IVIVV | | |
| | | Sampli | ng date / time | 14-Feb-2024 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2404752-001 | | |
| | | | | Result | | |
| EP080/071: Total Recoverable Hydroca | arbons - NEPM 201 | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | | |
| >C16 - C34 Fraction | | 100 | μg/L | <100 | | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | <100 | | |
| ^ >C10 - C16 Fraction minus Naphthalene | | 100 | μg/L | <100 | | |
| (F2) | | | | | | |
| EP080: BTEXN | | | ä | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | | |
| Toluene | 108-88-3 | 2 | μg/L | 2 | | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | | |
| ^ Total Xylenes | | 2 | μg/L | <2 | | |
| ^ Sum of BTEX | | 1 | μg/L | 2 | | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | | |
| EP075(SIM)S: Phenolic Compound Sui | rrogates | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 24.0 | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 51.6 | | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 43.7 | | |
| EP075(SIM)T: PAH Surrogates | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 67.3 | | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 72.2 | | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 71.5 | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 88.3 | | |
| Toluene-D8 | 2037-26-5 | 2 | % | 97.6 | | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 106 | | |

Page : 6 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery | Limits (%) |
|---|------------|----------|------------|
| Compound | CAS Number | Low | High |
| EP075(SIM)S: Phenolic Compound Surrogates | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 |
| EP075(SIM)T: PAH Surrogates | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 |
| Anthracene-d10 | 1719-06-8 | 27 | 113 |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 |
| EP080S: TPH(V)/BTEX Surrogates | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 |
| Toluene-D8 | 2037-26-5 | 75 | 131 |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 |



QA/QC Compliance Assessment to assist with Quality Review

:ES2404752 **Work Order** Page : 1 of 7

: Environmental Division Sydney Client : SENVERSA PTY LTD Laboratory

: Rowan Faint Telephone : +61-2-8784 8555 Contact **Project** : S20102 Wetherill Park WME **Date Samples Received** : 14-Feb-2024 **Issue Date** Site : 20-Feb-2024

Sampler : Rowan Faint No. of samples received : 1

Order number No. of samples analysed : 1

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- Matrix Spike outliers exist please see following pages for full details.
- For all regular sample matrices, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

• NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 7 Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Outliers: Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: WATER

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|--|----------------------|------------------|-----------|------------|------------|--------|----------------------------------|
| Matrix Spike (MS) Recoveries | | | | | | | |
| EG005(ED093)F: Dissolved Metals by ICP-AES | ES2404752001 | MW4 | Manganese | 7439-96-5 | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |

Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | | Co | unt | Rate | e (%) | Quality Control Specification |
|--------------------------------------|------------|----|---------|--------|----------|--------------------------------|
| Analytical Methods | Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 0 | 1 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 3 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 3 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 0 | 9 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 3 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 3 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: **x** = Holding time breach ; ✓ = Within holding time.

| Method | Sample Date | Ex | traction / Preparation | | | Analysis | |
|---|-------------|----------------|------------------------|------------|---------------|------------------|------------|
| Container / Client Sample ID(s) | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG005(ED093)F: Dissolved Metals by ICP-AES | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG005F) MW4 | 14-Feb-2024 | | | | 16-Feb-2024 | 12-Aug-2024 | √ |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) MW4 | 14-Feb-2024 | | | | 16-Feb-2024 | 12-Aug-2024 | ✓ |
| EG035F: Dissolved Mercury by FIMS | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) MW4 | 14-Feb-2024 | | | | 19-Feb-2024 | 13-Mar-2024 | √ |

Page : 3 of 7
Work Order : ES2404752

Amber VOC Vial - Sulfuric Acid (EP080)

MW4

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: **x** = Holding time breach; ✓ = Within holding time. Method Sample Date Extraction / Preparation Analysis Container / Client Sample ID(s) Due for extraction Evaluation Due for analysis Evaluation Date extracted Date analysed EK055G: Ammonia as N by Discrete Analyser Clear Plastic Bottle - Sulfuric Acid (EK055G) 14-Feb-2024 13-Mar-2024 MW4 19-Feb-2024 EK057G: Nitrite as N by Discrete Analyser Clear Plastic Bottle - Natural (EK057G) MW4 14-Feb-2024 14-Feb-2024 16-Feb-2024 EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analyser Clear Plastic Bottle - Sulfuric Acid (EK059G) MW4 14-Feb-2024 19-Feb-2024 13-Mar-2024 EK061G: Total Kjeldahl Nitrogen By Discrete Analyser Clear Plastic Bottle - Sulfuric Acid (EK061G) 14-Feb-2024 13-Mar-2024 16-Feb-2024 13-Mar-2024 MW4 16-Feb-2024 EK067G: Total Phosphorus as P by Discrete Analyser Clear Plastic Bottle - Sulfuric Acid (EK067G) 14-Feb-2024 16-Feb-2024 13-Mar-2024 16-Feb-2024 13-Mar-2024 MW4 EP075(SIM)B: Polynuclear Aromatic Hydrocarbons Amber Glass Bottle - Unpreserved (EP075(SIM)) 14-Feb-2024 15-Feb-2024 21-Feb-2024 17-Feb-2024 26-Mar-2024 MW4 EP080/071: Total Petroleum Hydrocarbons Amber Glass Bottle - Unpreserved (EP071) 21-Feb-2024 14-Feb-2024 15-Feb-2024 17-Feb-2024 26-Mar-2024 Amber VOC Vial - Sulfuric Acid (EP080) 15-Feb-2024 14-Feb-2024 15-Feb-2024 28-Feb-2024 28-Feb-2024 EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions Amber Glass Bottle - Unpreserved (EP071) 14-Feb-2024 15-Feb-2024 21-Feb-2024 17-Feb-2024 26-Mar-2024 MW4 Amber VOC Vial - Sulfuric Acid (EP080) 28-Feb-2024 28-Feb-2024 MW4 14-Feb-2024 15-Feb-2024 15-Feb-2024 EP080: BTEXN

14-Feb-2024

15-Feb-2024

28-Feb-2024

15-Feb-2024

28-Feb-2024

Page : 4 of 7
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

the expected rate. A listing of breaches is provided in the Summary of Outliers.

| Matrix: WATER | | | | Evaluatio | n: × = Quality Co | ntrol frequency r | not within specification ; \checkmark = Quality Control frequency within specification. |
|---|------------|----|---------|-----------|-------------------|-------------------|---|
| Quality Control Sample Type | | Co | unt | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 10 | 20.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 0 | 1 | 0.00 | 10.00 | 3c | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 1 | 100.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 14 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 8 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 3 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 14 | 14.29 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 16 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 3 | 0.00 | 10.00 | 3¢ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 1 | 100.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 1 | 100.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 3 | 33.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 14 | 21.43 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 3 | 16 | 18.75 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 3 | 33.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 1 | 100.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 1 | 100.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 9 | 11.11 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 8 | 12.50 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 3 | 33.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 14 | 7.14 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 3 | 33.33 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |

Page : 5 of 7
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: * = Quality Control frequency not within specification; * = Quality Control frequency within specification. Quality Control Sample Type Count Rate (%) Quality Control Specification Analytical Methods Method QC Evaluation Regular Actual Expected Matrix Spikes (MS) - Continued Ammonia as N by Discrete analyser 10 10.00 5.00 NEPM 2013 B3 & ALS QC Standard EK055G 1 1 Dissolved Mercury by FIMS 1 NEPM 2013 B3 & ALS QC Standard 1 100.00 5.00 EG035F 1 Dissolved Metals by ICP-AES 1 100.00 5.00 NEPM 2013 B3 & ALS QC Standard EG005F ✓ Dissolved Metals by ICP-MS - Suite A 0 9 0.00 NEPM 2013 B3 & ALS QC Standard EG020A-F 5.00 × Nitrite and Nitrate as N (NOx) by Discrete Analyser EK059G 1 14 7.14 5.00 ✓ NEPM 2013 B3 & ALS QC Standard Nitrite as N by Discrete Analyser 1 8 NEPM 2013 B3 & ALS QC Standard 12.50 5.00 EK057G ✓ PAH/Phenols (GC/MS - SIM) 0 3 0.00 5.00 NEPM 2013 B3 & ALS QC Standard EP075(SIM) × Total Kjeldahl Nitrogen as N By Discrete Analyser 1 14 7.14 5.00 NEPM 2013 B3 & ALS QC Standard EK061G 1 Total Phosphorus as P By Discrete Analyser 16 1 6.25 5.00 1 NEPM 2013 B3 & ALS QC Standard EK067G TRH - Semivolatile Fraction 0 3 0.00 NEPM 2013 B3 & ALS QC Standard EP071 5.00 × TRH Volatiles/BTEX EP080 1 20 5.00 5.00 NEPM 2013 B3 & ALS QC Standard

Page : 6 of 7
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|--|---|--|--|
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered |
| | | | samples, emitting a characteristic spectrum which is compared against matrix matched standards. This |
| | | | method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered |
| | | | prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions |
| | | | are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct |
| | | | mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are |
| | | | 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A |
| | | | bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic |
| | | | mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. |
| | | | Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM |
| | | | Schedule B(3). |
| Ammonia as N by Discrete analyser | EK055G | WATER | In house: Referenced to APHA 4500-NH3 G Ammonia is determined by direct colorimetry by Discrete Analyser. |
| | | | This method is compliant with NEPM Schedule B(3) |
| Nitrite as N by Discrete Analyser | EK057G | WATER | In house: Referenced to APHA 4500-NO2- B. Nitrite is determined by direct colourimetry by Discrete Analyser. |
| | | | This method is compliant with NEPM Schedule B(3) |
| Nitrate as N by Discrete Analyser | EK058G | WATER | In house: Referenced to APHA 4500-NO3- F. Nitrate is reduced to nitrite by way of a chemical reduction followed |
| | | | by quantification by Discrete Analyser. Nitrite is determined seperately by direct colourimetry and result for Nitrate |
| | | \\\\\ TEB | calculated as the difference between the two results. This method is compliant with NEPM Schedule B(3) |
| Nitrite and Nitrate as N (NOx) by Discrete | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by |
| Analyser | | | Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM |
| | | \\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\ | Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high |
| Analyser | | | temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined |
| | =:::::::::::::::::::::::::::::::::::::: | MATER | colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Discrete Analyser | =:: | VALATED | |
| Total Phosphorus as P By Discrete | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid |
| Analyser | | | digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with |
| | | | ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its |
| TRH - Semivolatile Fraction | ED074 | \\\\\ | concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRD - Semiyolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and |
| | | | quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This |
| | | | method is compliant with the QC requirements of NEPM Schedule B(3) |

Page : 7 of 7
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Analytical Methods | Method | Matrix | Method Descriptions |
|--------------------------------------|-----------------------|------------------------|--|
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode |
| | | | and quantification is by comparison against an established 5 point calibration curve. This method is compliant |
| | | | with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary |
| | | | GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a |
| | | | sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This |
| | | | method is compliant with the QC requirements of NEPM Schedule B(3) |
| | | | |
| Preparation Methods | Method | Matrix | Method Descriptions |
| Preparation Methods TKN/TP Digestion | Method EK061/EK067 | <i>Matrix</i> WATER | Method Descriptions In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| | | | |
| | | | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated |



Address

QUALITY CONTROL REPORT

Address

Work Order : **ES2404752** Page : 1 of 6

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : Rowan Faint : Sandy Phan

SYDNEY NSW 2000

Telephone : ---- Telephone : +61-2-8784 8555

Project : \$20102 Wetherill Park WME Date Samples Received : 14-Feb-2024

Order number : ---- Date Analysis Commenced : 14-Feb-2024

C-O-C number : ---- Issue Date

: Level 24, 1 Market St, Sydney NSW 2000

Sampler : Rowan Faint

Site : ---Quote number : EN/000

No. of samples received : 1

No. of samples analysed : 1

Accreditation No. 825
Accredited for compliance with ISO/IEC 17025 - Testing

: 277-289 Woodpark Road Smithfield NSW Australia 2164

· 20-Feb-2024

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

SignatoriesPositionAccreditation CategoryAnkit JoshiSenior Chemist - InorganicsSydney Inorganics, Smithfield, NSWEdwandy FadjarOrganic CoordinatorSydney Organics, Smithfield, NSW

Page : 2 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit: Result between 10 and 20 times LOR: 0% - 50%: Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory L | Duplicate (DUP) Report | | |
|----------------------|------------------------|--|------------|------------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | solved Metals by ICP | -AES (QC Lot: 5604225) | | | | | | | |
| ES2404752-001 | MW4 | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 5.03 | 5.03 | 0.0 | 0% - 20% |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 2.04 | 2.06 | 0.6 | 0% - 20% |
| EG020F: Dissolved | Metals by ICP-MS (QC | C Lot: 5604222) | | | | | | | |
| ES2404563-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.025 | 0.024 | 0.0 | 0% - 20% |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.001 | 0.002 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| EK055G: Ammonia | as N by Discrete Analy | yser (QC Lot: 5607396) | | | | | | | |
| ME2400277-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.64 | 0.64 | 0.0 | 0% - 20% |
| ES2404468-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.01 | 0.01 | 0.0 | No Limit |
| EK057G: Nitrite as | N by Discrete Analyse | er (QC Lot: 5601798) | | | | | | | |
| ES2404552-003 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK059G: Nitrite plu | s Nitrate as N (NOx) t | by Discrete Analyser (QC Lot: 5607397) | | | | | | | |
| ES2404549-002 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.03 | 0.03 | 0.0 | No Limit |
| ES2404468-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK061G: Total Kjeld | lahl Nitrogen By Discr | rete Analyser (QC Lot: 5607394) | | | | | | | |
| ES2404424-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (2.0)* | mg/L | 30.2 | 29.2 | 3.5 | 0% - 50% |

Page : 3 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | | Laboratory I | Ouplicate (DUP) Report | | |
|----------------------|------------------------|---|------------|--------------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EK061G: Total Kjeld | lahl Nitrogen By Discr | ete Analyser (QC Lot: 5607394) - continued | | | | | | | |
| ES2404549-002 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | <0.1 | 0.0 | No Limit |
| EK067G: Total Phos | sphorus as P by Discre | ete Analyser (QC Lot: 5607395) | | | | | | | |
| ES2404424-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 (0.20)* | mg/L | 3.04 | 3.21 | 5.2 | 0% - 50% |
| ES2404549-002 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.02 | 0.02 | 0.0 | No Limit |
| EP080/071: Total Pe | troleum Hydrocarbons | s (QC Lot: 5601542) | | | | | | | |
| ES2404452-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2404516-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Re | coverable Hydrocarbo | ons - NEPM 2013 Fractions (QC Lot: 5601542) | | | | | | | |
| ES2404452-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2404516-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5601542) | | | | | | | | |
| ES2404452-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| ES2404516-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 4 of 6 Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|--|--------------------|---------|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 560 |)4225) | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 110 | 82.0 | 114 |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 106 | 81.0 | 113 |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5604222) | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 101 | 85.0 | 114 |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 100 | 84.0 | 110 |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 99.9 | 85.0 | 111 |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 95.8 | 81.0 | 111 |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 101 | 83.0 | 111 |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 98.0 | 82.0 | 112 |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 99.8 | 81.0 | 117 |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5604224) | | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 97.8 | 83.0 | 105 |
| EK055G: Ammonia as N by Discrete Analyser (QCLot: 560 | 7396) | | | | | | | |
| EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 91.4 | 90.0 | 114 |
| EK057G: Nitrite as N by Discrete Analyser (QCLot: 560179 | 8) | | | | | | | |
| EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 95.4 | 82.0 | 114 |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analys | ser (QCLot: 56 | (07397) | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 106 | 91.0 | 113 |
| EK061G: Total Kieldahl Nitrogen By Discrete Analyser (QC | Lot: 5607394) | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 100 | 69.0 | 101 |
| | | | | <0.1 | 1 mg/L | 114 | 70.0 | 118 |
| | | | | <0.1 | 5 mg/L | 103 | 70.0 | 130 |
| EK067G: Total Phosphorus as P by Discrete Analyser (QC | Lot: 5607395) | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 94.0 | 71.3 | 126 |
| | | | | <0.01 | 0.442 mg/L | 99.4 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 104 | 70.0 | 130 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLo | : 56013 <u>97)</u> | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 72.2 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 76.1 | 63.6 | 114 |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 91.3 | 62.2 | 113 |
| <u> </u> | | | | 1 | | | | |

Page : 5 of 6
Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|------------------|------|-------------------|---------------|------------------------------|------------|------------|
| | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound CAS Num | ber LOR | Unit | Result | Concentration | LCS | Low | High |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLot: 5601397 | | | | | | | |
| EP075(SIM): Fluorene 86-73 | | μg/L | <1.0 | 5 μg/L | 74.4 | 63.9 | 115 |
| EP075(SIM): Phenanthrene 85-01 | | μg/L | <1.0 | 5 μg/L | 74.8 | 62.6 | 116 |
| EP075(SIM): Anthracene 120-12 | -7 1 | μg/L | <1.0 | 5 μg/L | 78.2 | 64.3 | 116 |
| EP075(SIM): Fluoranthene 206-44 | -0 1 | μg/L | <1.0 | 5 μg/L | 80.7 | 63.6 | 118 |
| EP075(SIM): Pyrene 129-00 | -0 1 | μg/L | <1.0 | 5 μg/L | 78.1 | 63.1 | 118 |
| EP075(SIM): Benz(a)anthracene 56-55 | -3 1 | μg/L | <1.0 | 5 μg/L | 77.2 | 64.1 | 117 |
| EP075(SIM): Chrysene 218-01 | -9 1 | μg/L | <1.0 | 5 μg/L | 72.6 | 62.5 | 116 |
| EP075(SIM): Benzo(b+j)fluoranthene 205-99 205-82 | | μg/L | <1.0 | 5 μg/L | 73.0 | 61.7 | 119 |
| EP075(SIM): Benzo(k)fluoranthene 207-08 | -9 1 | μg/L | <1.0 | 5 μg/L | 75.8 | 63.0 | 115 |
| EP075(SIM): Benzo(a)pyrene 50-32 | -8 0.5 | μg/L | <0.5 | 5 μg/L | 72.4 | 63.3 | 117 |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | -5 1 | μg/L | <1.0 | 5 μg/L | 80.3 | 59.9 | 118 |
| EP075(SIM): Dibenz(a.h)anthracene 53-70 | -3 1 | μg/L | <1.0 | 5 μg/L | 71.9 | 61.2 | 117 |
| EP075(SIM): Benzo(g.h.i)perylene 191-24 | -2 1 | μg/L | <1.0 | 5 μg/L | 75.9 | 59.1 | 118 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5601398) | | | | | | | |
| EP071: C10 - C14 Fraction - | 50 | μg/L | <50 | 400 μg/L | 79.5 | 53.7 | 97.0 |
| EP071: C15 - C28 Fraction - | 100 | μg/L | <100 | 600 μg/L | 84.9 | 63.3 | 107 |
| EP071: C29 - C36 Fraction - | 50 | μg/L | <50 | 400 μg/L | 88.8 | 58.3 | 120 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5601542) | | | | | | | |
| EP080: C6 - C9 Fraction | 20 | μg/L | <20 | 260 μg/L | 81.5 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions | (QCLot: 5601398) | | | | | | |
| EP071: >C10 - C16 Fraction - | 100 | μg/L | <100 | 500 μg/L | 66.2 | 53.9 | 95.5 |
| EP071: >C16 - C34 Fraction - | 100 | μg/L | <100 | 700 μg/L | 74.6 | 57.8 | 110 |
| EP071: >C34 - C40 Fraction - | 100 | μg/L | <100 | 300 μg/L | 73.7 | 50.5 | 115 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions | (QCLot: 5601542) | | | | | | |
| EP080: C6 - C10 Fraction C6_C | 10 20 | μg/L | <20 | 310 μg/L | 75.7 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 5601542) | | | | | | | |
| EP080: Benzene 71-43 | -2 1 | μg/L | <1 | 10 μg/L | 91.4 | 68.3 | 119 |
| EP080: Toluene 108-88 | -3 2 | μg/L | <2 | 10 μg/L | 99.4 | 73.5 | 120 |
| EP080: Ethylbenzene 100-41 | -4 2 | μg/L | <2 | 10 μg/L | 99.9 | 73.8 | 122 |
| EP080: meta- & para-Xylene 108-38 106-42 | | μg/L | <2 | 10 μg/L | 105 | 73.0 | 122 |
| EP080: ortho-Xylene 95-47 | -6 2 | μg/L | <2 | 10 μg/L | 102 | 76.4 | 123 |
| EP080: Naphthalene 91-20 | -3 5 | μg/L | <5 | 10 μg/L | 99.3 | 75.5 | 124 |

Page : 6 of 6 Work Order : ES2404752

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | | Ma | atrix Spike (MS) Report | | |
|----------------------|--|---------------------------------------|------------|---------------|-------------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable L | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG005(ED093)F: [| Dissolved Metals by ICP-AES (QCLot: 5604225) | | | | | | |
| ES2404752-001 | MW4 | EG005F: Manganese | 7439-96-5 | 1 mg/L | # Not Determined | 70.0 | 130 |
| EG035F: Dissolve | d Mercury by FIMS (QCLot: 5604224) | | | | Determined | | |
| ES2404752-001 | MW4 | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 93.8 | 70.0 | 130 |
| EK055G: Ammoni | a as N by Discrete Analyser (QCLot: 5607396) | | | | | | |
| ES2404468-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.5 mg/L | 85.7 | 70.0 | 130 |
| EK057G: Nitrite a | s N by Discrete Analyser (QCLot: 5601798) | | | | | | |
| ES2404552-003 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.5 mg/L | 92.2 | 70.0 | 130 |
| EK059G: Nitrite p | lus Nitrate as N (NOx) by Discrete Analyser (QCLot: 56 | | | | | | |
| ES2404468-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 99.8 | 70.0 | 130 |
| EK061G: Total Kje | eldahl Nitrogen By Discrete Analyser (QCLot: 5607394) | | | | | | |
| ES2404468-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 5 mg/L | 90.6 | 70.0 | 130 |
| EK067G: Total Ph | osphorus as P by Discrete Analyser (QCLot: 5607395) | | | | | | |
| ES2404468-001 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 90.5 | 70.0 | 130 |
| EP080/071: Total I | Petroleum Hydrocarbons (QCLot: 5601542) | | | | | | |
| ES2404452-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 81.8 | 70.0 | 130 |
| EP080/071: Total I | Recoverable Hydrocarbons - NEPM 2013 Fractions (QCI | .ot: 5601542) | | | | | |
| ES2404452-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 76.0 | 70.0 | 130 |
| EP080: BTEXN (C | (CLot: 5601542) | | | | | | |
| ES2404452-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 99.7 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 98.6 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 98.9 | 70.0 | 130 |
| | | , , , , , , , , , , , , , , , , , , , | 108-38-3 | 25 μg/L | 103 | 70.0 | 130 |
| | | | 106-42-3 | | | | 100 |
| | | a. ecc. craio Ayiene | 95-47-6 | 25 μg/L | 99.0 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 94.7 | 70.0 | 130 |

senversa

Chain of Custody Documentation

| Senversa Pty I | | | | Laboratory: | | | | | | | | Δ | nalysis R | equire | 4 | - | | | | |
|--------------------------------|---|-------------------|---|--------------------------------|--------------------------|---------------|---------------------|------------------------------------|--|--|--|------------------------------------|------------------|--------|-------|----------------|-------------------------------|---|-----------|--|
| www.senversa. ABN 89 132 23 | com.au | | ~ | Address: Contact: Phone: | ALS NSW Sample Receipt | | Level | | | | | | | | | | | Comments: e.g. Highly contaminate sample; hazardous materials prese LORs etc. | | |
| Job Number: | | S20 | 0102 | Purchase Order: | | | rd Le | Level | | Total Phosphorus | sta sa sa | | | | | | | | | |
| Project Name: | | Wetherill | Park WME | Quote No: | EN\103\ | 121 | Standard | ard L | Mn) | hosp | W 8/8 | | | | | | | | | |
| Sampled By: | • | Rowa | ın Faint | Turn Around Time: | Standa | rd | | Standard | and | otal P | enols | 7 | | | | | | | | |
| Project Manag | er: | Emma | a Walsh | Page: | 1 | of 1 | 15H Solids | 1 | S (Fe | nd To | H/H | TEX | 0 | | . | | | | | |
| Email Report 1 | o: | emma.walsh@ | senversa.com.au senversa.com.au senversa.com.au | Phone/Mobile: | 0420 218 | 0420 218 472 | | WATER - EA025H Suspended Solids | Suspended Solids WATER - EG005F Dissolved Metals (| Dissolved Metals (H WATER - NT-11 Total Nitrogen and T | WATER - W-27 TRH/BTEXN/PAH/Phenols/8 Metals | WATER - W-18 TRH(C6 - C9)/BTEXN | - EP080 | | | | | | | |
| | | Sample Informatio | | | Container Info | | TER - | TER | TER | HE IN | TER 1/BT | TER (C6 | WATER - BTEXN | | | | 40LD | | | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | WATER Total D | WA | WA | WA | X X | NAT TRI | WA | | | | 오 | | | |
| | SW1 | Water | 9/07/2024 | AM | | 6 | Х | X | X | X | X | | | | | | | | | |
| | SW2 | Water | 9/07/2024 | AM | | 6 | Х | X | X | X | X | | | | | | | | | |
| | QC405 | Water | 9/07/2024 | AM | | 1 | | | | | | X | | | | | and the state of the state of | | | |
| | QC505 | Water | 9/07/2024 | AM | | 1 | | | | | | | X | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | |
| | | 9 N | | | | Envir | onm | enta | l Divi | sion | | | | | | | | | | |
| | | | 1 | | | Sydn | ey | | | 0.011 | | | | | | | | | | |
| | | | | | | Siydn Wo | rk Or | der Re | eferend | ce | | | | | | | | | | |
| | | | FED | | | \bot E | S2 | :42 | 25 | 53 | | | | | | | | | | |
| | | | | | | | | | | - | | | | | | | | | - | |
| | | | | | | | HIL | | MATE I | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | |
| | | | | | | | | 144 | 100 | | | | | - | - | - | | | | |
| | | | | | | | N.V | | 113 | | | | | | | | 200 | | | |
| | | | | | | | 1 1 1 1 1 1 1 1 1 1 | (Martin | II., i je i | | | | | | | | | | | |
| | | | | | | Te ephon | e: +61 | 1-2-8784 | 3555 | | | | | | | | | | | |
| | | | | | | | L | e e | | | | | | | | | | | | |
| | | | | | | | ļ | ļ | | ļ | | | | | | | | | | |
| | | | | | | | | | | | | | | | 00202 | | | | | |
| Total | | | | | | 14 | | | | | | | | | | | | | | |
| | st that proper field sar were used during the | | | Senversa standard prod | cedures and/or project | Sampler Name: | | Rowa | an Faint | | Signat | ure: | Q: | au | X | | Date: | . 9 | 9/07/2024 | |
| Relinquished | Ву: | | | | Method of Shipment (if a | applicable): | | | Receiv | ed by: | | | | 1 | | | | - | | |
| Name/Signatur | e: | Rowan Faint | | Date: 9/7/2024 | | | | | | Signatur | e: 7 | TAD | 1 | /6 | | | | Date: | | |
| Of: | | Senversa | | Time: 12:30 PM | : 12:30 PM Date/Time: | | | | Of: | ~ | | | // | A | | | | Time: | | |
| | lame/Signature: Date: Carrier / Reference #: | | Carrier / Reference #: Date/Time: | | | | Name/s Of: | Signatur | e: | | -6 | MAR | 4 | 12:4 | 10/ | Date: Time: | | | | |
| Of: Name/Signatur | 0' | | | Time: | Carrier / Reference #: | | | | | Signatur | о. | | | | | | | Date: | | |
| Name/Signatur Of: | ь. | | | | Date/Time: | | | | Of: | Jignatur | ·. | | | | | | | Time: | | |
| V = | Time: Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO ₁) Preserved Plastic; ORC = V = VOA Vial Hydochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; | | | Preserved Amber Glass; H = He | CI Preserved Plastic; F | IS = HCI | Preserve | ed Speciat | ion Battle | e; SP = S | Julphuric F | reserve | l Plastic; | | | | served plastic; | | | |

Completed by: _____ Checked by: ____



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2422553

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : sandy.phan@alsqlobal.com

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555 Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 2

 Order number
 : --- Quote number
 : EB2023SENVER0001 (EN/000)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Sampler : Rowan Faint

Dates

Site

Date Samples Received : 09-Jul-2024 12:40 Issue Date : 09-Jul-2024 Client Requested Due : 16-Jul-2024 Scheduled Reporting Date : 16-Jul-2024

Date

Delivery Details

Mode of Delivery : Client Drop Off Security Seal : Intact.

No. of coolers/boxes : 1 Temperature : 3.6'C 4.4'C - Ice present

Receipt Detail : No. of samples received / analysed : 4 / 4

General Comments

• This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Sample(s) requiring volatile organic compound analysis received in airtight containers (ZHE).
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Unless otherwise stated, analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 09-Jul-2024 Issue Date

Page

2 of 2 ES2422553 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

Some items described below may be part of a laboratory process necessary for the execution of client requested tasks. Packages may contain additional analyses, such as the determination of moisture content and preparation tasks, that are included in the package. Fotal Dissolved Solids - Standard Level otal Nitrogen and Total Phosphorus If no sampling time is provided, the sampling time will TRH/BTEXN/PAH/Phenols/8 Metals Suspended Solids - Standard Level default 00:00 on the date of sampling. If no sampling date is provided, the sampling date will be assumed by the laboratory and displayed in brackets without a time WATER - W-18 IRH(C6 - C9)/BTEXN component VATER - EA015H VATER - EG020F VATER - EA025H VATER - NT-11 Matrix: WATER VATER - E Sampling date / Sample ID Laboratory sample time ES2422553-001 09-Jul-2024 00:00 ✓ ES2422553-002 09-Jul-2024 00:00 SW2 ES2422553-003 09-Jul-2024 00:00 QC405 ES2422553-004 09-Jul-2024 00:00 QC505

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

EMMA WALSH

| - *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
|--|-------|---------------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Tax Invoice (INV) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| SUPPLIER ACCOUNTS | | _ |
| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
| | | U |



CERTIFICATE OF ANALYSIS

Work Order : ES2422553

Client SENVERSA PTY LTD

Contact EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number

C-O-C number

Sampler : Rowan Faint

Site

Quote number : EN/000

No. of samples received : 4 No. of samples analysed : 4 Page : 1 of 7

Laboratory : Environmental Division Sydney

Contact : Sandy Phan

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61-2-8784 8555

Date Samples Received : 09-Jul-2024 12:40

Date Analysis Commenced : 10-Jul-2024

Issue Date : 16-Jul-2024 13:48



ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with **Quality Review and Sample Receipt Notification.**

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Sydney Organics, Smithfield, NSW Edwandy Fadjar Organic Coordinator

Page : 2 of 7

Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- Unless otherwise stated, analytical work for this work order will be conducted at ALS Sydney.
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.

Page : 3 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | Sample ID | SW1 | SW2 | QC405 | QC505 | |
|---|----------------------|-----------------|-------------------|-------------------|-------------------|-------------------|---------------|--|
| (Matrix: WATER) | Sampling date / time | | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | | |
| Compound | CAS Number | LOR | Unit | ES2422553-001 | ES2422553-002 | ES2422553-003 | ES2422553-004 | |
| Compound | CAS Number | LON | Onn | Result | Result | Result | Result | |
| EA015: Total Dissolved Solids dried at 18 | 80 + 5 °C | | | resuit | resuit | resuit | resuit | |
| Total Dissolved Solids @180°C | | 10 | mg/L | 228 | 282 | | | |
| EA025: Total Suspended Solids dried at | 104 + 2°C | | | | | | | |
| Suspended Solids (SS) | | 5 | mg/L | 7260 | 90 | | | |
| EG020F: Dissolved Metals by ICP-MS | 11 11 11 11 | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| Chromium | 7440-47-3 | 0.001 | mg/L | 0.001 | 0.001 | | | |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.003 | 0.004 | | | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Manganese | 7439-96-5 | 0.001 | mg/L | 0.045 | 0.015 | | | |
| Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.023 | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | | | |
| EG035F: Dissolved Mercury by FIMS | 1 11 1 | Till the second | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| EK059G: Nitrite plus Nitrate as N (NOx) | bv Discrete Ana | lvser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.32 | 0.51 | | | |
| EK061G: Total Kjeldahl Nitrogen By Disc | rete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 22.6 | 3.8 | | | |
| EK062G: Total Nitrogen as N (TKN + NOx | () by Discrete Ar | nalyser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 22.9 | 4.3 | | | |
| EK067G: Total Phosphorus as P by Disci | rete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 6.68 | 0.63 | | | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Phenol | 108-95-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Chlorophenol | 95-57-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2-Methylphenol | 95-48-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |

Page : 4 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | SW2 | QC405 | QC505 | |
|---------------------------------------|-------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2422553-001 | ES2422553-002 | ES2422553-003 | ES2422553-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| 3- & 4-Methylphenol | 1319-77-3 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| 2-Nitrophenol | 88-75-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dimethylphenol | 105-67-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4-Dichlorophenol | 120-83-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.6-Dichlorophenol | 87-65-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 4-Chloro-3-methylphenol | 59-50-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.6-Trichlorophenol | 88-06-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| 2.4.5-Trichlorophenol | 95-95-4 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pentachlorophenol | 87-86-5 | 2.0 | μg/L | <2.0 | <2.0 | | | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| ^ Sum of polycyclic aromatic hydrocar | bons | 0.5 | μg/L | <0.5 | <0.5 | | | |

Page : 5 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | SW2 | QC405 | QC505 | |
|--|---------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2422553-001 | ES2422553-002 | ES2422553-003 | ES2422553-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons - Cont | inued | | | | | | |
| ^ Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | | | |
| EP080/071: Total Petroleum Hydroca | ırbons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | | | |
| C15 - C28 Fraction | | 100 | μg/L | 290 | <100 | | | |
| C29 - C36 Fraction | | 50 | μg/L | 130 | <50 | | | |
| ^ C10 - C36 Fraction (sum) | | 50 | μg/L | 420 | <50 | | | |
| EP080/071: Total Recoverable Hydro | carbons - NEPM 201 | 3 Fraction | าร | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | | |
| C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | | | |
| >C16 - C34 Fraction | | 100 | μg/L | 380 | <100 | | | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | | | |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | 380 | <100 | | | |
| ^ >C10 - C16 Fraction minus Naphthaler (F2) | ne | 100 | μg/L | <100 | <100 | | | |
| EP080: BTEXN | 11 19 13 | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | 14 | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | 15 | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | 16 | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | 16 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | 14 | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | 30 | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | 75 | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | 21 | |
| EP075(SIM)S: Phenolic Compound S | Gurrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 27.9 | 19.6 | | | |

Page : 6 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | SW1 | SW2 | QC405 | QC505 | |
|-----------------------------------|----------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | 09-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2422553-001 | ES2422553-002 | ES2422553-003 | ES2422553-004 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)S: Phenolic Compound Su | rrogates - Continued | 1 | | | | | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 55.9 | 41.2 | | | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 65.8 | 41.5 | | | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 57.5 | 51.0 | | | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 59.7 | 58.5 | | | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 70.5 | 60.3 | | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 113 | 114 | 82.2 | 90.8 | |
| Toluene-D8 | 2037-26-5 | 2 | % | 115 | 114 | 90.9 | 103 | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 109 | 106 | 106 | 106 | |

Page : 7 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | Recovery Limits (%) | | |
|---|---------------------|-----|------|
| Compound | CAS Number | Low | High |
| EP075(SIM)S: Phenolic Compound Surrogates | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 |
| EP075(SIM)T: PAH Surrogates | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 |
| Anthracene-d10 | 1719-06-8 | 27 | 113 |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 |
| EP080S: TPH(V)/BTEX Surrogates | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 |
| Toluene-D8 | 2037-26-5 | 75 | 131 |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2422553** Page : 1 of 7

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact: EMMA WALSHTelephone: +61-2-8784 8555Project: \$20102 Wetherill Park WMEDate Samples Received: 09-Jul-2024

Site : --- Issue Date : 16-Jul-2024

Sampler : Rowan Faint No. of samples received : 4
Order number : ---- No. of samples analysed : 4

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- NO Matrix Spike outliers occur.
- For all regular sample matrices, where applicable to the methodology, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

NO Analysis Holding Time Outliers exist.

Outliers : Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 7 Work Order : ES2422553

 Client
 : SENVERSA PTY LTD

 Project
 · S20102 Wetherill Park WME



Outliers: Frequency of Quality Control Samples

Matrix: WATER

| Quality Control Sample Type | | Count | | Rate (%) | | Quality Control Specification |
|-----------------------------|------------|-------|---------|----------|----------|--------------------------------|
| Analytical Methods | Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 5 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 5 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive <u>or</u> Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: **x** = Holding time breach ; ✓ = Within holding time.

| Method | | Sample Date | Ex | Extraction / Preparation | | | Analysis | | |
|---|-------|-------------|----------------|--------------------------|------------|---------------|------------------|------------|--|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C | | | | | | | | | |
| Clear Plastic Bottle - Natural (EA015H) SW1, | SW2 | 09-Jul-2024 | | | | 11-Jul-2024 | 16-Jul-2024 | ✓ | |
| EA025: Total Suspended Solids dried at 104 ± 2°C | | | | | | | | | |
| Clear Plastic Bottle - Natural (EA025H) SW1, | SW2 | 09-Jul-2024 | | | | 11-Jul-2024 | 16-Jul-2024 | ✓ | |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) SW1, | SW2 | 09-Jul-2024 | | | | 10-Jul-2024 | 05-Jan-2025 | ✓ | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) SW1, | SW2 | 09-Jul-2024 | | | | 12-Jul-2024 | 06-Aug-2024 | ✓ | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Ana | lyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) SW1, | SW2 | 09-Jul-2024 | | | | 11-Jul-2024 | 06-Aug-2024 | ✓ | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) SW1, | SW2 | 09-Jul-2024 | 11-Jul-2024 | 06-Aug-2024 | 1 | 11-Jul-2024 | 06-Aug-2024 | ✓ | |
| EK067G: Total Phosphorus as P by Discrete Analyser | | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK067G) SW1, | SW2 | 09-Jul-2024 | 11-Jul-2024 | 06-Aug-2024 | ✓ | 11-Jul-2024 | 06-Aug-2024 | ✓ | |

Page : 3 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Matrix: WATER | | | | | Lvaluation | . × = Holding time | breach; ▼ = withi | ir noluling time. |
|--|----------------|-------------|----------------|------------------------|------------|--------------------|-------------------|-------------------|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP075(SIM)A: Phenolic Compounds | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) SW1, | SW2 | 09-Jul-2024 | 10-Jul-2024 | 16-Jul-2024 | ✓ | 11-Jul-2024 | 19-Aug-2024 | √ |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) SW1, | SW2 | 09-Jul-2024 | 10-Jul-2024 | 16-Jul-2024 | ✓ | 11-Jul-2024 | 19-Aug-2024 | √ |
| EP080/071: Total Petroleum Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) SW1, | SW2 | 09-Jul-2024 | 10-Jul-2024 | 16-Jul-2024 | ✓ | 11-Jul-2024 | 19-Aug-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) QC405 | | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | 1 | 11-Jul-2024 | 23-Jul-2024 | √ |
| Amber VOC Vial - Sulfuric Acid (EP080) SW1, | SW2 | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | 1 | 12-Jul-2024 | 23-Jul-2024 | √ |
| EP080/071: Total Recoverable Hydrocarbons - NEPM | 2013 Fractions | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) SW1, | SW2 | 09-Jul-2024 | 10-Jul-2024 | 16-Jul-2024 | 1 | 11-Jul-2024 | 19-Aug-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) QC405 | | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | 1 | 11-Jul-2024 | 23-Jul-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) SW1, | SW2 | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | 1 | 12-Jul-2024 | 23-Jul-2024 | ✓ |
| EP080: BTEXN | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) QC405, | QC505 | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | 1 | 11-Jul-2024 | 23-Jul-2024 | √ |
| Amber VOC Vial - Sulfuric Acid (EP080) SW1, | SW2 | 09-Jul-2024 | 11-Jul-2024 | 23-Jul-2024 | ✓ | 12-Jul-2024 | 23-Jul-2024 | ✓ |

Page : 4 of 7 Work Order ES2422553

Client SENVERSA PTY LTD S20102 Wetherill Park WME Project



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to

| the expected rate. A listing of breaches is provided in the Summary | y of Outliers. | | | | | | |
|---|----------------|----|---------|-----------|-------------------|------------------|--|
| Matrix: WATER | | | | Evaluatio | n: × = Quality Co | ontrol frequency | not within specification; ✓ = Quality Control frequency within specification |
| Quality Control Sample Type | | Co | ount | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Regular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 5 | 20.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 5 | 20.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 7 | 28.57 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 4 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 4 | 39 | 10.26 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 7 | 28.57 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 11 | 18.18 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 5 | 0.00 | 10.00 | 3c | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 4 | 25.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 5 | 40 | 12.50 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 5 | 39 | 12.82 | 12.50 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 3 | 7 | 42.86 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 3 | 11 | 27.27 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 40 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 4 | 25.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Suspended Solids (High Level) | EA025H | 2 | 40 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Dissolved Solids (High Level) | EA015H | 2 | 39 | 5.13 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 11 | 9.09 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 40 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |
| Dissolved Mercury by FIMS | EG035F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 5 | 20.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |

Page : 5 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: **x** = Quality Control frequency not within specification; ✓ = Quality Control frequency within specification. Quality Control Sample Type Count Rate (%) Quality Control Specification Analytical Methods Method QC Reaular Expected Evaluation Actual Matrix Spikes (MS) - Continued PAH/Phenols (GC/MS - SIM) EP075(SIM) 0 4 0.00 5.00 NEPM 2013 B3 & ALS QC Standard Total Kjeldahl Nitrogen as N By Discrete Analyser 1 7 14.29 NEPM 2013 B3 & ALS QC Standard EK061G 5.00 1 Total Phosphorus as P By Discrete Analyser 11 EK067G 1 9.09 5.00 ✓ NEPM 2013 B3 & ALS QC Standard TRH - Semivolatile Fraction 0 5 NEPM 2013 B3 & ALS QC Standard 0.00 5.00 EP071 x TRH Volatiles/BTEX 2 40 NEPM 2013 B3 & ALS QC Standard EP080 5.00 5.00 ✓

Page : 6 of 7
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|---|------------|--------|---|
| Total Dissolved Solids (High Level) | EA015H | WATER | In house: Referenced to APHA 2540C. A gravimetric procedure that determines the amount of 'filterable' residue in an aqueous sample. A well-mixed sample is filtered through a glass fibre filter (1.2um). The filtrate is evaporated to dryness and dried to constant weight at 180+/-5C. This method is compliant with NEPM Schedule B(3) |
| Suspended Solids (High Level) | EA025H | WATER | In house: Referenced to APHA 2540D. A gravimetric procedure employed to determine the amount of `non-filterable` residue in a aqueous sample. The prescribed GFC (1.2um) filter is rinsed with deionised water, oven dried and weighed prior to analysis. A well-mixed sample is filtered through a glass fibre filter (1.2um). The residue on the filter paper is dried at 104+/-2C. This method is compliant with NEPM Schedule B(3) |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3) |

Page : 7 of 7 Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Analytical Methods | Method | Matrix | Method Descriptions |
|---|-------------|--------|--|
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This method is compliant with the QC requirements of NEPM Schedule B(3) |
| Preparation Methods | Method | Matrix | Method Descriptions |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| Separatory Funnel Extraction of Liquids | ORG14 | WATER | In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated and concentrated for analysis. This method is compliant with NEPM Schedule B(3). ALS default excludes sediment which may be resident in the container. |
| Volatiles Water Preparation | ORG16-W | WATER | A 5 mL aliquot or 5 mL of a diluted sample is added to a 40 mL VOC vial for purging. |



QUALITY CONTROL REPORT

Work Order : **ES2422553** Page : 1 of 9

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555

Project : \$20102 Wetherill Park WME Date Samples Received : 09-Jul-2024
Order number : ---- Date Analysis Commenced : 10-Jul-2024

C-O-C number : ---- Issue Date : 16-Jul-2024

Sampler : Rowan Faint

Site : ---

No. of samples analysed : 4

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall

Accreditation No. 825

Accredited for compliance with

not be reproduced, except in full.

This Quality Control Report contains the following information:

: EN/000

: 4

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

Quote number

No. of samples received

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW

Edwandy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 9 Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key: Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit: Result between 10 and 20 times LOR: 0% - 50%: Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | Laboratory Duplicate (DUP) Report | | | | | | |
|----------------------|--------------------------------|---------------------------------------|------------|-----------------------------------|------|-----------------|------------------|---------|--------------------|--|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) | |
| EA015: Total Dissolv | ved Solids dried at 180 ± 5 °C | C (QC Lot: 5918870) | | | | | | | | |
| EN2406536-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 444 | 454 | 2.3 | 0% - 20% | |
| EN2406634-001 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 810 | 810 | 0.0 | 0% - 20% | |
| ES2422089-005 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 32200 | 32300 | 0.2 | 0% - 20% | |
| ES2422511-004 | Anonymous | EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | 16 | 32 | 68.8 | No Limit | |
| EA025: Total Susper | nded Solids dried at 104 ± 2° | C (QC Lot: 5918869) | | | | | | | | |
| EN2406536-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 8 | 8 | 0.0 | No Limit | |
| EN2406634-001 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 11 | 12 | 13.0 | No Limit | |
| ES2422089-005 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | <5 | 0.0 | No Limit | |
| ES2422511-004 | Anonymous | EA025H: Suspended Solids (SS) | | 5 | mg/L | 18 | 25 | 30.2 | No Limit | |
| EG020F: Dissolved I | Metals by ICP-MS (QC Lot: 5 | 914847) | | | | | | | | |
| ES2422557-001 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit | |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.005 | 0.005 | 0.0 | No Limit | |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit | |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 0.001 | 0.001 | 0.0 | No Limit | |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | 0.001 | <0.001 | 0.0 | No Limit | |
| | | EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | 0.048 | 0.048 | 0.0 | 0% - 20% | |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.004 | 0.004 | 0.0 | No Limit | |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 0.062 | 0.062 | 0.0 | 0% - 50% | |
| | | EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | 0.0 | No Limit | |
| EG035F: Dissolved I | Mercury by FIMS (QC Lot: 5 | 914848) | | | | | | | | |

Page : 3 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | | Laboratory L | Ouplicate (DUP) Report | | |
|----------------------|-----------------------------|---------------------------------------|----------------------|--------------|--------------|-----------------|------------------------|---------|----------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG035F: Dissolved N | Mercury by FIMS (QC Lot: 8 | 5914848) - continued | | | | | | | |
| ES2422557-002 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| EK059G: Nitrite plus | Nitrate as N (NOx) by Dis | crete Analyser (QC Lot: 5918576) | | | | | | | |
| ES2422915-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.55 | 0.62 | 11.7 | 0% - 20% |
| ES2422660-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK061G: Total Kjelda | ahl Nitrogen By Discrete Ar | nalyser (QC Lot: 5918574) | | | | | | | |
| ES2422553-001 | SW1 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (2.0)* | mg/L | 22.6 | 25.4 | 11.9 | 0% - 50% |
| ES2422915-002 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (0.5)* | mg/L | 19.6 | 19.9 | 1.3 | 0% - 20% |
| EK067G: Total Phosp | phorus as P by Discrete An | alyser (QC Lot: 5918575) | | | | | | | |
| ES2422553-001 | SW1 | EK067G: Total Phosphorus as P | | 0.01 (0.20)* | mg/L | 6.68 | 6.55 | 2.0 | 0% - 20% |
| ES2422915-002 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 (0.05)* | mg/L | 0.17 | 0.18 | 7.5 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbons (QC | Lot: 5915799) | | | | | | | |
| EN2406506-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2422255-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Pet | roleum Hydrocarbons (QC | Lot: 5915806) | | | | | | | |
| ES2422453-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2422604-001 | Anonymous | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Rec | coverable Hydrocarbons - N | NEPM 2013 Fractions (QC Lot: 5915799) | | | | | | | |
| EN2406506-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2422255-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Rec | coverable Hydrocarbons - N | NEPM 2013 Fractions (QC Lot: 5915806) | | | | | | | |
| ES2422453-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2422604-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC I | Lot: 5915799) | | | | | | | | |
| EN2406506-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | 0.0 | N. 1 |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| F02422255 004 | Ananymaya | EP080: Naphthalene | 91-20-3 71-43-2 | 5 1 | μg/L | <5 <1 | <5 <1 | 0.0 | No Limit |
| ES2422255-001 | Anonymous | EP080: Benzene | | 2 | μg/L | | | | No Limit |
| | | EP080: Toluene | 108-88-3 100-41-4 | 2 | μg/L | <2 <2 | <2 <2 | 0.0 | No Limit No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L μg/L | <2 | <2 | 0.0 | No Limit No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | - | µg/L | ~~ | -2 | 0.0 | INO LITTIL |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 4 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | | Laboratory L | Ouplicate (DUP) Report | | |
|----------------------|---------------|----------------------------|------------|-----|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP080: BTEXN (QC | Lot: 5915806) | | | | | | | | |
| ES2422453-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |
| ES2422604-001 | Anonymous | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | 2 | 2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit |

Page : 5 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | | | |
|--|------------|--------|------|-------------------|---------------------------------------|--------------------|------------|------------|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| Method: Compound | AS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EA015: Total Dissolved Solids dried at 180 ± 5 °C (QCLot: 5918 | 8870) | | | | | | | | |
| EA015H: Total Dissolved Solids @180°C | | 10 | mg/L | <10 | 2000 mg/L | 101 | 87.0 | 109 | |
| | | | | <10 | 293 mg/L | 108 | 75.2 | 126 | |
| | | | | <10 | 2410 mg/L | 109 | 83.0 | 124 | |
| EA025: Total Suspended Solids dried at 104 ± 2°C (QCLot: 591 | 18869) | | | | | | | | |
| EA025H: Suspended Solids (SS) | | 5 | mg/L | <5 | 150 mg/L | 96.0 | 83.0 | 129 | |
| | | | | <5 | 1000 mg/L | 98.6 | 82.0 | 110 | |
| | | | | <5 | 928 mg/L | 96.9 | 83.0 | 118 | |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5914847) | | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.5 | 85.0 | 114 | |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 95.0 | 84.0 | 110 | |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.3 | 85.0 | 111 | |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 91.3 | 81.0 | 111 | |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 92.4 | 83.0 | 111 | |
| EG020A-F: Manganese | 7439-96-5 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 91.4 | 82.0 | 110 | |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 91.1 | 82.0 | 112 | |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 101 | 81.0 | 117 | |
| EG020A-F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 91.5 | 82.0 | 112 | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5914848) | | | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 97.6 | 83.0 | 105 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analyser | (QCLot: 59 | 18576) | | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 104 | 91.0 | 113 | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser (QCLot | : 5918574) | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 95.3 | 69.0 | 123 | |
| | | | | <0.1 | 1 mg/L | 99.7 | 70.0 | 123 | |
| | | | | <0.1 | 5 mg/L | 100 | 70.0 | 123 | |
| EK067G: Total Phosphorus as P by Discrete Analyser (QCLot: | 5918575) | | | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 99.0 | 71.3 | 126 | |
| | | | | <0.01 | 0.442 mg/L | 92.1 | 71.3 | 126 | |
| | | | | <0.01 | 1 mg/L | 102 | 70.0 | 130 | |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5912236) | | | | | | | | | |
| EP075(SIM): Phenol | 108-95-2 | 1 | μg/L | <1.0 | 5 μg/L | 38.1 | 24.5 | 61.9 | |

Page : 6 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|----------------------|-----|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP075(SIM)A: Phenolic Compounds (QCLot: 5912236) | - continued | | | | | | | |
| EP075(SIM): 2-Chlorophenol | 95-57-8 | 1 | μg/L | <1.0 | 5 μg/L | 73.8 | 52.0 | 90.0 |
| EP075(SIM): 2-Methylphenol | 95-48-7 | 1 | μg/L | <1.0 | 5 μg/L | 67.0 | 51.0 | 91.0 |
| EP075(SIM): 3- & 4-Methylphenol | 1319-77-3 | 2 | μg/L | <2.0 | 10 μg/L | 58.7 | 44.0 | 88.0 |
| EP075(SIM): 2-Nitrophenol | 88-75-5 | 1 | μg/L | <1.0 | 5 μg/L | 73.1 | 48.0 | 100 |
| EP075(SIM): 2.4-Dimethylphenol | 105-67-9 | 1 | μg/L | <1.0 | 5 μg/L | 67.5 | 49.0 | 99.0 |
| EP075(SIM): 2.4-Dichlorophenol | 120-83-2 | 1 | μg/L | <1.0 | 5 μg/L | 71.7 | 53.0 | 105 |
| EP075(SIM): 2.6-Dichlorophenol | 87-65-0 | 1 | μg/L | <1.0 | 5 μg/L | 76.0 | 57.0 | 105 |
| EP075(SIM): 4-Chloro-3-methylphenol | 59-50-7 | 1 | μg/L | <1.0 | 5 μg/L | 75.2 | 53.0 | 99.0 |
| EP075(SIM): 2.4.6-Trichlorophenol | 88-06-2 | 1 | μg/L | <1.0 | 5 μg/L | 67.4 | 50.0 | 106 |
| EP075(SIM): 2.4.5-Trichlorophenol | 95-95-4 | 1 | μg/L | <1.0 | 5 μg/L | 79.4 | 51.0 | 105 |
| EP075(SIM): Pentachlorophenol | 87-86-5 | 2 | μg/L | <2.0 | 10 μg/L | 38.5 | 10.0 | 95.0 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QC | Lot: 5912236) | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 68.0 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene | 208-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 69.8 | 63.6 | 114 |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 70.5 | 62.2 | 113 |
| EP075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 70.6 | 63.9 | 115 |
| EP075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 85.4 | 62.6 | 116 |
| EP075(SIM): Anthracene | 120-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 85.0 | 64.3 | 116 |
| EP075(SIM): Fluoranthene | 206-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 84.4 | 63.6 | 118 |
| EP075(SIM): Pyrene | 129-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 84.9 | 63.1 | 118 |
| EP075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 78.0 | 64.1 | 117 |
| EP075(SIM): Chrysene | 218-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 81.0 | 62.5 | 116 |
| EP075(SIM): Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1 | μg/L | <1.0 | 5 μg/L | 83.5 | 61.7 | 119 |
| EP075(SIM): Benzo(k)fluoranthene | 207-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 75.8 | 63.0 | 115 |
| EP075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 81.6 | 63.3 | 117 |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | 193-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 76.6 | 59.9 | 118 |
| EP075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 80.0 | 61.2 | 117 |
| EP075(SIM): Benzo(g.h.i)perylene | 191-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 78.2 | 59.1 | 118 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 591 | 2235) | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 59.4 | 53.7 | 97.0 |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 µg/L | 65.1 | 63.3 | 107 |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 92.2 | 58.3 | 120 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 591 | 5799) | | | | | | | |

Page : 7 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|---|---------------------|---------------|------|-------------------|---------------|------------------------------|--------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5915) | 799) - continue | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 79.8 | 75.0 | 127 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 59158 | 306) | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 85.9 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | Fractions (QC | Lot: 5912235) | | | | | | |
| EP071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 58.1 | 53.9 | 95.5 |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 61.8 | 57.8 | 110 |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 86.6 | 50.5 | 115 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | Fractions (QC | Lot: 5915799) | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 78.1 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 | Fractions (QC | Lot: 5915806) | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 87.0 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 5915799) | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 91.5 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 96.9 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 92.5 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 90.9 | 73.0 | 122 |
| | 106-42-3 | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 87.8 | 76.4 | 123 |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 96.8 | 75.5 | 124 |
| EP080: BTEXN (QCLot: 5915806) | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 94.8 | 68.3 | 119 |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 107 | 73.5 | 120 |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 99.6 | 73.8 | 122 |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 98.0 | 73.0 | 122 |
| ED000, atha Vidana | 106-42-3 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 400 | 76.4 | 400 |
| EP080: ortho-Xylene | 91-20-3 | 5 | | <5 | | 100 | 75.4 75.5 | 123 |
| EP080: Naphthalene | 91-20-3 | ð | μg/L | <5 | 10 μg/L | 102 | 70.5 | 124 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | Matrix Spike (MS) Report | | | | | |
|----------------------|-----------|--------------------------|------------|---------------|------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable l | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |

Page : 8 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | M | atrix Spike (MS) Report | | | | |
|----------------------|--|--------------------------------------|-------------------------|---------------|------------------|------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG020F: Dissolved | Metals by ICP-MS (QCLot: 5914847) | | | | | | |
| ES2422553-002 | SW2 | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 97.3 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 98.2 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 95.2 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 96.2 | 70.0 | 130 |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 91.9 | 70.0 | 130 |
| | | EG020A-F: Manganese | 7439-96-5 | 1 mg/L | 98.0 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 95.2 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 103 | 70.0 | 130 |
| EG035F: Dissolved | Mercury by FIMS (QCLot: 5914848) | | | | | | |
| ES2422553-001 | SW1 | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 88.0 | 70.0 | 130 |
| EK059G: Nitrite plu | us Nitrate as N (NOx) by Discrete Analyser (QCLot: 5 | 918576) | | | | | |
| ES2422660-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 114 | 70.0 | 130 |
| EK061G: Total Kiel | dahl Nitrogen By Discrete Analyser (QCLot: 5918574 | | | | | | |
| ES2422553-002 | SW2 | EK061G: Total Kjeldahl Nitrogen as N | | 50 mg/L | 88.9 | 70.0 | 130 |
| | sphorus as P by Discrete Analyser(QCLot: 5918575) | | | 3 | | | |
| ES2422553-002 | SW2 | EK067G: Total Phosphorus as P | | 10 mg/L | 102 | 70.0 | 130 |
| | etroleum Hydrocarbons (QCLot: 5915799) | Ertoor of Total Thoophorus do T | | 3 | | | |
| EN2406506-001 | Anonymous | EDOOD, CC. CO Frantier | <u></u> | 325 μg/L | 73.9 | 70.0 | 130 |
| | - | EP080: C6 - C9 Fraction | | 323 μg/L | 75.9 | 70.0 | 150 |
| | etroleum Hydrocarbons (QCLot: 5915806) | | | | | | |
| ES2422604-001 | Anonymous | EP080: C6 - C9 Fraction | | 325 μg/L | 77.2 | 70.0 | 130 |
| EP080/071: Total R | ecoverable Hydrocarbons - NEPM 2013 Fractions(Q | CLot: 5915799) | | | | | |
| EN2406506-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 75.9 | 70.0 | 130 |
| EP080/071: Total R | ecoverable Hydrocarbons - NEPM 2013 Fractions (Qu | CLot: 5915806) | | | | | |
| ES2422604-001 | Anonymous | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 76.3 | 70.0 | 130 |
| EP080: BTEXN (QC | CLot: 5915799) | | | | | | |
| EN2406506-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 95.3 | 70.0 | 130 |
| | , then, mead | EP080: Toluene | 108-88-3 | 25 μg/L | 99.6 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 106 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 104 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 103 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 109 | 70.0 | 130 |
| EP080: BTEXN (QC | CLot: 5915806) | | | | | | |
| ES2422604-001 | Anonymous | EP080: Benzene | 71-43-2 | 25 μg/L | 81.6 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 92.5 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 87.1 | 70.0 | 130 |

Page : 9 of 9
Work Order : ES2422553

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | Matrix Spike (MS) Report | | | | | |
|----------------------|-----------------------------|----------------------------|------------|---------------|------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable i | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP080: BTEXN (| QCLot: 5915806) - continued | | | | | | |
| ES2422604-001 | Anonymous | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 85.9 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 89.1 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 104 | 70.0 | 130 |

senversa

Chain of Custody Documentation

13096 Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. Please forward to Eurofins 41/11 Time: 571 Date: Date: Time: ОГО Date: | Time: | Date/Time: | Date/Tim REGIMA NT bns 9 ×× MATER - EP080 NXETX × Thenh NATER - W-18 FRH(C6 - C9)/BTEXN Signature: NSTER - W-26 REH/BTEXN/PAH/8 Metals × × × × Name/Signature: Vame/Signature Name/Signature: WATER - EG005F × × × Rowan Faint × Total Bottles Sampler Name 47 Container Information Method of Shipment (if applicable) of 1 0420 218 472 1xP, 2xVSA, 1xN, 1xUA, 1xSP xP, 2xVSA, 1xN, 1xUA, 1xSP Standard EN1103\21 2xVSA, 1xN, 1xUA, 1xSP 2xVSA, 1xN, 1xUA, 1xSP 2xVSA, 1xN, 1xUA, 1xSP Type / Code Sarrier / Reference #: Carrier / Reference #: Samier / Reference #: Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project VSA VSA Sample Receipt Date/Time: ALS NSW Turn Around Time: Purchase Order: Phone/Mobile: Time Laboratory: Address: Contact: Phone: Quote No: Page: Date: Date: Date: Time: Sydney Work Order Reference ES2423038 Environmental Division 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 11/07/2024 emma.walsh@senversa.com.au Emma Walsh rowan.faint@senversa.com.au Date Wetherill Park WME Rowan Faint specifications were used during the collection of these samples: Talephone : + 61-2-878* 8555 Sample Information Matrix * Water Rowan Faint Sample ID QC405 QC204 QC304 QC104 QC505 MW6 MW2 MW3 MW1 MW4 Senversa Pty Ltd www.senversa.com.au ABN 89 132 231 380 oject Manager: mail Report To: Relinquished By: lame/Signature: Vame/Signature: dame/Signature: roject Name ampled By: Lab ID 2 20 1 0 3 2 1

Completed by: Checked by:

V = VOA Vial Hydocholro Add (HOI) Preserved: VS = VOA Vial Sulphurio Preserved. VSA = Sulphurio Preserved Amber Glass; H = HOI Preserved Plastic; HS = HOI Preserved Speciation Bottle; SV = Sulphurio Preserved Dottle; BA Preserved Bottle; ST = Sterile Bottle; UA = Unpriserved Amber Glass; L= Lugot's lodine preserved white plastic bottle; SW= sulfurir and preserved wide mouth glass jar

S20102 GW COC



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2423038

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW Address : 277-289 Woodpark Road Smithfield

NSW Australia 2164

SYDNEY NSW 2000

2000

E-mail : Emma.Walsh@senversa.com.au E-mail : sandy.phan@alsqlobal.com

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555 Facsimile : ---- Facsimile : +61-2-8784 8500

Project : S20102 Wetherill Park WME Page : 1 of 3

 Order number
 : --- Quote number
 : EB2023SENVER0001 (EN/000)

 C-O-C number
 : --- QC Level
 : NEPM 2013 B3 & ALS QC Standard

Sampler : Rowan Faint

Dates

Date Samples Received : 12-Jul-2024 17:15 Issue Date : 12-Jul-2024
Client Requested Due : 22-Jul-2024
Scheduled Reporting Date : 22-Jul-2024

Date

Mode of Delivery : Client Drop Off Security Seal : Not Available

No. of coolers/boxes : 1 Temperature : 18.2'C, 15.3'C & 12.4'C -

Ice present

Receipt Detail : No. of samples received / analysed : 9 / 9

General Comments

Delivery Details

• This report contains the following information:

- Sample Container(s)/Preservation Non-Compliances
- Summary of Sample(s) and Requested Analysis
- Proactive Holding Time Report
- Requested Deliverables
- The sampling date listed on the COC is 11/07/24, however the soil jar was dated 08/07/24 for sample 8 & 9.
- Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.
- Sample(s) requiring volatile organic compound analysis received in airtight containers (ZHE).
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Unless otherwise stated, analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical
 analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this
 temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS
 recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.

: 12-Jul-2024 Issue Date

Page

: 2 of 3 : ES2423038 Amendment 0 Work Order Client : SENVERSA PTY LTD



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

• No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

| process necessal tasks. Packages as the determin tasks, that are included in the sampling default 00:00 on | may contain ad ation of moisture uded in the package. time is provided, the date of sampling date wi | be part of a laboratory on of client requested ditional analyses, such content and preparation the sampling time will be assumed by the ckets without a time | WATER - EG005F Dissolved Metals by ICPAES | WATER - EP080 BTEXN | WATER - NT-08 Total Nitrogen + NO2 + NO3 + NH3 + Total P | WATER - NT-11 Total Nitrogen and Total Phosphorus | WATER - W-18 TRH(C6 - C9)/BTEXN | WATER - W-26 TRH/BTEXN/PAH/8 Metals |
|--|--|--|--|------------------------|---|--|------------------------------------|--|
| ES2423038-001 | 11-Jul-2024 00:00 | MW1 | ✓ | | ✓ | | | ✓ |
| ES2423038-002 | 11-Jul-2024 00:00 | MW2 | ✓ | | ✓ | | | 1 |
| ES2423038-003 | 11-Jul-2024 00:00 | MW3 | ✓ | | ✓ | | | ✓ |
| ES2423038-004 | 11-Jul-2024 00:00 | MW4 | ✓ | | ✓ | | | ✓ |
| ES2423038-005 | 11-Jul-2024 00:00 | MW6 | ✓ | | ✓ | | | ✓ |
| ES2423038-006 | 11-Jul-2024 00:00 | QC104 | ✓ | | | ✓ | | ✓ |
| ES2423038-007 | 11-Jul-2024 00:00 | QC304 | ✓ | | | 1 | | ✓ |
| ES2423038-008 | 08-Jul-2024 00:00 | QC405 | | | | | ✓ | |
| ES2423038-009 | 08-Jul-2024 00:00 | QC505 | | 1 | | | | |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

BEC CHAPPLE

| - *AU Certificate of Analysis - NATA (COA) | Email | bec.chapple@senversa.com.au |
|---|-------|---------------------------------|
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | bec.chapple@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | bec.chapple@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | bec.chapple@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | bec.chapple@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | bec.chapple@senversa.com.au |
| EMMA GUY | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | emma.guy@senversa.com.au |
| - *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | emma.guy@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | emma.guy@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | emma.guy@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | emma.guy@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | emma.guy@senversa.com.au |
| EMMA WALSH | | |
| - *AU Certificate of Analysis - NATA (COA) | Email | Emma.Walsh@senversa.com.au |
| *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) | Email | Emma.Walsh@senversa.com.au |
| - *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Sample Receipt Notification - Environmental HT (SRN) | Email | Emma.Walsh@senversa.com.au |
| - A4 - AU Tax Invoice (INV) | Email | Emma.Walsh@senversa.com.au |
| - Chain of Custody (CoC) (COC) | Email | Emma.Walsh@senversa.com.au |
| - EDI Format - ESDAT (ESDAT) | Email | Emma.Walsh@senversa.com.au |
| SUPPLIER ACCOUNTS | | |
| - A4 - AU Tax Invoice (INV) | Email | supplieraccounts@senversa.com.a |
| | | u |

: 12-Jul-2024 Issue Date

Page

: 3 of 3 : ES2423038 Amendment 0 Work Order Client : SENVERSA PTY LTD





CERTIFICATE OF ANALYSIS

Work Order : ES2423038

Client : SENVERSA PTY LTD

Contact : EMMA WALSH

Address : Level 24, 1 Market St, Sydney NSW 2000

SYDNEY NSW 2000

Telephone : 02 8252 0000

Project : S20102 Wetherill Park WME

Order number : ---C-O-C number : ----

Sampler : Rowan Faint

Site : ---

Quote number : EN/000

No. of samples received : 9
No. of samples analysed : 9

Page : 1 of 9

Laboratory : Environmental Division Sydney

Contact : Sandy Phan

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : +61-2-8784 8555

Date Samples Received : 12-Jul-2024 17:15

Date Analysis Commenced : 13-Jul-2024

Issue Date : 22-Jul-2024 16:26



ed by ALS. This document shall

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW

Edwardy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key: CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

- ^ = This result is computed from individual analyte detections at or above the level of reporting
- ø = ALS is not NATA accredited for these tests.
- ~ = Indicates an estimated value.
- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- Unless otherwise stated, analytical work for this work order will be conducted at ALS Sydney.
- EP080: Sample TRIP SPIKE contains volatile compounds spiked into the sample containers prior to dispatch from the laboratory. BTEXN compounds spiked at 20 ug/L.

Page : 3 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|--------------------------------------|----------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 11-Jul-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2423038-001 | ES2423038-002 | ES2423038-003 | ES2423038-004 | ES2423038-005 |
| | | | | Result | Result | Result | Result | Result |
| EG005(ED093)F: Dissolved Metals b | y ICP-AES | | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 87.2 | 2.30 | 7.01 | 1.90 | <0.05 |
| Manganese | 7439-96-5 | 0.01 | mg/L | 4.84 | 3.28 | 6.79 | 4.00 | <0.01 |
| EG020F: Dissolved Metals by ICP-M | S | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.007 | 0.008 | 0.003 | 0.007 | <0.001 |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | 0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Copper | 7440-50-8 | 0.001 | mg/L | 0.003 | <0.001 | 0.002 | <0.001 | <0.001 |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.156 | 0.006 | 0.200 | 0.011 | <0.001 |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.174 | <0.005 | 0.243 | <0.005 | <0.005 |
| EG035F: Dissolved Mercury by FIMS | S | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| EK055G: Ammonia as N by Discrete | Analyser | | | | | | | |
| Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.19 | 0.26 | 0.28 | 0.28 | <0.01 |
| EK057G: Nitrite as N by Discrete Ar | nalyser | | | | | | | |
| Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| EK058G: Nitrate as N by Discrete A | nalyser | | | | | | | |
| Nitrate as N | 14797-55-8 | 0.01 | mg/L | <0.01 | 0.02 | 0.01 | <0.01 | 1.64 |
| EK059G: Nitrite plus Nitrate as N (N | IOx) by Discrete Ana | lyser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.02 | 0.01 | <0.01 | 1.64 |
| EK061G: Total Kjeldahl Nitrogen By | Discrete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 1.1 | 0.6 | 0.6 | 0.4 | 0.3 |
| EK062G: Total Nitrogen as N (TKN + | NOx) by Discrete An | alyser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 1.1 | 0.6 | 0.6 | 0.4 | 1.9 |
| EK067G: Total Phosphorus as P by | Discrete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.04 | 0.05 | 0.04 | 0.04 | 0.05 |
| EP075(SIM)B: Polynuclear Aromatic | 1 | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |

Page : 4 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|--|--------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| (many to the large) | | Samplii | ng date / time | 11-Jul-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2423038-001 | ES2423038-002 | ES2423038-003 | ES2423038-004 | ES2423038-005 |
| | | | | Result | Result | Result | Result | Result |
| EP075(SIM)B: Polynuclear Aromatic H | ydrocarbons - Cont | | | | | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Sum of polycyclic aromatic hydrocarbon | ns | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | <0.5 | <0.5 | <0.5 |
| EP080/071: Total Petroleum Hydrocarl | bons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |
| C10 - C14 Fraction | | 50 | μg/L | 250 | <50 | <50 | <50 | <50 |
| C15 - C28 Fraction | | 100 | μg/L | 170 | <100 | 810 | <100 | <100 |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | <50 | <50 | <50 |
| C10 - C36 Fraction (sum) | | 50 | μg/L | 420 | <50 | 810 | <50 | <50 |
| EP080/071: Total Recoverable Hydroc | arbons - NEPM 201 | 3 Fraction | าร | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |
| C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | <20 | <20 |

Page : 5 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | MW1 | MW2 | MW3 | MW4 | MW6 |
|---|-------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | Sampli | ng date / time | 11-Jul-2024 00:00 |
| Compound | CAS Number | LOR | Unit | ES2423038-001 | ES2423038-002 | ES2423038-003 | ES2423038-004 | ES2423038-005 |
| | | | | Result | Result | Result | Result | Result |
| EP080/071: Total Recoverable Hydroc | arbons - NEPM 201 | | | | | | | |
| >C10 - C16 Fraction | | 100 | μg/L | 260 | <100 | <100 | <100 | <100 |
| >C16 - C34 Fraction | | 100 | μg/L | 260 | <100 | 400 | <100 | <100 |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | <100 | <100 | <100 |
| ^ >C10 - C40 Fraction (sum) | | 100 | μg/L | 520 | <100 | 400 | <100 | <100 |
| ^ >C10 - C16 Fraction minus Naphthalene (F2) | | 100 | μg/L | 260 | <100 | <100 | <100 | <100 |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | <1 | <1 |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | <2 | <2 |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | <2 | <2 |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | <2 | <2 |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | <2 | <2 |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | <2 | <2 |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | <1 | <1 |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | <5 | <5 |
| EP075(SIM)S: Phenolic Compound Su | rrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 31.6 | 30.9 | 20.2 | 24.7 | 27.4 |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 62.0 | 59.3 | 38.7 | 52.2 | 55.7 |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 74.8 | 62.3 | 46.5 | 44.2 | 52.6 |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 74.0 | 73.3 | 46.8 | 66.1 | 72.7 |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 81.8 | 77.4 | 60.8 | 72.8 | 76.4 |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 85.2 | 84.8 | 58.0 | 78.3 | 82.7 |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 96.6 | 98.2 | 104 | 100 | 99.9 |
| Toluene-D8 | 2037-26-5 | 2 | % | 89.4 | 95.8 | 94.0 | 95.7 | 95.9 |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 109 | 113 | 115 | 113 | 116 |

Page : 6 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC104 | QC304 | QC405 | QC505 | |
|--------------------------------------|---------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| | | Sampli | ng date / time | 11-Jul-2024 00:00 | 11-Jul-2024 00:00 | 08-Jul-2024 00:00 | 08-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2423038-006 | ES2423038-007 | ES2423038-008 | ES2423038-009 | |
| | | | | Result | Result | Result | Result | |
| EG005(ED093)F: Dissolved Metals b | _ | | | | | | | |
| Iron | 7439-89-6 | 0.05 | mg/L | 6.96 | <0.05 | | | |
| Manganese | 7439-96-5 | 0.01 | mg/L | 6.80 | <0.01 | | | |
| EG020F: Dissolved Metals by ICP-M | S | | | | | | | |
| Arsenic | 7440-38-2 | 0.001 | mg/L | 0.003 | <0.001 | | | |
| Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | | | |
| Nickel | 7440-02-0 | 0.001 | mg/L | 0.200 | <0.001 | | | |
| Zinc | 7440-66-6 | 0.005 | mg/L | 0.239 | <0.005 | | | |
| EG035F: Dissolved Mercury by FIMS | 3 | | | | | | | |
| Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | | | |
| EK059G: Nitrite plus Nitrate as N (N | Ox) by Discrete Ana | lyser | | | | | | |
| Nitrite + Nitrate as N | | 0.01 | mg/L | 0.02 | 3.34 | | | |
| EK061G: Total Kjeldahl Nitrogen By | Discrete Analyser | | | | | | | |
| Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.6 | 0.1 | | | |
| EK062G: Total Nitrogen as N (TKN + | NOx) by Discrete An | alyser | | | | | | |
| ^ Total Nitrogen as N | | 0.1 | mg/L | 0.6 | 3.4 | | | |
| EK067G: Total Phosphorus as P by | Discrete Analyser | | | | | | | |
| Total Phosphorus as P | | 0.01 | mg/L | 0.04 | <0.01 | | | |
| EP075(SIM)B: Polynuclear Aromatic | Hydrocarbons | | | | | | | |
| Naphthalene | 91-20-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthylene | 208-96-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Acenaphthene | 83-32-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Fluorene | 86-73-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Phenanthrene | 85-01-8 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Anthracene | 120-12-7 | 1.0 | μg/L | <1.0 | <1.0 | | | |

Page : 7 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC104 | QC304 | QC405 | QC505 | |
|--|--------------------|------------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| , | | Sampli | ng date / time | 11-Jul-2024 00:00 | 11-Jul-2024 00:00 | 08-Jul-2024 00:00 | 08-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2423038-006 | ES2423038-007 | ES2423038-008 | ES2423038-009 | |
| | | | | Result | Result | Result | Result | |
| EP075(SIM)B: Polynuclear Aromatic H | ydrocarbons - Cont | inued | | | | | | |
| Fluoranthene | 206-44-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Pyrene | 129-00-0 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benz(a)anthracene | 56-55-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Chrysene | 218-01-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(b+j)fluoranthene | 205-99-2 205-82-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(k)fluoranthene | 207-08-9 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | <0.5 | | | |
| Indeno(1.2.3.cd)pyrene | 193-39-5 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Dibenz(a.h)anthracene | 53-70-3 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Benzo(g.h.i)perylene | 191-24-2 | 1.0 | μg/L | <1.0 | <1.0 | | | |
| Sum of polycyclic aromatic hydrocarbon | ıs | 0.5 | μg/L | <0.5 | <0.5 | | | |
| Benzo(a)pyrene TEQ (zero) | | 0.5 | μg/L | <0.5 | <0.5 | | | |
| EP080/071: Total Petroleum Hydrocarb | oons | | | | | | | |
| C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | <20 | | |
| C10 - C14 Fraction | | 50 | μg/L | <50 | <50 | | | |
| C15 - C28 Fraction | | 100 | μg/L | 800 | <100 | | | |
| C29 - C36 Fraction | | 50 | μg/L | <50 | <50 | | | |
| C10 - C36 Fraction (sum) | | 50 | μg/L | 800 | <50 | | | |
| EP080/071: Total Recoverable Hydroca | arbons - NEPM 201 | 3 Fraction | ns | | | | | |
| C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | <20 | | |
| C6 - C10 Fraction minus BTEX (F1) | C6_C10-BTEX | 20 | μg/L | <20 | <20 | <20 | | |
| >C10 - C16 Fraction | | 100 | μg/L | <100 | <100 | | | |
| >C16 - C34 Fraction | | 100 | μg/L | 480 | <100 | | | |
| >C34 - C40 Fraction | | 100 | μg/L | <100 | <100 | | | |
| >C10 - C40 Fraction (sum) | | 100 | μg/L | 480 | <100 | | | |
| >C10 - C16 Fraction minus Naphthalene (F2) | | 100 | μg/L | <100 | <100 | | | |

Page : 8 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER (Matrix: WATER) | | | Sample ID | QC104 | QC304 | QC405 | QC505 | |
|-----------------------------------|-------------------|--------|----------------|-------------------|-------------------|-------------------|-------------------|--|
| (Maurix. WATER) | | Sampli | ng date / time | 11-Jul-2024 00:00 | 11-Jul-2024 00:00 | 08-Jul-2024 00:00 | 08-Jul-2024 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2423038-006 | ES2423038-007 | ES2423038-008 | ES2423038-009 | |
| | | | | Result | Result | Result | Result | |
| EP080: BTEXN | | | | | | | | |
| Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | <1 | 14 | |
| Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | <2 | 15 | |
| Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | <2 | 14 | |
| meta- & para-Xylene | 108-38-3 106-42-3 | 2 | μg/L | <2 | <2 | <2 | 14 | |
| ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | <2 | 16 | |
| ^ Total Xylenes | | 2 | μg/L | <2 | <2 | <2 | 30 | |
| ^ Sum of BTEX | | 1 | μg/L | <1 | <1 | <1 | 73 | |
| Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | <5 | 19 | |
| EP075(SIM)S: Phenolic Compound | Surrogates | | | | | | | |
| Phenol-d6 | 13127-88-3 | 1.0 | % | 33.2 | 30.2 | | | |
| 2-Chlorophenol-D4 | 93951-73-6 | 1.0 | % | 66.6 | 65.3 | | | |
| 2.4.6-Tribromophenol | 118-79-6 | 1.0 | % | 77.0 | 66.1 | | | |
| EP075(SIM)T: PAH Surrogates | | | | | | | | |
| 2-Fluorobiphenyl | 321-60-8 | 1.0 | % | 73.7 | 70.9 | | | |
| Anthracene-d10 | 1719-06-8 | 1.0 | % | 90.4 | 90.0 | | | |
| 4-Terphenyl-d14 | 1718-51-0 | 1.0 | % | 97.0 | 97.4 | | | |
| EP080S: TPH(V)/BTEX Surrogates | | | | | | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 2 | % | 98.6 | 103 | 100 | 104 | |
| Toluene-D8 | 2037-26-5 | 2 | % | 91.8 | 97.1 | 90.7 | 85.5 | |
| 4-Bromofluorobenzene | 460-00-4 | 2 | % | 109 | 118 | 112 | 117 | |

Page : 9 of 9
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery | Limits (%) |
|---|------------|----------|------------|
| Compound | CAS Number | Low | High |
| EP075(SIM)S: Phenolic Compound Surrogates | | | |
| Phenol-d6 | 13127-88-3 | 10 | 44 |
| 2-Chlorophenol-D4 | 93951-73-6 | 14 | 94 |
| 2.4.6-Tribromophenol | 118-79-6 | 17 | 125 |
| EP075(SIM)T: PAH Surrogates | | | |
| 2-Fluorobiphenyl | 321-60-8 | 20 | 104 |
| Anthracene-d10 | 1719-06-8 | 27 | 113 |
| 4-Terphenyl-d14 | 1718-51-0 | 32 | 112 |
| EP080S: TPH(V)/BTEX Surrogates | | | |
| 1.2-Dichloroethane-D4 | 17060-07-0 | 72 | 143 |
| Toluene-D8 | 2037-26-5 | 75 | 131 |
| 4-Bromofluorobenzene | 460-00-4 | 73 | 137 |



QA/QC Compliance Assessment to assist with Quality Review

Work Order : **ES2423038** Page : 1 of 8

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

 Contact
 : EMMA WALSH
 Telephone
 : +61-2-8784 8555

 Project
 : S20102 Wetherill Park WME
 Date Samples Received
 : 12-Jul-2024

 Site
 : --- Issue Date
 : 22-Jul-2024

Sampler : Rowan Faint No. of samples received : 9

Order number : ---- No. of samples analysed : 9

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers: Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- Matrix Spike outliers exist please see following pages for full details.
- For all regular sample matrices, where applicable to the methodology, NO surrogate recovery outliers occur.

Outliers: Analysis Holding Time Compliance

• NO Analysis Holding Time Outliers exist.

Outliers: Frequency of Quality Control Samples

Quality Control Sample Frequency Outliers exist - please see following pages for full details.

Page : 2 of 8 Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME

ALS

Outliers: Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: WATER

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|---|----------------------|------------------|------------------------|------------|------------|--------|----------------------------------|
| Matrix Spike (MS) Recoveries | | | | | | | |
| EG005(ED093)F: Dissolved Metals by ICP-AES | ES2423038001 | MW1 | Manganese | 7439-96-5 | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete A | EN2406968001 | Anonymous | Nitrite + Nitrate as N | | Not | | MS recovery not determined, |
| | | | | | Determined | | background level greater than or |
| | | | | | | | equal to 4x spike level. |

Outliers: Frequency of Quality Control Samples

Matrix: WATER

| WOULK, WATER | | | | | | |
|-----------------------------|------------|----|---------|--------|----------|--------------------------------|
| Quality Control Sample Type | | Co | unt | Rate | e (%) | Quality Control Specification |
| Analytical Methods | Method | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 10 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 10 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 10 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 10 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for <u>VOC in soils</u> vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive <u>or</u> Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: **×** = Holding time breach ; ✓ = Within holding time.

| madistri in the little | | | | | | | , | g |
|--|----------|-------------|----------------|------------------------|------------|---------------|------------------|------------|
| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG005(ED093)F: Dissolved Metals by ICI | P-AES | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered | (EG005F) | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | | | | 17-Jul-2024 | 07-Jan-2025 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |

Page : 3 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Method | | Sample Date | Ex | traction / Preparation | | | Analysis | |
|---|--------|-------------|----------------|------------------------|------------|---------------|------------------|------------|
| Container / Client Sample ID(s) | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EG020F: Dissolved Metals by ICP-MS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | | | | 17-Jul-2024 | 07-Jan-2025 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EG035F: Dissolved Mercury by FIMS | | | | | | | | |
| Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | | | | 18-Jul-2024 | 08-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EK055G: Ammonia as N by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK055G) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | | | | 18-Jul-2024 | 08-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6 | | | | | | | | |
| EK057G: Nitrite as N by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Natural (EK057G) | | | | | | | 40 1 1 0004 | |
| MW1, | MW2, | 11-Jul-2024 | | | | 13-Jul-2024 | 13-Jul-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6 | | | | | | | | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Ana | ılyser | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK059G) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | | | | 18-Jul-2024 | 08-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK061G) | | | | 00.4 | _ | | | |
| MW1, | MW2, | 11-Jul-2024 | 17-Jul-2024 | 08-Aug-2024 | ✓ | 18-Jul-2024 | 08-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EK067G: Total Phosphorus as P by Discrete Analyser | | | | | | | | |
| Clear Plastic Bottle - Sulfuric Acid (EK067G) | | | | 00.4 005. | _ | | 00.4 005: | |
| MW1, | MW2, | 11-Jul-2024 | 17-Jul-2024 | 08-Aug-2024 | ✓ | 18-Jul-2024 | 08-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |

Page : 4 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: ▼ = Holding time breach; ✓ = Within holding time.

| Method | | Sample Date | Ev | traction / Preparation | Lvaldation | Holding time | Analysis | IT Holding time |
|--|----------------|----------------------------|----------------------------|----------------------------|------------|----------------------------|----------------------------|--|
| Container / Client Sample ID(s) | | Sample Date | | | Francisco | 5 | | Fratrotion |
| | | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) | | | | 40 1-1-0004 | | | 04 4 0004 | |
| MW1, | MW2, | 11-Jul-2024 | 15-Jul-2024 | 18-Jul-2024 | ✓ | 17-Jul-2024 | 24-Aug-2024 | ✓ |
| MW3 | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP075(SIM)) | | 44 1 1 0004 | 45 1 1 0004 | 40 1.1 0004 | | 40 1 1 0004 | 04 4 0004 | |
| MW4, | MW6, | 11-Jul-2024 | 15-Jul-2024 | 18-Jul-2024 | ✓ | 18-Jul-2024 | 24-Aug-2024 | ✓ |
| QC104, | QC304 | | | | | | | |
| EP080/071: Total Petroleum Hydrocarbons | | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | 15-Jul-2024 | 18-Jul-2024 | ✓ | 18-Jul-2024 | 24-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| QC405 | | 08-Jul-2024 | 15-Jul-2024 | 22-Jul-2024 | ✓ | 15-Jul-2024 | 22-Jul-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | 15-Jul-2024 | 25-Jul-2024 | ✓ | 15-Jul-2024 | 25-Jul-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EP080/071: Total Recoverable Hydrocarbons - NEPM | 2013 Fractions | | | | | | | |
| Amber Glass Bottle - Unpreserved (EP071) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | 15-Jul-2024 | 18-Jul-2024 | ✓ | 18-Jul-2024 | 24-Aug-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| QC405 | | 08-Jul-2024 | 15-Jul-2024 | 22-Jul-2024 | ✓ | 15-Jul-2024 | 22-Jul-2024 | ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| MW1, | MW2, | 11-Jul-2024 | 15-Jul-2024 | 25-Jul-2024 | ✓ | 15-Jul-2024 | 25-Jul-2024 | ✓ |
| MW3, | MW4, | | | | | | | |
| MW6, | QC104, | | | | | | | |
| QC304 | | | | | | | | |
| EP080: BTEXN | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| | | | | | | | | T. Control of the Con |
| QC405, | QC505 | 08-Jul-2024 | 15-Jul-2024 | 22-Jul-2024 | ✓ | 15-Jul-2024 | 22-Jul-2024 | ✓ |
| QC405, Amber VOC Vial - Sulfuric Acid (EP080) | QC505 | 08-Jul-2024 | 15-Jul-2024 | 22-Jul-2024 | ✓ | 15-Jul-2024 | 22-Jul-2024 | ✓ |
| · · · · · · · · · · · · · · · · · · · | QC505 MW2, | 08-Jul-2024 11-Jul-2024 | 15-Jul-2024 15-Jul-2024 | 22-Jul-2024 25-Jul-2024 | <i>J</i> | 15-Jul-2024 15-Jul-2024 | 22-Jul-2024 25-Jul-2024 | ✓ ✓ |
| Amber VOC Vial - Sulfuric Acid (EP080) | | | | | | | | |
| Amber VOC Vial - Sulfuric Acid (EP080) MW1, | MW2, | | | | | | | |

Page : 5 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

the expected rate. A listing of breaches is provided in the Summary of Oddiers.

| Matrix: WATER | | | | Evaluatio | n: × = Quality Co | not within specification; ✓ = Quality Control frequency within specification. | |
|---|------------|----|---------|-----------|-------------------|---|--------------------------------|
| Quality Control Sample Type | | Co | ount | | Rate (%) | | Quality Control Specification |
| Analytical Methods | Method | QC | Reaular | Actual | Expected | Evaluation | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 2 | 7 | 28.57 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 4 | 32 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 2 | 15 | 13.33 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 0 | 10 | 0.00 | 10.00 | × | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 4 | 40 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 2 | 20 | 10.00 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 0 | 10 | 0.00 | 10.00 | 3¢ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 2 | 16 | 12.50 | 10.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 32 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 15 | 6.67 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 6 | 40 | 15.00 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 3 | 20 | 15.00 | 15.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Ammonia as N by Discrete analyser | EK055G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Mercury by FIMS | EG035F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-AES | EG005F | 1 | 7 | 14.29 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | 2 | 32 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Nitrite as N by Discrete Analyser | EK057G | 1 | 15 | 6.67 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | 2 | 40 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Total Phosphorus as P By Discrete Analyser | EK067G | 1 | 20 | 5.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH - Semivolatile Fraction | EP071 | 1 | 10 | 10.00 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| TRH Volatiles/BTEX | EP080 | 1 | 16 | 6.25 | 5.00 | ✓ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |

Page : 6 of 8 Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Matrix: WATER Evaluation: * = Quality Control frequency not within specification; * = Quality Control frequency within specification. Quality Control Sample Type Count Rate (%) Quality Control Specification Analytical Methods Method QC Evaluation Regular Actual Expected Matrix Spikes (MS) - Continued Ammonia as N by Discrete analyser 20 5.00 NEPM 2013 B3 & ALS QC Standard EK055G 1 5.00 1 Dissolved Mercury by FIMS 1 20 5.00 5.00 NEPM 2013 B3 & ALS QC Standard EG035F 1 7 Dissolved Metals by ICP-AES 1 14.29 5.00 NEPM 2013 B3 & ALS QC Standard EG005F ✓ Dissolved Metals by ICP-MS - Suite A 1 20 5.00 NEPM 2013 B3 & ALS QC Standard EG020A-F 5.00 1 Nitrite and Nitrate as N (NOx) by Discrete Analyser EK059G 2 32 6.25 5.00 1 NEPM 2013 B3 & ALS QC Standard Nitrite as N by Discrete Analyser 1 15 NEPM 2013 B3 & ALS QC Standard 6.67 5.00 EK057G ✓ PAH/Phenols (GC/MS - SIM) 0 10 0.00 5.00 NEPM 2013 B3 & ALS QC Standard EP075(SIM) × Total Kjeldahl Nitrogen as N By Discrete Analyser 2 40 5.00 5.00 NEPM 2013 B3 & ALS QC Standard EK061G 1 Total Phosphorus as P By Discrete Analyser 20 1 5.00 5.00 1 NEPM 2013 B3 & ALS QC Standard EK067G TRH - Semivolatile Fraction 0 10 0.00 NEPM 2013 B3 & ALS QC Standard EP071 5.00 × TRH Volatiles/BTEX EP080 1 16 6.25 5.00 NEPM 2013 B3 & ALS QC Standard

Page : 7 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|---|----------|--------|---|
| Dissolved Metals by ICP-AES | EG005F | WATER | In house: Referenced to APHA 3120; USEPA SW 846 - 6010. The ICPAES technique ionises the 0.45µm filtered samples, emitting a characteristic spectrum which is compared against matrix matched standards. This method is compliant with NEPM Schedule B(3). |
| Dissolved Metals by ICP-MS - Suite A | EG020A-F | WATER | In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector. |
| Dissolved Mercury by FIMS | EG035F | WATER | In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl2)(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl2 which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3). |
| Ammonia as N by Discrete analyser | EK055G | WATER | In house: Referenced to APHA 4500-NH3 G Ammonia is determined by direct colorimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrite as N by Discrete Analyser | EK057G | WATER | In house: Referenced to APHA 4500-NO2- B. Nitrite is determined by direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Nitrate as N by Discrete Analyser | EK058G | WATER | In house: Referenced to APHA 4500-NO3- F. Nitrate is reduced to nitrite by way of a chemical reduction followed by quantification by Discrete Analyser. Nitrite is determined seperately by direct colourimetry and result for Nitrate calculated as the difference between the two results. This method is compliant with NEPM Schedule B(3) |
| Nitrite and Nitrate as N (NOx) by Discrete Analyser | EK059G | WATER | In house: Referenced to APHA 4500-NO3- F. Combined oxidised Nitrogen (NO2+NO3) is determined by Chemical Reduction and direct colourimetry by Discrete Analyser. This method is compliant with NEPM Schedule B(3) |
| Total Kjeldahl Nitrogen as N By Discrete Analyser | EK061G | WATER | In house: Referenced to APHA 4500-Norg D (In house). An aliquot of sample is digested using a high temperature Kjeldahl digestion to convert nitrogenous compounds to ammonia. Ammonia is determined colorimetrically by discrete analyser. This method is compliant with NEPM Schedule B(3) |
| Total Nitrogen as N (TKN + Nox) By Discrete Analyser | EK062G | WATER | In house: Referenced to APHA 4500-Norg / 4500-NO3 This method is compliant with NEPM Schedule B(3) |
| Total Phosphorus as P By Discrete Analyser | EK067G | WATER | In house: Referenced to APHA 4500-P H, Jirka et al, Zhang et al. This procedure involves sulphuric acid digestion of a sample aliquot to break phosphorus down to orthophosphate. The orthophosphate reacts with ammonium molybdate and antimony potassium tartrate to form a complex which is then reduced and its concentration measured at 880nm using discrete analyser. This method is compliant with NEPM Schedule B(3) |
| TRH - Semivolatile Fraction | EP071 | WATER | In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3) |

Page : 8 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Analytical Methods | Method | Matrix | Method Descriptions |
|--------------------------------------|-----------------------|------------------------|--|
| PAH/Phenols (GC/MS - SIM) | EP075(SIM) | WATER | In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode |
| | | | and quantification is by comparison against an established 5 point calibration curve. This method is compliant |
| | | | with NEPM Schedule B(3) |
| TRH Volatiles/BTEX | EP080 | WATER | In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary |
| | | | GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a |
| | | | sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This |
| | | | method is compliant with the QC requirements of NEPM Schedule B(3) |
| | | | |
| Preparation Methods | Method | Matrix | Method Descriptions |
| Preparation Methods TKN/TP Digestion | Method EK061/EK067 | <i>Matrix</i> WATER | Method Descriptions In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| | | | |
| | | | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel |
| TKN/TP Digestion | EK061/EK067 | WATER | In house: Referenced to APHA 4500 Norg - D; APHA 4500 P - H. This method is compliant with NEPM Schedule B(3) In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated |



QUALITY CONTROL REPORT

Work Order : **ES2423038** Page : 1 of 8

Client : SENVERSA PTY LTD Laboratory : Environmental Division Sydney

Contact : EMMA WALSH Contact : Sandy Phan

Address : Level 24, 1 Market St, Sydney NSW 2000 Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

SYDNEY NSW 2000

Telephone : 02 8252 0000 Telephone : +61-2-8784 8555

Project : \$20102 Wetherill Park WME Date Samples Received : 12-Jul-2024

Order number Date Analysis Commenced : 13-Jul-2024

C-O-C number : ---- Issue Date

Sampler : Rowan Faint

Site : ---Quote number : EN/000

No. of samples analysed : 9

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

· 22-Jul-2024

Accreditation No. 825

Accredited for compliance with ISO/IEC 17025 - Testing

This Quality Control Report contains the following information:

: 9

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

No. of samples received

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories Position Accreditation Category

Ankit Joshi Senior Chemist - Inorganics Sydney Inorganics, Smithfield, NSW Edwardy Fadjar Organic Coordinator Sydney Organics, Smithfield, NSW

Page : 2 of 8 · ES2423038 Work Order

· SENVERSA PTY LTD Client **Project** · S20102 Wetherill Park WME



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot Key:

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit: Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: WATER | | | | | | Laboratory I | Duplicate (DUP) Report | | |
|----------------------|-----------------------|------------------------|------------|--------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG005(ED093)F: Dis | ssolved Metals by ICP | -AES (QC Lot: 5927824) | | | | | | | |
| ES2423038-002 | MW2 | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 3.28 | 3.30 | 0.7 | 0% - 20% |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | 2.30 | 2.27 | 1.6 | 0% - 20% |
| ES2423171-004 | Anonymous | EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | 0.29 | 0.31 | 5.7 | 0% - 20% |
| | | EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EG020F: Dissolved | Metals by ICP-MS (Q | C Lot: 5927822) | | | | | | | |
| ES2422571-003 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | <0.005 | 0.0 | No Limit |
| ES2423171-007 | Anonymous | EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | 0.0001 | 0.0001 | 0.0 | No Limit |
| | | EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | 0.006 | 0.006 | 0.0 | No Limit |
| | | EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | 0.010 | 0.010 | 0.0 | 0% - 50% |
| | | EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | 0.006 | 0.007 | 0.0 | No Limit |
| | | EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | <0.001 | 0.0 | No Limit |
| | | EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | 0.009 | 0.009 | 0.0 | No Limit |
| | | EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | 0.352 | 0.348 | 1.0 | 0% - 20% |

Page : 3 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | | Laboratory E | Ouplicate (DUP) Report | | |
|----------------------|------------------------------|--------------------------------------|------------|--------------|------|-----------------|------------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EG035F: Dissolved I | Mercury by FIMS (QC Lot: 5 | 927821) - continued | | | | | | | |
| ES2422571-004 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| ES2423038-006 | QC104 | EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | <0.0001 | 0.0 | No Limit |
| EK055G: Ammonia a | as N by Discrete Analyser (| QC Lot: 5931487) | | | | | | | |
| EN2406968-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.68 | 0.65 | 3.2 | 0% - 20% |
| ES2423149-002 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | 0.04 | 0.03 | 0.0 | No Limit |
| EK057G: Nitrite as I | N by Discrete Analyser (QC | Lot: 5922064) | | | | | | | |
| ES2422894-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| ES2423027-007 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | <0.01 | 0.0 | No Limit |
| EK059G: Nitrite plus | s Nitrate as N (NOx) by Disc | crete Analyser (QC Lot: 5931486) | | | | | | | |
| EN2406976-006 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 19.2 | 19.1 | 0.5 | 0% - 20% |
| EN2406968-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 4.30 | 4.20 | 2.3 | 0% - 20% |
| EK059G: Nitrite plus | s Nitrate as N (NOx) by Disc | crete Analyser (QC Lot: 5931488) | | | | | | | |
| ES2423149-002 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.73 | 0.73 | 0.0 | 0% - 20% |
| ME2401138-003 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | 0.34 | 0.36 | 5.2 | 0% - 20% |
| EK061G: Total Kjeld | ahl Nitrogen By Discrete An | alyser (QC Lot: 5931483) | | | | | | | |
| EN2406976-007 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (0.5)* | mg/L | 0.8 | 0.9 | 12.1 | No Limit |
| EN2406950-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (2.0)* | mg/L | 101 | 110 | 8.2 | 0% - 20% |
| EK061G: Total Kield | ahl Nitrogen By Discrete An | | | | | | | | |
| ES2423038-005 | MW6 | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | 0.3 | 0.3 | 0.0 | No Limit |
| ES2423536-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 0.1 (1.0)* | mg/L | 50.4 | 49.4 | 1.9 | 0% - 20% |
| EK067G: Total Phos | phorus as P by Discrete An | alvser (QC Lot: 5931482) | | | | | | | |
| EN2406950-001 | Anonymous | EK067G: Total Phosphorus as P | | 0.01 (0.20)* | mg/L | 0.81 | 1.13 | 32.5 | No Limit |
| ES2423038-005 | MW6 | EK067G: Total Phosphorus as P | | 0.01 | mg/L | 0.05 | 0.05 | 0.0 | No Limit |
| EP080/071: Total Pe | troleum Hydrocarbons (QC | Lot: 5924176) | | | | | | | |
| ES2423038-001 | MW1 | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2423038-008 | QC405 | EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080/071: Total Re | coverable Hydrocarbons - N | EPM 2013 Fractions (QC Lot: 5924176) | | | | | | | |
| ES2423038-001 | MW1 | EP080: C6 - C10 Fraction | C6 C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| ES2423038-008 | QC405 | EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | <20 | 0.0 | No Limit |
| EP080: BTEXN (QC | Lot: 5924176) | | | | | | | | |
| ES2423038-001 | MW1 | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |
| | | | 106-42-3 | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit |

Page : 4 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | | Laboratory Duplicate (DUP) Report | | | | | | | |
|----------------------|---------------------------|----------------------------|------------|-----|-----------------------------------|-----------------|------------------|---------|--------------------|--|--|--|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) | | | |
| EP080: BTEXN (QC | Lot: 5924176) - continued | | | | | | | | | | | |
| ES2423038-001 | MW1 | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit | | | |
| ES2423038-008 | QC405 | EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | <1 | 0.0 | No Limit | | | |
| | | EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | | | |
| | | EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | | | |
| | | EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | | | |
| | | | 106-42-3 | | | | | | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | <2 | 0.0 | No Limit | | | |
| | | EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | <5 | 0.0 | No Limit | | | |

Page : 5 of 8 Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | | |
|--|----------------|--------|------|-------------------|---------------|------------------------------|------------|--------------|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | e Limits (%) | |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EG005(ED093)F: Dissolved Metals by ICP-AES (QCLot: 592 | | | | | | | | | |
| EG005F: Iron | 7439-89-6 | 0.05 | mg/L | <0.05 | 0.5 mg/L | 109 | 82.0 | 114 | |
| EG005F: Manganese | 7439-96-5 | 0.01 | mg/L | <0.01 | 0.1 mg/L | 103 | 81.0 | 113 | |
| EG020F: Dissolved Metals by ICP-MS (QCLot: 5927822) | | | | | | | | | |
| EG020A-F: Arsenic | 7440-38-2 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 99.3 | 85.0 | 114 | |
| EG020A-F: Cadmium | 7440-43-9 | 0.0001 | mg/L | <0.0001 | 0.1 mg/L | 97.4 | 84.0 | 110 | |
| EG020A-F: Chromium | 7440-47-3 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.1 | 85.0 | 111 | |
| EG020A-F: Copper | 7440-50-8 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.9 | 81.0 | 111 | |
| EG020A-F: Lead | 7439-92-1 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 93.7 | 83.0 | 111 | |
| EG020A-F: Nickel | 7440-02-0 | 0.001 | mg/L | <0.001 | 0.1 mg/L | 96.3 | 82.0 | 112 | |
| EG020A-F: Zinc | 7440-66-6 | 0.005 | mg/L | <0.005 | 0.1 mg/L | 94.8 | 81.0 | 117 | |
| EG035F: Dissolved Mercury by FIMS (QCLot: 5927821) | | | | | | | | | |
| EG035F: Mercury | 7439-97-6 | 0.0001 | mg/L | <0.0001 | 0.01 mg/L | 93.6 | 83.0 | 105 | |
| EK055G: Ammonia as N by Discrete Analyser (QCLot: 593° | 1487) | | | | | | | | |
| EK055G: Ammonia as N | 7664-41-7 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 95.4 | 90.0 | 114 | |
| EK057G: Nitrite as N by Discrete Analyser (QCLot: 592206 | 4) | | | | | | | | |
| EK057G: Nitrite as N | 14797-65-0 | 0.01 | mg/L | <0.01 | 0.5 mg/L | 100 | 82.0 | 114 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analys | ser (QCLot: 59 | 31486) | | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 108 | 91.0 | 113 | |
| EK059G: Nitrite plus Nitrate as N (NOx) by Discrete Analys | ser (QCLot: 59 | 31488) | | | | | | | |
| EK059G: Nitrite + Nitrate as N | | 0.01 | mg/L | <0.01 | 0.5 mg/L | 107 | 91.0 | 113 | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser(QC | Lot: 5931483) | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 88.7 | 69.0 | 123 | |
| | | | | <0.1 | 1 mg/L | 116 | 70.0 | 123 | |
| | | | | <0.1 | 5 mg/L | 112 | 70.0 | 123 | |
| EK061G: Total Kjeldahl Nitrogen By Discrete Analyser(QC | Lot: 5931484) | | | | | | | | |
| EK061G: Total Kjeldahl Nitrogen as N | | 0.1 | mg/L | <0.1 | 10 mg/L | 87.1 | 69.0 | 123 | |
| | | | | <0.1 | 1 mg/L | 112 | 70.0 | 123 | |
| | | | | <0.1 | 5 mg/L | 108 | 70.0 | 123 | |
| EK067G: Total Phosphorus as P by Discrete Analyser (QCI | Lot: 5931482) | | | | | | | | |

Page : 6 of 8
Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Method Blank (MB) | | Laboratory Control Spike (LC | S) Report | |
|--|--------------------|---------------|------|-------------------|---------------|------------------------------|------------|------------|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) |
| Method: Compound CAS | S Number | LOR | Unit | Result | Concentration | LCS | Low | High |
| EK067G: Total Phosphorus as P by Discrete Analyser (QCLot: 5 | 5931482) - | - continued | | | | | | |
| EK067G: Total Phosphorus as P | | 0.01 | mg/L | <0.01 | 4.42 mg/L | 104 | 71.3 | 126 |
| | | | | <0.01 | 0.442 mg/L | 98.2 | 71.3 | 126 |
| | | | | <0.01 | 1 mg/L | 101 | 70.0 | 130 |
| EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLot: 592 | 3133) | | | | | | | |
| EP075(SIM): Naphthalene | 91-20-3 | 1 | μg/L | <1.0 | 5 μg/L | 68.8 | 50.0 | 94.0 |
| EP075(SIM): Acenaphthylene 2 | 08-96-8 | 1 | μg/L | <1.0 | 5 μg/L | 71.8 | 63.6 | 114 |
| EP075(SIM): Acenaphthene | 83-32-9 | 1 | μg/L | <1.0 | 5 μg/L | 71.5 | 62.2 | 113 |
| EP075(SIM): Fluorene | 86-73-7 | 1 | μg/L | <1.0 | 5 μg/L | 68.4 | 63.9 | 115 |
| EP075(SIM): Phenanthrene | 85-01-8 | 1 | μg/L | <1.0 | 5 μg/L | 75.7 | 62.6 | 116 |
| EP075(SIM): Anthracene 1 | 20-12-7 | 1 | μg/L | <1.0 | 5 μg/L | 78.6 | 64.3 | 116 |
| EP075(SIM): Fluoranthene 2 | 06-44-0 | 1 | μg/L | <1.0 | 5 μg/L | 84.5 | 63.6 | 118 |
| EP075(SIM): Pyrene 1 | 29-00-0 | 1 | μg/L | <1.0 | 5 μg/L | 87.7 | 63.1 | 118 |
| EP075(SIM): Benz(a)anthracene | 56-55-3 | 1 | μg/L | <1.0 | 5 μg/L | 80.9 | 64.1 | 117 |
| EP075(SIM): Chrysene 2 | 18-01-9 | 1 | μg/L | <1.0 | 5 μg/L | 92.3 | 62.5 | 116 |
| . , | 05-99-2 05-82-3 | 1 | μg/L | <1.0 | 5 μg/L | 86.5 | 61.7 | 119 |
| EP075(SIM): Benzo(k)fluoranthene 2 | 07-08-9 | 1 | μg/L | <1.0 | 5 μg/L | 85.9 | 63.0 | 115 |
| EP075(SIM): Benzo(a)pyrene | 50-32-8 | 0.5 | μg/L | <0.5 | 5 μg/L | 85.5 | 63.3 | 117 |
| EP075(SIM): Indeno(1.2.3.cd)pyrene | 93-39-5 | 1 | μg/L | <1.0 | 5 μg/L | 73.2 | 59.9 | 118 |
| EP075(SIM): Dibenz(a.h)anthracene | 53-70-3 | 1 | μg/L | <1.0 | 5 μg/L | 74.8 | 61.2 | 117 |
| EP075(SIM): Benzo(g.h.i)perylene 1 | 91-24-2 | 1 | μg/L | <1.0 | 5 μg/L | 77.8 | 59.1 | 118 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5923134) | | | | | | | | |
| EP071: C10 - C14 Fraction | | 50 | μg/L | <50 | 400 μg/L | 85.8 | 53.7 | 97.0 |
| EP071: C15 - C28 Fraction | | 100 | μg/L | <100 | 600 μg/L | 83.2 | 63.3 | 107 |
| EP071: C29 - C36 Fraction | | 50 | μg/L | <50 | 400 μg/L | 97.1 | 58.3 | 120 |
| EP080/071: Total Petroleum Hydrocarbons (QCLot: 5924176) | | | | | | | | |
| EP080: C6 - C9 Fraction | | 20 | μg/L | <20 | 260 μg/L | 88.1 | 75.0 | 127 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fracti | ons (QCL | .ot: 5923134) | | | | | | |
| EP071: >C10 - C16 Fraction | | 100 | μg/L | <100 | 500 μg/L | 73.3 | 53.9 | 95.5 |
| EP071: >C16 - C34 Fraction | | 100 | μg/L | <100 | 700 μg/L | 82.1 | 57.8 | 110 |
| EP071: >C34 - C40 Fraction | | 100 | μg/L | <100 | 300 μg/L | 105 | 50.5 | 115 |
| EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fracti | ons (QCL | .ot: 5924176) | | | | | | |
| EP080: C6 - C10 Fraction | C6_C10 | 20 | μg/L | <20 | 310 μg/L | 82.9 | 75.0 | 127 |
| EP080: BTEXN (QCLot: 5924176) | | | | | | | | |

Page : 7 of 8 Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | Method Blank (MB) | Laboratory Control Spike (LCS) Report | | | | | | | |
|---|-------------------|---------------------------------------|------|--------|---------------|--------------------|------------|------------|--|
| | | | | Report | Spike | Spike Recovery (%) | Acceptable | Limits (%) | |
| Method: Compound | CAS Number | LOR | Unit | Result | Concentration | LCS | Low | High | |
| EP080: BTEXN (QCLot: 5924176) - continued | | | | | | | | | |
| EP080: Benzene | 71-43-2 | 1 | μg/L | <1 | 10 μg/L | 84.9 | 68.3 | 119 | |
| EP080: Toluene | 108-88-3 | 2 | μg/L | <2 | 10 μg/L | 95.7 | 73.5 | 120 | |
| EP080: Ethylbenzene | 100-41-4 | 2 | μg/L | <2 | 10 μg/L | 89.7 | 73.8 | 122 | |
| EP080: meta- & para-Xylene | 108-38-3 | 2 | μg/L | <2 | 10 μg/L | 97.5 | 73.0 | 122 | |
| | 106-42-3 | | | | | | | | |
| EP080: ortho-Xylene | 95-47-6 | 2 | μg/L | <2 | 10 μg/L | 96.1 | 76.4 | 123 | |
| EP080: Naphthalene | 91-20-3 | 5 | μg/L | <5 | 10 μg/L | 103 | 75.5 | 124 | |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

| Sub-Matrix: WATER | | | | Ma | atrix Spike (MS) Report | • | |
|----------------------|--|--------------------------------|------------|---------------|-------------------------|--------------|-----------|
| | | | | Spike | SpikeRecovery(%) | Acceptable i | imits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EG005(ED093)F: D | issolved Metals by ICP-AES (QCLot: 5927824) | | | | | | |
| ES2423038-001 | MW1 | EG005F: Manganese | 7439-96-5 | 1 mg/L | # Not Determined | 70.0 | 130 |
| EG020F: Dissolved | Metals by ICP-MS (QCLot: 5927822) | | | | | | |
| ES2422571-002 | Anonymous | EG020A-F: Arsenic | 7440-38-2 | 1 mg/L | 96.6 | 70.0 | 130 |
| | | EG020A-F: Cadmium | 7440-43-9 | 0.25 mg/L | 98.8 | 70.0 | 130 |
| | | EG020A-F: Chromium | 7440-47-3 | 1 mg/L | 95.1 | 70.0 | 130 |
| | | EG020A-F: Copper | 7440-50-8 | 1 mg/L | 96.7 | 70.0 | 130 |
| | | EG020A-F: Lead | 7439-92-1 | 1 mg/L | 92.5 | 70.0 | 130 |
| | | EG020A-F: Nickel | 7440-02-0 | 1 mg/L | 97.7 | 70.0 | 130 |
| | | EG020A-F: Zinc | 7440-66-6 | 1 mg/L | 99.2 | 70.0 | 130 |
| EG035F: Dissolved | Mercury by FIMS (QCLot: 5927821) | | | | | | |
| ES2422571-001 | Anonymous | EG035F: Mercury | 7439-97-6 | 0.01 mg/L | 84.1 | 70.0 | 130 |
| EK055G: Ammonia | as N by Discrete Analyser (QCLot: 5931487) | | | | | | |
| EN2406968-001 | Anonymous | EK055G: Ammonia as N | 7664-41-7 | 0.5 mg/L | 113 | 70.0 | 130 |
| EK057G: Nitrite as | N by Discrete Analyser (QCLot: 5922064) | | | | | | |
| ES2422894-001 | Anonymous | EK057G: Nitrite as N | 14797-65-0 | 0.5 mg/L | 101 | 70.0 | 130 |
| EK059G: Nitrite pl | us Nitrate as N (NOx) by Discrete Analyser (QCLot: | 5931486) | | | | | |
| EN2406968-001 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | # Not Determined | 70.0 | 130 |
| EK059G: Nitrite pl | us Nitrate as N (NOx) by Discrete Analyser (QCLot: | 5931488) | | | | | |

Page : 8 of 8 Work Order : ES2423038

Client : SENVERSA PTY LTD
Project : S20102 Wetherill Park WME



| Sub-Matrix: WATER | | | | Ma | trix Spike (MS) Repor | t | |
|----------------------|--|--------------------------------------|------------|---------------|-----------------------|--------------|------------|
| | | | | Spike | SpikeRecovery(%) | Acceptable I | Limits (%) |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EK059G: Nitrite pl | us Nitrate as N (NOx) by Discrete Analyser (QCLot: 593 | 1488) - continued | | | | | |
| ME2401138-002 | Anonymous | EK059G: Nitrite + Nitrate as N | | 0.5 mg/L | 125 | 70.0 | 130 |
| EK061G: Total Kjel | dahl Nitrogen By Discrete Analyser (QCLot: 5931483) | | | | | | |
| EN2406950-002 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 100 mg/L | 99.8 | 70.0 | 130 |
| EK061G: Total Kjel | dahl Nitrogen By Discrete Analyser (QCLot: 5931484) | | | | | | |
| ES2423536-001 | Anonymous | EK061G: Total Kjeldahl Nitrogen as N | | 50 mg/L | 104 | 70.0 | 130 |
| EK067G: Total Pho | sphorus as P by Discrete Analyser (QCLot: 5931482) | | | | | | |
| EN2406950-002 | Anonymous | EK067G: Total Phosphorus as P | | 1 mg/L | 97.0 | 70.0 | 130 |
| EP080/071: Total P | etroleum Hydrocarbons (QCLot: 5924176) | | | | | | |
| ES2423038-001 | MW1 | EP080: C6 - C9 Fraction | | 325 μg/L | 108 | 70.0 | 130 |
| EP080/071: Total R | ecoverable Hydrocarbons - NEPM 2013 Fractions (QCL | ot: 5924176) | | | | | |
| ES2423038-001 | MW1 | EP080: C6 - C10 Fraction | C6_C10 | 375 μg/L | 100 | 70.0 | 130 |
| EP080: BTEXN (Q | CLot: 5924176) | | | | | | |
| ES2423038-001 | MW1 | EP080: Benzene | 71-43-2 | 25 μg/L | 76.5 | 70.0 | 130 |
| | | EP080: Toluene | 108-88-3 | 25 μg/L | 79.4 | 70.0 | 130 |
| | | EP080: Ethylbenzene | 100-41-4 | 25 μg/L | 81.6 | 70.0 | 130 |
| | | EP080: meta- & para-Xylene | 108-38-3 | 25 μg/L | 78.9 | 70.0 | 130 |
| | | | 106-42-3 | | | | |
| | | EP080: ortho-Xylene | 95-47-6 | 25 μg/L | 92.7 | 70.0 | 130 |
| | | EP080: Naphthalene | 91-20-3 | 25 μg/L | 97.4 | 70.0 | 130 |

, ev rsa

Chain of Custody Documentation

| | Pty Ltd | | | Laboratory: | ALS NSW | | | | | | | | / | \nalysis | Require | ed | | |
|-------------------------------|------------------------------|---------------------|--------------------------------------|--------------------------------------|----------------------------|-------------------|--|------------------------------|--|---------------------------------------|-------------------|------------------|--------------|-----------|---------|----|-------|---|
| WWW.SORVE ABN 89 13 | | | | Address: Contact: Phone: | Sample Receipt | | | | | | | | | | | | | <u>Comments</u> : e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. |
| Job Numb | er; | S2 | 20102 | Purchase Order: | | | 1 | TALS | TALS | Ş | | | | | | | | Environmental Divisi |
| Project Na: | me: | Wetherill | Park WME | Quote No: | EN/103/21 | | | 8 ME | 8 ME | NS A | | | | · · | | | i | Sydney Work Order Reference ES230401 |
| Sampled B | у: | Bec (| Chapple | Turn Around Time: | Standard 7 |)ays | 9 | PAH | PAH | ANIC | } | | | NN NN | | | | Work Order Reference |
| Project Ma | nager: | | a Walsh | Page: | 1 | of 1 . | Ĭ. | JEX/ | ĪÄ | SNC | a. |) _@ | | AND MN) | İ | | | E3230401 |
| Email Repo | ort To: | | senversa.com.au. Psenversa.com.au | Phone/Mobile: | 0408038593, 040 | 4011544 | M-18 (TRH/BTEXN) | N-26 (TRH/BTEX/PAH/8 METALS) | W:27 (TRH/BTEX/PAH/8 METALS/ PHENOLS) | NT-14 (CATIONS, ANIONS AND NUTRIENTS) | NT-11 (TN, TP | EA015H (TDS) | EA025H (TSS) | 뿐 | | | | 2 111 2 11 7 75.2. 11 17 |
| _ | | Sample Information | on | | Container Infor | mation | 8 | ၂ | | 4 E |] E . | <u> 후</u> | 1251 | <u>ĝ</u> | | | ا ا | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | 1 ₹ | M-2 | % E | Εğ | 🛓 | I % | EA0 | EG005F (| | i | НОГЪ | |
| - 1 | QC401 | w | 8/02/2023 | АМ | VOA | 1 | х | | - | | <u> </u> | | | | | | | |
| 2 | QC501 | w | 8/02/2023 | AM | VOA | 1 | х | | | | | | | | | | | |
| 3 | QC301 | w | 8/02/2023 | AM | VS x2, N, UA, VSA | 5 | | х | | | х | | - | х | | | | Telephone: + 61-2-6784 6555 |
| <u>'</u> | MW1 | W | 8/02/2023 | AM | P; VS x2. N. UA. VSA | 6 | | | х | х | | | | x i | | | | 1 |
| ₹ | MW2 | w | 8/02/2023 | АМ | P, VS x2, N, UA, VSA | l 6 | | | х | х | | | | х | | | | |
| 6 | MW3 | W | 8/02/2023 | AM | P, VS x2, N, UA, VSA | 6 | | | X | х | | | | х | *. | | | |
| 7 | MW4 | w | 8/02/2023 | AM | P. VS x2, N, UA, VSA | 6 | - | | X. | х | | | | х | | | | |
| 8 | MW6 | w | 8/02/2023 | АМ | P. VS x2, N, UA, VSA | 6 | | | х | Χ̈ | _ | | | х | | | | |
| 9 | QC101 | W | 8/02/2023 | АМ | VS x2, N. UA. VSA | 5. | | х | | | х | | | x | | | | |
| X_ | QC201 | w | 8/02/2023 | AM | VS x2, N, UA, VSA | _ 5 | | | | Envir | olab Se | | | | | | | Please forward to Envirolab |
| | | | | | | | _FI | LOSTON | ÀB_ | | 12 Asi | ley St | | | | | | , |
| | | | | | | , | | _ | , , | natswo | od NSI n2) 991 | 0 6200 | | | | | | |
| | | | | | | | | h No | - 2 | 11 15 | -9 | | | | | | | |
| | Subcon | Corward Lab | / Split WO | | | <u>'</u> | | | | \\ \(\) | , , | 15 | ,] | | | | | · · · · · · · · · · · · · · · · · · · |
| | | l to | (A) | 1211 | WRUAB | | D | ate Re | ceived | Y. | 10,4 | 10 | ٧ | , | | | | Event I |
| | Lab / A | 7 7 | | | | | T | me Re | ceive | <u> ,(v</u> | 00 | | | | | | | |
| | Organis | ed By / Date: | | | - | | R | eceive | g BA | UST | , | | | | | | | 4. 47 W |
| | Relinqu | ished By / Dat | 31 | | | | | empro | 12 | epack | | 7 | | | | | | |
| Γotal | - Contot | e / Courier: | | | | 47 🕳 | - | ecutit | | | ep/No | ie) | | | | | | |
| Sampler: l a | ttest that proper field samp | ing procedures in a | ccordance with Se | nversa standard proce | dures and/or project | Sampler Name: | | Bec C | happle | | Signatu | re: | AL, | 版 |) _ | | Date: | 8/02/2023 |
| Relinguishe | | d By PO / Inte | 1.65 | | Method of Shipment (if app | lianhlair | | | Receive | | | 2 | <u> </u> | <u>**</u> | / | | | |
| Vame/Signa | | · Bec Chapple | - Persey/ | Date: 8/2/23 | Carrier / Reference #: | mca <u>ble</u> j. | | | | ignature | | 7-79 | | 1 | | | _ | Date: SIUI3 |
| Of: | | | | Time: 12:00 PM | Date/Time: | - | | | Of: | ng natur C | • | / π Δ | 1 | 17 | | | | Time: 12-29_ |
| lame/Signa | ure: | | | Date: | Carrier / Reference #: | | | | Name/S | ignature | <u>Ci</u> | 1/-1/ | Fix | | | 1_ | | Date: 09 (02/73 |
| Of: | | | | Time: | Date/Time: | | <u>. </u> | | Of: | | 1 / | <u> 27</u> | 0 | | | | | Time: 1,6(1(1) |
| lame/Signal Of: | ure: | | | Date: | Carrier / Reference #: | | | _ | Name/S | ignature | : | | | | | | | Date: |
| | | | _ | Time: eserved Plastic; ORC = Niti | Date/Time: | 4 | | | Of: | | | | | | | | | Time: |

Simon Song

From:

Emma Walsh < Emma. Walsh@senversa.com.au>

Sent:

Wednesday, 15 February 2023 9:55 AM

To:

Simon Song

Cc:

Bec Chapple

Subject:

RE: Sample Receipt for 316159 S20102, Wetherill Park WME

CAUTION: This email originated from outside of the organisation. Do not act on instructions, click links or open attachments unless you recognise the sender and know the content is authentic and safe.

Hi Simon,

Sorry for the delay in getting back to you - can you pleased analyse sample QC201 for the following:

- TRH/BTEXN/PAH/8 metals (As, Cd, Cr, Cu, Hg, Ni and Zn)
- Total N, total P
- Additional metals iron and manganese

APPENDED THE PROPERTY OF A STATE OF THE STAT

10.00

Thanks.

Kind regards,

senversa

Emma Walsh

Senior Associate Environmental Scientist

M: +61 404 011 544 www.senversa.com.au

Level 24, 1 Market St, Djubuguli, Eora Country Sydney, NSW, 2000, Australia etnerill Pa. 4N:

tructons, c. Make o. . .

Octob for the follow

From: Simon Song <SSong@envirolab.com.au>

Sent: Friday, 10 February 2023 1:16 PM

To: Bec Chapple <bec.chapple@senversa.com.au>; Emma Walsh <Emma.Walsh@senversa.com.au>

Subject: Sample Receipt for 316159 S20102, Wetherill Park WME

Please refer to attached for:

a copy of the COC/paperwork received from you

a copy of our Sample Receipt Advice (SRA)

Please open and read the SRA as it contains important information.

Please let the lab know immediately if there are any issues.

echemi Pa AV

Results will be available by 6.30pm on the date indicated.

PLEASE NOTE COMBO PRICES WILL ONLY APPLY IF COMBOS ARE SELECTED ON COC. Company of the property of the combos are selected on coc.

We have a new reporting format and would welcome your feedback. Sydney@envirolab.com.au

Please note that subcontracted testing or non routine testing may take significantly longer than just the standard 5 day TAT, contact the lab to get an approximate due date.

Enquiries should be made directly to: customerservice@envirolab.com.au

DOLDLIGHT IN FOR GA

Regards

Envirolab Services
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

The Property State of the

5. . .

Mari Par A

in Emiles in the in-



Envirolab Services Pty Ltd
ABN 37 112 535 645
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

SAMPLE RECEIPT ADVICE

| Client Details | |
|----------------|-------------------------|
| Client | Senversa Pty Ltd |
| Attention | Bec Chapple, Emma Walsh |

| Sample Login Details | |
|--------------------------------------|----------------------------|
| Your reference | S20102, Wetherill Park WME |
| Envirolab Reference | 316159 |
| Date Sample Received | 09/02/2023 |
| Date Instructions Received | 15/02/2023 |
| Date Results Expected to be Reported | 22/02/2023 |

| Sample Condition | |
|--|----------|
| Samples received in appropriate condition for analysis | Yes |
| No. of Samples Provided | 1 Water |
| Turnaround Time Requested | Standard |
| Temperature on Receipt (°C) | 3 |
| Cooling Method | Ice |
| Sampling Date Provided | YES |

| Comments |
|--|
| last day of holding time for organics 15/2 |

Please direct any queries to:

| Aileen Hie | Jacinta Hurst | |
|------------------------------|--------------------------------|--|
| Phone: 02 9910 6200 | Phone: 02 9910 6200 | |
| Fax: 02 9910 6201 | Fax: 02 9910 6201 | |
| Email: ahie@envirolab.com.au | Email: jhurst@envirolab.com.au | |

Analysis Underway, details on the following page:



Envirolab Services Pty Ltd
ABN 37 112 535 645
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

| Sample ID | vTRH(C6-C10)/BTEXN in Water | svTRH (C10-C40) in Water | PAHsin Water | HM in water - dissolved | Total Nitrogen in water | Metals in Waters -Total |
|-----------|-----------------------------|--------------------------|--------------|-------------------------|-------------------------|-------------------------|
| QC201 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ |

The '\sqrt{'} indicates the testing you have requested. THIS IS NOT A REPORT OF THE RESULTS.

Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.



Envirolab Services Pty Ltd

ABN 37 112 535 645 12 Ashley St Chatswood NSW 2067 ph 02 9910 6200 fax 02 9910 6201 customerservice@envirolab.com.au www.envirolab.com.au

CERTIFICATE OF ANALYSIS 316159

| Client Details | |
|----------------|---------------------------------------|
| Client | Senversa Pty Ltd |
| Attention | Bec Chapple, Emma Walsh |
| Address | 6/15 William St, Melbourne, VIC, 3000 |

| Sample Details | |
|--------------------------------------|----------------------------|
| Your Reference | S20102, Wetherill Park WME |
| Number of Samples | 1 Water |
| Date samples received | 09/02/2023 |
| Date completed instructions received | 15/02/2023 |

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Please refer to the last page of this report for any comments relating to the results.

| Report Details | | |
|--|------------|--|
| Date results requested by | 22/02/2023 | |
| Date of Issue | 22/02/2023 | |
| NATA Accreditation Number 2901. This document shall not be reproduced except in full. | | |
| Accredited for compliance with ISO/IEC 17025 - Testing. Tests not covered by NATA are denoted with * | | |

Results Approved By

Diego Bigolin, Inorganics Supervisor Hannah Nguyen, Metals Supervisor Josh Williams, Organics Supervisor Kyle Gavrily, Senior Chemist Authorised By

Nancy Zhang, Laboratory Manager



| vTRH(C6-C10)/BTEXN in Water | | |
|---|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date extracted | - | 21/02/2023 |
| Date analysed | - | 21/02/2023 |
| TRH C ₆ - C ₉ | μg/L | <10 |
| TRH C ₆ - C ₁₀ | μg/L | <10 |
| TRH C ₆ - C ₁₀ less BTEX (F1) | μg/L | <10 |
| Benzene | μg/L | <1 |
| Toluene | μg/L | <1 |
| Ethylbenzene | μg/L | <1 |
| m+p-xylene | μg/L | <2 |
| o-xylene | μg/L | <1 |
| Naphthalene | μg/L | <1 |
| Surrogate Dibromofluoromethane | % | 113 |
| Surrogate toluene-d8 | % | 103 |
| Surrogate 4-BFB | % | 104 |

| svTRH (C10-C40) in Water | | |
|--|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date extracted | - | 16/02/2023 |
| Date analysed | - | 16/02/2023 |
| TRH C ₁₀ - C ₁₄ | μg/L | <50 |
| TRH C ₁₅ - C ₂₈ | μg/L | 140 |
| TRH C ₂₉ - C ₃₆ | μg/L | <100 |
| Total +ve TRH (C10-C36) | μg/L | 140 |
| TRH >C ₁₀ - C ₁₆ | μg/L | 130 |
| TRH >C ₁₀ - C ₁₆ less Naphthalene (F2) | μg/L | 130 |
| TRH >C ₁₆ - C ₃₄ | μg/L | <100 |
| TRH >C ₃₄ - C ₄₀ | μg/L | <100 |
| Total +ve TRH (>C10-C40) | μg/L | 130 |
| Surrogate o-Terphenyl | % | 67 |

| PAHs in Water | | |
|---------------------------|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date extracted | - | 16/02/2023 |
| Date analysed | - | 20/02/2023 |
| Naphthalene | μg/L | <2 |
| Acenaphthylene | μg/L | <1 |
| Acenaphthene | μg/L | <1 |
| Fluorene | μg/L | <1 |
| Phenanthrene | μg/L | <1 |
| Anthracene | μg/L | <1 |
| Fluoranthene | μg/L | <1 |
| Pyrene | μg/L | <1 |
| Benzo(a)anthracene | μg/L | <1 |
| Chrysene | μg/L | <1 |
| Benzo(b,j+k)fluoranthene | μg/L | <2 |
| Benzo(a)pyrene | μg/L | <1 |
| Indeno(1,2,3-c,d)pyrene | μg/L | <1 |
| Dibenzo(a,h)anthracene | μg/L | <1 |
| Benzo(g,h,i)perylene | μg/L | <1 |
| Benzo(a)pyrene TEQ | μg/L | <5 |
| Total +ve PAH's | μg/L | NIL (+)VE |
| Surrogate p-Terphenyl-d14 | % | 74 |

Envirolab Reference: 316159

Revision No: R00

| HM in water - dissolved | | |
|-------------------------|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date prepared | - | 17/02/2023 |
| Date analysed | - | 20/02/2023 |
| Arsenic-Dissolved | μg/L | 4 |
| Cadmium-Dissolved | μg/L | 0.1 |
| Chromium-Dissolved | μg/L | 2 |
| Copper-Dissolved | μg/L | <1 |
| Lead-Dissolved | μg/L | 1 |
| Mercury-Dissolved | μg/L | <0.05 |
| Nickel-Dissolved | μg/L | 180 |
| Zinc-Dissolved | μg/L | 230 |
| Iron-Dissolved | μg/L | 5,700 |
| Manganese-Dissolved | μg/L | 5,800 |

| Miscellaneous Inorganics | | |
|--------------------------|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date prepared | - | 16/02/2023 |
| Date analysed | - | 16/02/2023 |
| Total Nitrogen in water | mg/L | 0.5 |

| Metals in Waters - Total | | |
|--------------------------|-------|------------|
| Our Reference | | 316159-1 |
| Your Reference | UNITS | QC201 |
| Date Sampled | | 8/02/2023 |
| Type of sample | | Water |
| Date prepared | - | 20/02/2023 |
| Date analysed | - | 20/02/2023 |
| Phosphorus - Total | mg/L | 0.8 |

| Method ID | Methodology Summary |
|-------------------|--|
| Inorg-055/062/127 | Total Nitrogen - Calculation sum of TKN and oxidised Nitrogen. Alternatively analysed by combustion and chemiluminescence. |
| Metals-020 | Determination of various metals by ICP-AES. |
| Metals-021 | Determination of Mercury by Cold Vapour AAS. |
| Metals-022 | Determination of various metals by ICP-MS. |
| Org-020 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis. |
| Org-022/025 | Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS/GC-MSMS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013. |
| Org-023 | Water samples are analysed directly by purge and trap GC-MS. |
| Org-023 | Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater. |

Envirolab Reference: 316159

Revision No: R00

| QUALITY CONT | ROL: vTRH(| C6-C10)/E | BTEXN in Water | | Duplicate | | | | Spike Recovery % | | |
|--------------------------------------|------------|-----------|----------------|------------|-----------|------|------|------|------------------|------|--|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] | |
| Date extracted | - | | | 21/02/2023 | [NT] | | [NT] | [NT] | 21/02/2023 | | |
| Date analysed | - | | | 21/02/2023 | [NT] | | [NT] | [NT] | 21/02/2023 | | |
| TRH C ₆ - C ₉ | μg/L | 10 | Org-023 | <10 | [NT] | | [NT] | [NT] | 95 | | |
| TRH C ₆ - C ₁₀ | μg/L | 10 | Org-023 | <10 | [NT] | | [NT] | [NT] | 95 | | |
| Benzene | μg/L | 1 | Org-023 | <1 | [NT] | | [NT] | [NT] | 94 | | |
| Toluene | μg/L | 1 | Org-023 | <1 | [NT] | | [NT] | [NT] | 97 | | |
| Ethylbenzene | μg/L | 1 | Org-023 | <1 | [NT] | | [NT] | [NT] | 98 | | |
| m+p-xylene | μg/L | 2 | Org-023 | <2 | [NT] | | [NT] | [NT] | 93 | | |
| o-xylene | μg/L | 1 | Org-023 | <1 | [NT] | | [NT] | [NT] | 96 | | |
| Naphthalene | μg/L | 1 | Org-023 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Surrogate Dibromofluoromethane | % | | Org-023 | 110 | [NT] | | [NT] | [NT] | 97 | | |
| Surrogate toluene-d8 | % | | Org-023 | 104 | [NT] | | [NT] | [NT] | 100 | | |
| Surrogate 4-BFB | % | | Org-023 | 103 | [NT] | | [NT] | [NT] | 101 | | |

Envirolab Reference: 316159

Revision No: R00

| QUALITY CON | QUALITY CONTROL: svTRH (C10-C40) in Water | | | | | | | Duplicate | | |
|--|---|-----|---------|------------|------|------|------|-----------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] |
| Date extracted | - | | | 16/02/2023 | [NT] | | [NT] | [NT] | 16/02/2023 | |
| Date analysed | - | | | 16/02/2023 | [NT] | | [NT] | [NT] | 16/02/2023 | |
| TRH C ₁₀ - C ₁₄ | μg/L | 50 | Org-020 | <50 | [NT] | | [NT] | [NT] | 86 | |
| TRH C ₁₅ - C ₂₈ | μg/L | 100 | Org-020 | <100 | [NT] | | [NT] | [NT] | 120 | |
| TRH C ₂₉ - C ₃₆ | μg/L | 100 | Org-020 | <100 | [NT] | | [NT] | [NT] | 100 | |
| TRH >C ₁₀ - C ₁₆ | μg/L | 50 | Org-020 | <50 | [NT] | | [NT] | [NT] | 86 | |
| TRH >C ₁₆ - C ₃₄ | μg/L | 100 | Org-020 | <100 | [NT] | | [NT] | [NT] | 120 | |
| TRH >C ₃₄ - C ₄₀ | μg/L | 100 | Org-020 | <100 | [NT] | | [NT] | [NT] | 100 | |
| Surrogate o-Terphenyl | % | | Org-020 | 75 | [NT] | | [NT] | [NT] | 82 | |

| QUAL | ITY CONTRO | L: PAHs ir | ı Water | | | Duplicate | | | Spike Recovery % | | |
|---------------------------|------------|------------|-------------|------------|------|-----------|------|------|------------------|------|--|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] | |
| Date extracted | - | | | 16/02/2023 | [NT] | | [NT] | [NT] | 16/02/2023 | | |
| Date analysed | - | | | 20/02/2023 | [NT] | | [NT] | [NT] | 20/02/2023 | | |
| Naphthalene | μg/L | 2 | Org-022/025 | <2 | [NT] | | [NT] | [NT] | 72 | | |
| Acenaphthylene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Acenaphthene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 73 | | |
| Fluorene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 74 | | |
| Phenanthrene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 82 | | |
| Anthracene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Fluoranthene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 80 | | |
| Pyrene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 85 | | |
| Benzo(a)anthracene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Chrysene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 69 | | |
| Benzo(b,j+k)fluoranthene | μg/L | 2 | Org-022/025 | <2 | [NT] | | [NT] | [NT] | [NT] | | |
| Benzo(a)pyrene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | 86 | | |
| Indeno(1,2,3-c,d)pyrene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Dibenzo(a,h)anthracene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Benzo(g,h,i)perylene | μg/L | 1 | Org-022/025 | <1 | [NT] | | [NT] | [NT] | [NT] | | |
| Surrogate p-Terphenyl-d14 | % | | Org-022/025 | 77 | [NT] | | [NT] | [NT] | 77 | | |

| QUALITY CONTROL: HM in water - dissolved | | | | | | | Duplicate | | | Spike Recovery % | |
|--|-------|------|------------|------------|------|------|-----------|------|------------|------------------|--|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] | |
| Date prepared | - | | | 17/02/2023 | [NT] | | [NT] | [NT] | 17/02/2023 | | |
| Date analysed | - | | | 20/02/2023 | [NT] | | [NT] | [NT] | 20/02/2023 | | |
| Arsenic-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 93 | | |
| Cadmium-Dissolved | μg/L | 0.1 | Metals-022 | <0.1 | [NT] | | [NT] | [NT] | 95 | | |
| Chromium-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 94 | | |
| Copper-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 94 | | |
| Lead-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 99 | | |
| Mercury-Dissolved | μg/L | 0.05 | Metals-021 | <0.05 | [NT] | | [NT] | [NT] | 97 | | |
| Nickel-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 94 | | |
| Zinc-Dissolved | μg/L | 1 | Metals-022 | <1 | [NT] | | [NT] | [NT] | 95 | | |
| Iron-Dissolved | μg/L | 10 | Metals-022 | <10 | [NT] | | [NT] | [NT] | 93 | | |
| Manganese-Dissolved | μg/L | 5 | Metals-022 | <5 | [NT] | | [NT] | [NT] | 94 | | |

| QUALITY CONTROL: Miscellaneous Inorganics | | | | | | Duplicate | | | Spike Recovery % | |
|---|-------|-----|-------------------|------------|------|-----------|------|------|------------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] |
| Date prepared | - | | | 16/02/2023 | [NT] | [NT] | | [NT] | 16/02/2023 | |
| Date analysed | - | | | 16/02/2023 | [NT] | [NT] | | [NT] | 16/02/2023 | |
| Total Nitrogen in water | mg/L | 0.1 | Inorg-055/062/127 | <0.1 | [NT] | [NT] | | [NT] | 111 | |

| QUALITY CONTROL: Metals in Waters - Total | | | | | | Duplicate | | | Spike Recovery % | |
|---|-------|------|------------|------------|------|-----------|------|------|------------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] |
| Date prepared | - | | | 20/02/2023 | [NT] | | [NT] | [NT] | 20/02/2023 | |
| Date analysed | - | | | 20/02/2023 | [NT] | | [NT] | [NT] | 20/02/2023 | |
| Phosphorus - Total | mg/L | 0.05 | Metals-020 | <0.05 | [NT] | [NT] | [NT] | [NT] | 111 | |

Envirolab Reference: 316159

Page | **14 of 17** Revision No: R00

| Result Definiti | ons |
|-----------------|---|
| NT | Not tested |
| NA | Test not required |
| INS | Insufficient sample for this test |
| PQL | Practical Quantitation Limit |
| < | Less than |
| > | Greater than |
| RPD | Relative Percent Difference |
| LCS | Laboratory Control Sample |
| NS | Not specified |
| NEPM | National Environmental Protection Measure |
| NR | Not Reported |

Envirolab Reference: 316159

Revision No: R00

| Quality Control | ol Definitions |
|------------------------------------|--|
| Blank | This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples. |
| Duplicate | This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable. |
| Matrix Spike | A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist. |
| LCS (Laboratory Control Sample) | This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample. |
| Surrogate Spike | Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples. |

Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.

The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOH Exposure Standards Committee, 2016.

Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2

Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

Report Comments

Total metals: no unfiltered, preserved sample was received, therefore analysis was conducted from the unpreserved amber sample bottle

Note: there is a possibility some elements may be underestimated.

Envirolab Reference: 316159

Revision No: R00

Page | 17 of 17





Envirolab Services Pty Ltd
ABN 37 112 535 645
12 Ashley St Chatswood NSW 2067
ph 02 9910 6200 fax 02 9910 6201
customerservice@envirolab.com.au
www.envirolab.com.au

DATA QUALITY ASSESSMENT SUMMARY

| Report Details | |
|----------------------------|----------------------------|
| Envirolab Report Reference | <u>316159</u> |
| Client ID | Senversa Pty Ltd |
| Project Reference | S20102, Wetherill Park WME |
| Date Issued | 22/02/2023 |

QC DATA

All laboratory QC data was within the Envirolab Group's specifications.

HOLDING TIME COMPLIANCE EVALUATION

All preservation / holding times (based on AS/ASPHA/ISO/NEPM/USEPA reference documents and standards) are compliant except:

| Holding Time Exceedances | | | | | |
|--------------------------|-----------|--------------|----------------|---------------|----------|
| Analysis | Sample No | Date Sampled | Date Extracted | Date Analysed | Accepted |
| svTRH (C10-C40) in Water | | | | | |
| | 316159-1 | 8/02/2023 | 16/02/2023 | 16/02/2023 | Х |
| PAHs in Water | | | | | |
| | 316159-1 | 8/02/2023 | 16/02/2023 | 20/02/2023 | X |

Certain analyses have had their recommended technical holding times elongated by filtering and/or freezing on receipt at the laboratory (e.g. BOD, chlorophyll/Pheophytin, nutrients and acid sulphate soil tests).

COMPLIANCE TO QC FREQUENCY (NEPM)

Internal laboratory QC rate complies with NEPM requirements (LCS/MB/MS 1 in 20, Duplicates 1 in 10 samples). Note, samples are batched together with other sample consignments in order to assign QC sample frequency.

| QC Evaluation | |
|--|---|
| Duplicate(s) was performed as per NEPM frequency | ✓ |
| Laboratory Control Sample(s) were analysed with the samples received | ✓ |
| A Method Blank was performed with the samples received | ✓ |
| Matrix spike(s) was performed as per NEPM frequency (Not Applicable for Air samples) | ✓ |

Refer to Certificate of Analysis for all Quality Control data.

senversa

Chain of Custody Documentation

| Senversa Pty Ltd www.senversa.com.au | | | Laboratory: | mgt/Eurofins VIC | rofins VIC | | Analysis Required | | | | | | | | | | | |
|--|---|--|-------------------------------------|-----------------------------------|---|--------------------------------------|---------------------------|----------|------------|--------|---------------|-------------|------------|----------|------------|------------|----------|---|
| ABN 89 132 231 | | | | Address: Contact: Phone: | Sample Receipt | | | neg | | | | | | | | | | Comments: e.g. Highly contaminated sam hazardous materials present; trace LORs |
| Job Number: | | SZ | 20102 | Purchase Order: | | | tals | Nitro | | | | | | | | | | |
| Project Name: Redirect Sampled By: HY Project Manager: Bec Chapple | | Quote No: | | | Ş, | Total Phosphorous and Total Nitrogen | | | | | | | | | | | | |
| | | | | Turn Around Time: 24 Hours | | | Heav | Hea | <u>lo</u> | | | | | | | | | |
| | | Bec | | | 1 | | of 1 | | and In | | | | | | | | | |
| | | Page: 1 of 1 Phone/Mobile: 0408 038 593 | | TRH/BTEX/PAH/8 Heavy metals | osbhr | 88 88 | | | | | | | | | | | | |
| | | Sample Informati | | | Container Info | | <u> </u> | <u>₹</u> | Manganese | | 1 | 1 1 | | | | | | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | 崖 | 흅 | | | | | | | | | НОГР | |
| | QC202 | Water | 14/08/2023 | | | | Х | Х | X | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | \neg | \dashv | |
| | | | | | | | | | | | | | | | _ | \neg | \neg | |
| otal | | | | | | | | | | | | | | | | | | |
| ampler: I attest pecifications w | that proper field samp ere used during the col | ling procedures in a lection of these sam | ccordance with Sen | versa standard proced | lures and/or project | Sampler Name: | | layley Y | 'ellowlees | | Signatu | re: | | | | Da | ite: | 14/08/20: |
| Relinquished By | : | | | | Method of Shipment (if app | ilcable): | | | Receive | d by: | - | | | | | | | |
| | | | Date: 25/8/23 | Carrier / Reference #: | | | Name/Signature: Phenounce | | | | | Date: 15/8 | | | | | | |
| | | | Time: | Date/Time: | | | Of: Comm | | | | Т | Ime: {L: {L | | | | | | |
| | | | Date: Time: | Carrier / Reference #: | | | Name/Signature: Date: | | | | | | | | | | | |
| 700 | | | Date: | Date/Time: | | | Of: | | | | $\overline{}$ | îme; | | | | | | |
| a. | | | Time: | Carrier / Reference #: Date/Time: | | | | | | | | Pate: | | | | | | |
| Water | Container Codes: P = U | Inpreserved Plastic; N = | Nitric Acid (HNO ₃) Pre | served Plastic: ORC = Nit | ric Preserved ORC: SH = Sodium | Hydroxide (NaOH)/Ca | admium (| Cd) Pres | anrad: S - | Sodium | 1 Hydroxide | Preserved | l Plastic; | STH = Sc | dium thios | ulfate pre | served | îme: plastic; |
| V - VL | JA VIXII FIVOOCIIIONG ACIO (M | un Preserved: VS = VC | JA VIAI SUIDHURG Presei | vod: VSA = Subhburic Pro | served Amber Glass; H = HCl Pro = Sterile Bottle; UA = Unpreserved | control Pitantin, 110 - | 11010 | | | | | | | | | | | |

7.1 Report # 1020195

S20102 COC



www.eurofins.com.au

EnviroSales@eurofins.com

NZBN: 9429046024954

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne 6 Monterey Road Dandenong South VIC 3175 Tel: +61 3 8564 5000 Tel: +61 3 8564 5000 Tel: +61 2 9900 8400 NATA# 1261

Geelong 19/8 Lewalan Street Grovedale VIC 3216 NATA# 1261 Site# 25403

179 Magowar Road Girraween NSW 2145 NATA# 1261 Site# 18217

Canberra Unit 1.2 Dacre Street Mitchell ACT 2911 Tel: +61 2 6113 8091 NATA# 1261 Site# 25466

Brisbane Newcastle 1/21 Smallwood Place 1/2 Frost Drive Murarrie QLD 4172 Tel: +61 7 3902 4600 NATA# 1261 NATA# 1261 Site# 20794

Mayfield West NSW 2304 Tel: +61 2 4968 8448 Site# 25079 & 25289

Perth 46-48 Banksia Road Welshpool WA 6106 Tel: +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 91 05 0159 898

Eurofins ARL Pty Ltd

Auckland Christchurch 35 O'Rorke Road 43 Detroit Drive Penrose, Rolleston, Auckland 1061 Christchurch 7675 Tel: +64 9 526 4551 Tel: +64 3 343 5201 IANZ# 1327 IANZ# 1290

Eurofins Environment Testing NZ Ltd

Tauranga 1277 Cameron Road Gate Pa, Tauranga 3112 IANZ# 1402

Sample Receipt Advice

Company name:

Senversa Pty Ltd NSW

Contact name: Project name: Project ID:

Bec Chapple REDIRECT S20102 1 Day

Turnaround time: Date/Time received

Aug 25, 2023 12:11 PM

Eurofins reference 1020195

Sample Information

A detailed list of analytes logged into our LIMS, is included in the attached summary table.

All samples have been received as described on the above COC.

COC has been completed correctly.

Attempt to chill was evident.

Appropriately preserved sample containers have been used.

All samples were received in good condition.

Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.

Appropriate sample containers have been used.

Sample containers for volatile analysis received with zero headspace.

Split sample sent to requested external lab.

X Some samples have been subcontracted.

N/A Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Hannah Mawbey on phone: or by email: Hannah Mawbey@eurofins.com

Results will be delivered electronically via email to Bec Chapple - bec.chapple@senversa.com.au.

Note: A copy of these results will also be delivered to the general Senversa Pty Ltd NSW email address.





web: www.eurofins.com.au email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

NATA# 1261

Site# 25403

ABN: 50 005 085 521

NATA# 1261

Time

Water

Site# 1254

Melbourne 6 Monterey Road Dandenong South VIC 3175

Geelong Sydney 19/8 Lewalan Street 179 Magowar Road Grovedale Girraween VIC 3216 NSW 2145

NATA# 1261

Site# 18217

S23-Au0064866

Χ

1

Χ

Χ Χ Χ

1

Canberra Unit 1.2 Dacre Street Mitchell ACT 2911

NATA# 1261

Site# 25466

Brisbane Newcastle 1/21 Smallwood Place 1/2 Frost Drive Murarrie Mayfield West NSW 2304 QLD 4172 Tel: +61 2 4968 8448 Tel: +61 3 8564 5000 Tel: +61 3 8564 5000 Tel: +61 2 9900 8400 Tel: +61 2 6113 8091 Tel: +61 7 3902 4600 NATA# 1261 NATA# 1261 Site# 25079 & 25289 Site# 20794

ABN: 91 05 0159 898

46-48 Banksia Road

Tel: +61 8 6253 4444

Received:

Priority:

Contact Name:

Due:

Perth

Welshpool

WA 6106

NATA# 2377

Site# 2370

NZBN: 9429046024954

Auckland Christchurch Tauranga 35 O'Rorke Road 43 Detroit Drive 1277 Cameron Road. Penrose, Rolleston. Gate Pa, Auckland 1061 Christchurch 7675 Tauranga 3112 Tel: +64 9 526 4551 Tel: +64 3 343 5201 Tel: +64 9 525 0568 IANZ# 1327 IANZ# 1290 IANZ# 1402

Aug 25, 2023 12:11 PM

Aug 28, 2023

Bec Chapple

1 Dav

Company Name:

Address:

QC202

Test Counts

Senversa Pty Ltd NSW

Level 24, 1 Market Street SYDNEY

NSW 2000

Project Name: Project ID:

REDIRECT S20102

Aug 14, 2023

Order No.:

Phone:

Report #:

1020195 02 9994 8016

03 9606 0074 Fax:

Eurofins Analytical Services Manager: Hannah Mawbey

| | | Sa | mple Detail | | | ron | Manganese | Phosphate total (as P) | Total Nitrogen (as N) | Eurofins Suite B7 |
|--|--|-------------|-------------|--------|--------|-----|-----------|------------------------|-----------------------|-------------------|
| Melb | Melbourne Laboratory - NATA # 1261 Site # 1254 | | | | | | | | Х | Х |
| Sydney Laboratory - NATA # 1261 Site # 18217 | | | | | | | Х | Х | | Х |
| External Laboratory | | | | | | | | | | |
| No | Sample ID | Sample Date | Sampling | Matrix | LAB ID | | | | | |



Senversa Pty Ltd NSW Level 24, 1 Market Street SYDNEY NSW 2000





NATA Accredited Accreditation Number 1261 Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: Bec Chapple

Report1020195-WProject nameREDIRECTProject ID\$20102Received DateAug 25, 2023

| Client Sample ID | | | QC202 |
|---|-----------|------|-------------------|
| Sample Matrix | | | Water |
| Eurofins Sample No. | | | S23- Au0064866 |
| Date Sampled | | | Aug 14, 2023 |
| Test/Reference | LOR | Unit | |
| Total Recoverable Hydrocarbons | | 0 | |
| TRH C6-C9 | 0.02 | mg/L | < 0.02 |
| TRH C10-C14 | 0.05 | mg/L | < 0.05 |
| TRH C15-C28 | 0.1 | mg/L | < 0.1 |
| TRH C29-C36 | 0.1 | mg/L | < 0.1 |
| TRH C10-C36 (Total) | 0.1 | mg/L | < 0.1 |
| TRH C6-C10 | 0.02 | mg/L | < 0.02 |
| TRH C6-C10 less BTEX (F1)N04 | 0.02 | mg/L | < 0.02 |
| TRH >C10-C16 | 0.05 | mg/L | < 0.05 |
| TRH >C10-C16 less Naphthalene (F2) ^{N01} | 0.05 | mg/L | < 0.05 |
| TRH >C16-C34 | 0.1 | mg/L | < 0.1 |
| TRH >C34-C40 | 0.1 | mg/L | < 0.1 |
| TRH >C10-C40 (total)* | 0.1 | mg/L | < 0.1 |
| BTEX | | | |
| Benzene | 0.001 | mg/L | < 0.001 |
| Toluene | 0.001 | mg/L | < 0.001 |
| Ethylbenzene | 0.001 | mg/L | < 0.001 |
| m&p-Xylenes | 0.002 | mg/L | < 0.002 |
| o-Xylene | 0.001 | mg/L | < 0.001 |
| Xylenes - Total* | 0.003 | mg/L | < 0.003 |
| 4-Bromofluorobenzene (surr.) | 1 | % | 95 |
| Total Recoverable Hydrocarbons - 2013 NEPM | Fractions | | |
| Naphthalene ^{N02} | 0.01 | mg/L | < 0.01 |
| Polycyclic Aromatic Hydrocarbons | | | |
| Acenaphthene | 0.001 | mg/L | < 0.001 |
| Acenaphthylene | 0.001 | mg/L | < 0.001 |
| Anthracene | 0.001 | mg/L | < 0.001 |
| Benz(a)anthracene | 0.001 | mg/L | < 0.001 |
| Benzo(a)pyrene | 0.001 | mg/L | < 0.001 |
| Benzo(b&j)fluorantheneN07 | 0.001 | mg/L | < 0.001 |
| Benzo(g.h.i)perylene | 0.001 | mg/L | < 0.001 |
| Benzo(k)fluoranthene | 0.001 | mg/L | < 0.001 |
| Chrysene | 0.001 | mg/L | < 0.001 |
| Dibenz(a.h)anthracene | 0.001 | mg/L | < 0.001 |
| Fluoranthene | 0.001 | mg/L | < 0.001 |
| Fluorene | 0.001 | mg/L | < 0.001 |

Report Number: 1020195-W



| Client Sample ID Sample Matrix | | | QC202 Water S23- |
|-----------------------------------|--------|------|------------------------|
| Eurofins Sample No. | | | Au0064866 |
| Date Sampled | | | Aug 14, 2023 |
| Test/Reference | LOR | Unit | |
| Polycyclic Aromatic Hydrocarbons | | | |
| Indeno(1.2.3-cd)pyrene | 0.001 | mg/L | < 0.001 |
| Naphthalene | 0.001 | mg/L | < 0.001 |
| Phenanthrene | 0.001 | mg/L | < 0.001 |
| Pyrene | 0.001 | mg/L | < 0.001 |
| Total PAH* | 0.001 | mg/L | < 0.001 |
| 2-Fluorobiphenyl (surr.) | 1 | % | 71 |
| p-Terphenyl-d14 (surr.) | 11 | % | 130 |
| Nitrate & Nitrite (as N) | 0.05 | mg/L | < 0.05 |
| Nitrate (as N) | 0.02 | mg/L | < 0.02 |
| Nitrite (as N) | 0.02 | mg/L | < 0.02 |
| Phosphate total (as P) | 0.01 | mg/L | 0.03 |
| Total Kjeldahl Nitrogen (as N) | 0.2 | mg/L | 0.5 |
| Total Nitrogen (as N)* | 0.2 | mg/L | 0.5 |
| Heavy Metals | | | |
| Arsenic | 0.001 | mg/L | 0.002 |
| Cadmium | 0.0002 | mg/L | < 0.0002 |
| Chromium | 0.001 | mg/L | 0.002 |
| Copper | 0.001 | mg/L | 0.002 |
| Iron | 0.05 | mg/L | 2.3 |
| Lead | 0.001 | mg/L | 0.002 |
| Manganese | 0.005 | mg/L | 5.9 |
| Mercury | 0.0001 | mg/L | < 0.0001 |
| Nickel | 0.001 | mg/L | 0.18 |
| Zinc | 0.005 | mg/L | 0.086 |

Report Number: 1020195-W



Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

| Description | Testing Site | Extracted | Holding Time |
|--|--------------|--------------|---------------------|
| Total Recoverable Hydrocarbons - 1999 NEPM Fractions | Sydney | Aug 25, 2023 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Aug 25, 2023 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Aug 25, 2023 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| BTEX | Sydney | Aug 25, 2023 | 14 Days |
| - Method: LTM-ORG-2010 BTEX and Volatile TRH | | | |
| Polycyclic Aromatic Hydrocarbons | Sydney | Aug 25, 2023 | 7 Days |
| - Method: LTM-ORG-2130 PAH and Phenols in Soil and Water | | | |
| Metals M8 | Sydney | Aug 28, 2023 | 28 Days |
| - Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS | | | |
| Total Nitrogen Set (as N) | | | |
| Nitrate & Nitrite (as N) | Melbourne | Aug 31, 2023 | 28 Days |
| - Method: LTM-INO-4120 Analysis of NOx NO2 NH3 by FIA | | | |
| Nitrate (as N) | Melbourne | Aug 31, 2023 | 28 Days |
| - Method: LTM-INO-4120 Analysis of NOx NO2 NH3 by FIA | | | |
| Nitrite (as N) | Melbourne | Aug 31, 2023 | 2 Days |
| - Method: LTM-INO-4120 Analysis of NOx NO2 NH3 by FIA | | | |
| Total Kjeldahl Nitrogen (as N) | Melbourne | Aug 31, 2023 | 28 Days |
| - Method: APHA 4500-Norg B,D Total Kjeldahl Nitrogen by FIA | | | |
| Phosphate total (as P) | Sydney | Aug 25, 2023 | 28 Days |
| - Method: E052 Total Phosphate (as P) | | | |
| Heavy Metals | Sydney | Aug 28, 2023 | 28 Days |
| - Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS | | | |

Report Number: 1020195-W



web: www.eurofins.com.au email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

NATA# 1261

Site# 25403

ABN: 50 005 085 521

NATA# 1261

Site# 1254

Melbourne Geelong 6 Monterey Road 19/8 Lewalan Street Dandenong South Grovedale VIC 3175 VIC 3216

Sydney 179 Magowar Road Girraween NSW 2145

NATA# 1261

Site# 18217

Canberra Mitchell ACT 2911

NATA# 1261

Site# 25466

Brisbane Newcastle Unit 1.2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Murarrie Mayfield West NSW 2304 QLD 4172 Tel: +61 2 4968 8448 Tel: +61 3 8564 5000 Tel: +61 3 8564 5000 Tel: +61 2 9900 8400 Tel: +61 2 6113 8091 Tel: +61 7 3902 4600 NATA# 1261 NATA# 1261 Site# 25079 & 25289 Site# 20794

ABN: 91 05 0159 898

46-48 Banksia Road

Tel: +61 8 6253 4444

Received:

Due:

Perth

Welshpool

WA 6106

NATA# 2377

Site# 2370

NZBN: 9429046024954

Auckland Christchurch Tauranga 35 O'Rorke Road 43 Detroit Drive 1277 Cameron Road. Penrose, Rolleston. Gate Pa. Auckland 1061 Christchurch 7675 Tauranga 3112 Tel: +64 9 526 4551 Tel: +64 3 343 5201 Tel: +64 9 525 0568 IANZ# 1327 IANZ# 1290 IANZ# 1402

Aug 25, 2023 12:11 PM

Aug 28, 2023

Company Name:

Address:

Senversa Pty Ltd NSW Level 24, 1 Market Street

SYDNEY

NSW 2000

Project Name: Project ID:

REDIRECT S20102

Order No.:

Report #: 1020195 02 9994 8016

Phone: 03 9606 0074 Fax:

Priority: 1 Dav **Contact Name:** Bec Chapple

Eurofins Analytical Services Manager: Hannah Mawbey

| | | Sa | mple Detail | | | Iron | Manganese | Phosphate total (as P) | Total Nitrogen (as N) | Eurofins Suite B7 |
|------|--|---------------|------------------|--------|---------------|------|-----------|------------------------|-----------------------|-------------------|
| Melb | Melbourne Laboratory - NATA # 1261 Site # 1254 | | | | | | | | Х | Х |
| Sydr | ney Laboratory | - NATA # 1261 | Site # 18217 | • | | Х | Х | Х | | Х |
| Exte | rnal Laboratory | | | | | | | | | |
| No | Sample ID | Sample Date | Sampling Time | Matrix | LAB ID | | | | | |
| 1 | QC202 | Aug 14, 2023 | | Water | S23-Au0064866 | Χ | Х | Х | Х | Х |
| Test | Counts | | | | | 1 | 1 | 1 | 1 | 1 |



Internal Quality Control Review and Glossary

General

- 1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follows guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013 and are included in this QC report where applicable. Additional QC data may be available on request.
- 2. All soil/sediment/solid results are reported on a dry basis, unless otherwise stated.
- 3. All biota/food results are reported on a wet weight basis on the edible portion, unless otherwise stated.
- 4. Actual LORs are matrix dependant. Quoted LORs may be raised where sample extracts are diluted due to interferences.
- 5. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds.
- 6. SVOC analysis on waters are performed on homogenised, unfiltered samples, unless noted otherwise
- 7. Samples were analysed on an 'as received' basis.
- 8. Information identified on this report with blue colour, indicates data provided by customer that may have an impact on the results.
- 9. This report replaces any interim results previously issued.

Holding Times

Please refer to 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours prior to sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and regardless of any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the date of sampling, therefore compliance to these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether the holding time is 7 days however for all other VOCs such as BTEX or C6-10 TRH then the holding time is 14 days.

Units

mg/kg: milligrams per kilogram mg/L: milligrams per litre µg/L: micrograms per litre

ppm: parts per million **ppb**: parts per billion
%: Percentage

org/100 mL: Organisms per 100 millilitres NTU: Nephelometric Turbidity Units MPN/100 mL: Most Probable Number of organisms per 100 millilitres

CFU: Colony forming unit

Terms

APHA American Public Health Association

COC Chain of Custody

CP Client Parent - QC was performed on samples pertaining to this report

CRM Certified Reference Material (ISO17034) - reported as percent recovery.

Dry Where a moisture has been determined on a solid sample the result is expressed on a dry basis.

Duplicate A second piece of analysis from the same sample and reported in the same units as the result to show comparison.

LOR Limit of Reporting

LCS Laboratory Control Sample - reported as percent recovery.

Method Blank

In the case of solid samples these are performed on laboratory certified clean sands and in the case of water samples these are performed on de-ionised water.

NCP

Non-Client Parent - QC performed on samples not pertaining to this report, QC is representative of the sequence or batch that client samples were analysed within.

RPD Relative Percent Difference between two Duplicate pieces of analysis.

SPIKE Addition of the analyte to the sample and reported as percentage recovery

SRA Sample Receipt Advice

Surr - Surrogate The addition of a like compound to the analyte target and reported as percentage recovery.

TBTO Tributyltin oxide (bis-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment however free tributyltin was measured

and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.

TCLP Toxicity Characteristic Leaching Procedure
TEQ Toxic Equivalency Quotient or Total Equivalence

QSM US Department of Defense Quality Systems Manual Version 5.4

US EPA United States Environmental Protection Agency

WA DWER Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should be used as a guide only and may be different when site specific Sampling Analysis and Quality Plan (SAQP) have been implemented

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is 30% however the following acceptance guidelines are equally applicable:

Results <10 times the LOR: No Limit

Results between 10-20 times the LOR: RPD must lie between 0-50%

Results >20 times the LOR: RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 - 150%

PFAS field samples that contain surrogate recoveries in excess of the QC limit designated in QSM 5.4 where no positive PFAS results have been reported have been reviewed and no data was affected.

QC Data General Comments

- 1. Where a result is reported as a less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
- 2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch, but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown is not data from your samples.
- 3. pH and Free Chlorine analysed in the laboratory Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time. Analysis will begin as soon as possible after sample receipt.
- 4. Recovery Data (Spikes & Surrogates) where chromatographic interference does not allow the determination of recovery the term "INT" appears against that analyte
- 5. For Matrix Spikes and LCS results a dash "-" in the report means that the specific analyte was not added to the QC sample.
- 6. Duplicate RPDs are calculated from raw analytical data thus it is possible to have two sets of data.



Quality Control Results

| Test | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|---|--------|---------------------------------------|----------------------|----------------|--------------------|
| Method Blank | | | | | |
| Total Recoverable Hydrocarbons | | | | | |
| TRH C6-C9 | mg/L | < 0.02 | 0.02 | Pass | |
| TRH C10-C14 | mg/L | < 0.05 | 0.05 | Pass | |
| TRH C15-C28 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C29-C36 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C6-C10 | mg/L | < 0.02 | 0.02 | Pass | |
| TRH >C10-C16 | mg/L | < 0.05 | 0.05 | Pass | |
| TRH >C16-C34 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH >C34-C40 | mg/L | < 0.1 | 0.1 | Pass | |
| Method Blank | | | | | |
| BTEX | | | | | |
| Benzene | mg/L | < 0.001 | 0.001 | Pass | |
| Toluene | mg/L | < 0.001 | 0.001 | Pass | |
| Ethylbenzene | mg/L | < 0.001 | 0.001 | Pass | |
| m&p-Xylenes | mg/L | < 0.002 | 0.001 | Pass | |
| o-Xylene | mg/L | < 0.002 | 0.002 | Pass | |
| Xylenes - Total* | mg/L | < 0.003 | 0.003 | Pass | |
| Method Blank | | \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ | 0.000 | 1 455 | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fraction | ne | | | | |
| Naphthalene | mg/L | < 0.01 | 0.01 | Pass | |
| Method Blank | IIIg/L | V 0.01 | 0.01 | rass | |
| Polycyclic Aromatic Hydrocarbons | | | | | |
| Acenaphthene | mg/L | < 0.001 | 0.001 | Pass | |
| Acenaphthylene | mg/L | < 0.001 | 0.001 | Pass | |
| Anthracene | | < 0.001 | 0.001 | Pass | |
| | mg/L | | | Pass | |
| Benz(a)anthracene | mg/L | < 0.001 | 0.001 | | |
| Benzo(a)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(b&j)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(g.h.i)perylene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(k)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Chrysene | mg/L | < 0.001 | 0.001 | Pass | |
| Dibenz(a.h)anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluorene | mg/L | < 0.001 | 0.001 | Pass | |
| Indeno(1.2.3-cd)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Naphthalene | mg/L | < 0.001 | 0.001 | Pass | |
| Phenanthrene | mg/L | < 0.001 | 0.001 | Pass | |
| Pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Method Blank | | 1 | | | |
| Heavy Metals | 1 | | | <u> </u> | |
| Arsenic | mg/L | < 0.001 | 0.001 | Pass | |
| Cadmium | mg/L | < 0.0002 | 0.0002 | Pass | |
| Chromium | mg/L | < 0.001 | 0.001 | Pass | |
| Copper | mg/L | < 0.001 | 0.001 | Pass | |
| Iron | mg/L | < 0.05 | 0.05 | Pass | |
| Lead | mg/L | < 0.001 | 0.001 | Pass | |
| Manganese | mg/L | < 0.005 | 0.005 | Pass | |
| Mercury | mg/L | 0.0001 | 0.0001 | Pass | |
| Nickel | mg/L | < 0.001 | 0.001 | Pass | |
| | mg/L | < 0.005 | 0.005 | Pass | l . |



| Test | | | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|----------------------------------|-----------------|--------|--------------|----------|----------------------|----------------|--------------------|
| Total Recoverable Hydrocarbons | | | | | | | |
| TRH C6-C9 | | | % | 78 | 70-130 | Pass | |
| TRH C10-C14 | | | % | 91 | 70-130 | Pass | |
| TRH C6-C10 | | | % | 78 | 70-130 | Pass | |
| TRH >C10-C16 | | | % | 89 | 70-130 | Pass | |
| LCS - % Recovery | | | | | | | |
| ВТЕХ | | | | | | | |
| Benzene | | | % | 95 | 70-130 | Pass | |
| Toluene | | | % | 86 | 70-130 | Pass | |
| Ethylbenzene | | | % | 85 | 70-130 | Pass | |
| m&p-Xylenes | | | % | 85 | 70-130 | Pass | |
| o-Xylene | | | % | 82 | 70-130 | Pass | |
| Xylenes - Total* | | | % | 84 | 70-130 | Pass | |
| LCS - % Recovery | | | | | | | |
| Total Recoverable Hydrocarbons - | 2013 NEPM Fract | tions | | | | | |
| Naphthalene | | | % | 95 | 70-130 | Pass | |
| LCS - % Recovery | | | | | | | |
| Polycyclic Aromatic Hydrocarbons | <u> </u> | | | | | | |
| Acenaphthene | | | % | 99 | 70-130 | Pass | |
| Acenaphthylene | | | % | 98 | 70-130 | Pass | |
| Anthracene | | | % | 113 | 70-130 | Pass | |
| Benz(a)anthracene | | | % | 91 | 70-130 | Pass | |
| Benzo(a)pyrene | | | % | 111 | 70-130 | Pass | |
| Benzo(b&j)fluoranthene | | | % | 109 | 70-130 | Pass | |
| Benzo(g.h.i)perylene | | | % | 118 | 70-130 | Pass | |
| Benzo(k)fluoranthene | | | % | 128 | 70-130 | Pass | |
| Chrysene | | | % | 125 | 70-130 | Pass | |
| Dibenz(a.h)anthracene | | | % | 84 | 70-130 | Pass | |
| Fluoranthene | | | % | 114 | 70-130 | Pass | |
| Fluorene | | | % | 111 | 70-130 | Pass | |
| Indeno(1.2.3-cd)pyrene | | | % | 100 | 70-130 | Pass | |
| Naphthalene | | | % | 83 | 70-130 | Pass | |
| Phenanthrene | | | % | 95 | 70-130 | Pass | |
| Pyrene | | | % | 113 | 70-130 | Pass | |
| LCS - % Recovery | | | 70 | 1.0 | 70 100 | 1 400 | |
| Heavy Metals | | | | | | | |
| Arsenic | | | % | 101 | 80-120 | Pass | |
| Cadmium | | | % | 101 | 80-120 | Pass | |
| Chromium | | | % | 102 | 80-120 | Pass | |
| Copper | | | % | 101 | 80-120 | Pass | |
| Iron | | | <u> </u> | 95 | 80-120 | Pass | |
| Lead | | | <u> </u> | 106 | 80-120 | Pass | |
| Manganese | | | % | 100 | 80-120 | Pass | |
| Mercury | | | % | 100 | 80-120 | Pass | |
| Nickel | | | % | 109 | 80-120 | Pass | |
| Zinc | | | % | 101 | 80-120 | Pass | |
| | | QA | | | Acceptance | Pass | Qualifying |
| Test | Lab Sample ID | Source | Units | Result 1 | Limits | Limits | Code |
| Spike - % Recovery | | | | T - | | | |
| Total Recoverable Hydrocarbons | 12 | | | Result 1 | <u> </u> | _ | |
| TRH C10-C14 | S23-Au0058328 | NCP | % | 73 | 70-130 | Pass | |
| TRH >C10-C16 | S23-Au0058328 | NCP | % | 72 | 70-130 | Pass | |
| Spike - % Recovery | | | | | T | | |
| Heavy Metals | 1 | | | Result 1 | | | |
| Arsenic | S23-Au0054217 | NCP | % | 117 | 75-125 | Pass | |



| Test | Lab Sample ID | QA Source | Units | Result 1 | | | Acceptance Limits | Pass Limits | Qualifying Code |
|--------------------------------|-------------------|--------------|-------|----------|----------|-----|----------------------|----------------|--------------------|
| Cadmium | L23-Au0051993 | NCP | % | 99 | | | 75-125 | Pass | |
| Chromium | L23-Au0051993 | NCP | % | 88 | | | 75-125 | Pass | |
| Copper | S23-Au0054217 | NCP | % | 85 | | | 75-125 | Pass | |
| Iron | L23-Au0051993 | NCP | % | 82 | | | 75-125 | Pass | |
| Lead | L23-Au0051993 | NCP | % | 81 | | | 75-125 | Pass | |
| Manganese | L23-Au0051993 | NCP | % | 91 | | | 75-125 | Pass | |
| Mercury | S23-Au0054217 | NCP | % | 97 | | | 75-125 | Pass | |
| Nickel | S23-Au0054217 | NCP | % | 89 | | | 75-125 | Pass | |
| Zinc | S23-Au0054217 | NCP | % | 85 | | | 75-125 | Pass | |
| Test | Lab Sample ID | QA Source | Units | Result 1 | | | Acceptance Limits | Pass Limits | Qualifying Code |
| Duplicate | | | | ī | 1 | | | | |
| Total Recoverable Hydrocarbons | 1 | , , | | Result 1 | Result 2 | RPD | | | |
| TRH C6-C9 | S23-Au0058331 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| TRH C10-C14 | S23-Au0058327 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| TRH C15-C28 | S23-Au0058327 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH C29-C36 | S23-Au0058327 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH C6-C10 | S23-Au0058331 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| TRH >C10-C16 | S23-Au0058327 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| TRH >C16-C34 | S23-Au0058327 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH >C34-C40 | S23-Au0058327 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| BTEX | | | | Result 1 | Result 2 | RPD | | | |
| Benzene | S23-Au0058331 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Toluene | S23-Au0058331 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Ethylbenzene | S23-Au0058331 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| m&p-Xylenes | S23-Au0058331 | NCP | mg/L | < 0.002 | < 0.002 | <1 | 30% | Pass | |
| o-Xylene | S23-Au0058331 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Xylenes - Total* | S23-Au0058331 | NCP | mg/L | < 0.003 | < 0.003 | <1 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| Total Recoverable Hydrocarbons | - 2013 NEPM Fract | ions | | Result 1 | Result 2 | RPD | | | |
| Naphthalene | S23-Au0058331 | NCP | mg/L | < 0.01 | < 0.01 | <1 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| Heavy Metals | | | | Result 1 | Result 2 | RPD | | | |
| Arsenic | S23-Au0066997 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Cadmium | S23-Au0066997 | NCP | mg/L | < 0.0002 | < 0.0002 | <1 | 30% | Pass | |
| Chromium | S23-Au0066997 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Copper | S23-Au0066997 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Iron | S23-Au0066997 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| Lead | S23-Au0066997 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Manganese | S23-Au0066997 | NCP | mg/L | < 0.005 | < 0.005 | <1 | 30% | Pass | |
| Mercury | S23-Au0066997 | NCP | mg/L | 0.0001 | 0.0001 | 4.8 | 30% | Pass | |
| Nickel | S23-Au0066997 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| THOROT | 020710000000 | 1101 | mg/ L | ₹ 0.001 | <u> </u> | | 3070 | 1 033 | |



Comments

Sample Integrity

Custody Seals Intact (if used) N/A Attempt to Chill was evident Yes Sample correctly preserved Yes Appropriate sample containers have been used Yes Sample containers for volatile analysis received with minimal headspace Yes Samples received within HoldingTime Yes Some samples have been subcontracted No

Qualifier Codes/Comments

Code Description

F2 is determined by arithmetically subtracting the "naphthalene" value from the ">C10-C16" value. The naphthalene value used in this calculation is obtained from volatiles (Purge & Trap analysis).

N01

Where we have reported both volatile (P&T GCMS) and semivolatile (GCMS) naphthalene data, results may not be identical. Provided correct sample handling protocols have been followed, any observed differences in results are likely to be due to procedural differences within each methodology. Results determined by both techniques have passed all QAQC acceptance criteria, and are entirely technically valid.

F1 is determined by arithmetically subtracting the "Total BTEX" value from the "C6-C10" value. The "Total BTEX" value is obtained by summing the concentrations of BTEX analytes. The "C6-C10" value is obtained by quantitating against a standard of mixed aromatic/aliphatic analytes. N04

Please note:- These two PAH isomers closely co-elute using the most contemporary analytical methods and both the reported concentration (and the TEQ) apply specifically to the total of the two co-eluting PAHs N07

Authorised by:

N02

Hannah Mawbey Analytical Services Manager Fang Yee Tan Senior Analyst-Metal Mary Makarios Senior Analyst-Inorganic Roopesh Rangarajan Senior Analyst-Organic Roopesh Rangarajan Senior Analyst-Volatile Ryan Phillips Senior Analyst-Inorganic



Final Report - this report replaces any previously issued Report

- Indicates Not Requested
- * Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here

Eurofins shall not be liable for loss, cost, damages or expenses incurred by the client, or any other person or company, resulting from the use of any information or interpretation given in this report. In no case shall Eurofins be liable for consequential damages including, but not limited to, lost profits, damages for failure to meet deadlines and lost production arising from this report. This document shall not be reproduced except in full and relates only to the items tested. Unless indicated otherwise, the tests were performed on the samples as received.

| Checked by | Completion | Completed | |
|------------|------------|-----------|--|
| | 100 | 34 | |
| | | | |
| | - | | |
| | - | 1 | |

Subcon Torward Lab / Split WO

Chain of Custody Documentation C 203

the field

| Senversa Pty Ltd | | | Laboratory: | ALS NSW | | | Ken | delinquisho: | 1777 | au. | Anal | Analysis Required | |
|---|--|--|--|--|---------------|---|-------------|-----------------|-------------|------|------------------------|---|--|
| ABN 89 132 231 380 | | | Address: Contact: | Sample Receipt | | | Lon | onno e / Cc | riei. | | | 100 | one the side for many one the teacher. We have |
| | | | Phone: | - | | | WO'N: | | Z | | 2 | N | LORs etc. |
| Job Number: | S2 | \$20102 | Purchase Order: | (Company) and the second and the sec | | F 2. | > | ttached By | PO/Internal | | | | and the control of th |
| Project Name: | Wetherill | Wetherill Park WME | Quote No: | EN/103/21 | | | | | | | - 5 | A Distance of the Part of the | The state of the s |
| Sampled By: | Rowi | Rowan Faint | Turn Around Time: | Standard 7 Days | ays | | 411/0 | | | | | 14) | |
| Project Manager: | Emm | Emma Walsh | Page: | 3 | of 1 | | | | | | | , IVI | Environmental Division |
| | rowan,faint@ | rowan.faint@senversa.com.au | | | ų. | H/BTE | 7/018 | | | 1 | | E AN | Sydney |
| Email Report To: | <u>Bec.Chapple@</u> | Bec.Chapple@sonversa.com.au; | Phone/Mobile: | 0408038593, 0404011544 | 4011544 | | - | | | d Tî | |) T (| Work Order Reference |
| | Sample Information | on | | Container Information | mation | | -20 (| | -8 | and | | | ES2404239 |
| Lab ID Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | - | VV- | | NT | TP | | | |
| QC404 | W | 9/02/2024 | | VOA | 1 × | _ | | | | | | | |
| QC504 | W | 9/02/2024 | | VOA | X | | | .5 | , | | | | |
| Z ac303 | W | 9/02/2024 | | P, VS x2, N, UA, VSA | O | × | | | | × | | | |
| C. MW1 | W | 9/02/2024 | | P, VS x2, N, UA, VSA | 6 | × | | | × | | × | | |
| | W | 9/02/2024 | | P, VS x2, N, UA, VSA | on . | × | | | × | | × | - | ■ ここのであることであった。 ■ ここのであることである。 |
| N/M/3 | W | 9/02/2024 | | P, VS x2, N, UA, VSA | 0 | × | | | × | | × | | Telephone: +61-2-8784 8555 |
| | W | 9/02/2024 | | P, VS x2, N, UA, VSA | Ø | × | | | × | | × | | |
| 2 00103 | W | 9/02/2024 | | P, VS x2, N, UA, VSA | 6 | × | | | | × | × | | |
| QC203 | W | 9/02/2024 | | P, VS x2, N, UA, VSA | 6 | × | | | | × | | | Please forward to Eurofins |
| | * | | | | | | | | | | | | |
| | | | | | | | 1 | | | | _ | | |
| | | | | | | | | | | | | | |
| L L | | | | | | | | | | | | | |
| | | | | | | | | | | | | | |
| | | | | | | | | | | | | | |
| μŝ | | | | | | - | | | | | - | | |
| | | | | | | | | | | | | | |
| Total | Special and the second | ACCORD AND AND AND AND AND AND AND AND AND AN | is the factories of the contract of the contra | Approximation of the state of t | 44 | | | - | | | | | |
| Sampler: I aftest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: | sampling procedures in a | accordance with S | enversa standard proce | edures and/or project | Sampler Name: | R | Rowan Faint | | Signature: | re: | 2 rack | E A | Date: 9/02/2024 |
| Relinquished By: | de des en de des constructions en construction de des des des des des des des des des | scelosios/policepolicerenzoowy/sersoy-unitate-abolosione | | Method of Shipment (if applicable): | olicable): | | Rece | Received by: | | | NAME OF TAXABLE PARTY. | | Name y depoilement registrapoid and conscional conscional companies (secure property and constitution of the constitution of t |
| Name/Signature: | Rowan Faint | | Date: 9/2/24 | Carrier / Reference #: | | | Nam | Name/Signature: | e Was | - | なく | | Date: 9/2/24 |
| Of. | Manager of the Common C | | Time: 3:30 PM | Date/Time: | | | Of: | A | S | | | | Time: (630 |
| Name/Signature: | | | Date: | Carrier / Reference #: | | | Nam | Name/Signature: | 9 | | | | Date: 3 |
| Of: | Inflact ferminds do planetomente (consparant) eventas consistentes proposes | oth finicipally-structure description of the commence of commentation described the structure of the structu | Time: | Date/Time: | | W. C. | Of: | | | | | | l. |
| Name/Signature: | | | Date: | Carrier / Reference #: | | | Nam | Name/Signature | (D) | | | | Date: 10/0 11:10 |
| Of: | CONTROL (AND INCIDENT AND INCID | | Time: | Time: Date/Time: Time: | | | Q. | | | | | | 101 |

2996901

"VEX VIA Vial Hygochloric Acid (HCI) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCI Preserved Plastic; HS = HCI Preserved Speciation Bottle; SP = Sulphuric Preserved Vial Hygochloric Area (HCI) Preserved Southe; E = EDTA Preserved Wide mouth glass jar in the preserved Wide P

COC_RF_GW sampling.xlsx



www.eurofins.com.au

EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne 6 Monterey Road Dandenong South VIC 3175 +61 3 8564 5000 NATA# 1261 Site# 1254

Geelong Grovedale VIC 3216 +61 3 8564 5000 NATA# 1261 Site# 25403

Sydney Girraween NSW 2145 +61 2 9900 8400 NATA# 1261 Site# 18217

Canberra Brisbane Mitchell Murarrie ACT 2911 QLD 4172 +61 2 6113 8091 NATA# 1261 Site# 20794

Newcastle 19/8 Lewalan Street 179 Magowar Road Unit 1,2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Mayfield West NSW 2304 +61 2 4968 8448 T: +61 7 3902 4600 NATA# 1261 NATA# 1261 Site# 25079 & 25289

46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 91 05 0159 898

Auckland IANZ# 1327

Eurofins ARL Pty Ltd Eurofins Environment Testing NZ Ltd

NZBN: 9429046024954

Auckland (Asb) 35 O'Rorke Road Unit C1/4 Pacific Rise, 43 Detroit Drive Mount Wellington, IANZ# 1308

Tauranga 1277 Cameron Road Rolleston, Gate Pa, Christchurch 7675 Tauranga 3112 +64 3 343 5201 IANZ# 1290 IANZ# 1402

Sample Receipt Advice

Company name:

Senversa Pty Ltd NSW

Contact name:

Emma Walsh

Project name:

WETHERILL PARK WME

Project ID: Turnaround time:

S20102 5 Day

Date/Time received

Feb 12, 2024 10:00 AM

Eurofins reference

1067666

Sample Information

A detailed list of analytes logged into our LIMS, is included in the attached summary table.

All samples have been received as described on the above COC.

COC has been completed correctly.

Attempt to chill was evident.

Appropriately preserved sample containers have been used.

All samples were received in good condition.

Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.

Appropriate sample containers have been used.

Sample containers for volatile analysis received with zero headspace.

Split sample sent to requested external lab.

X Some samples have been subcontracted.

N/A Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Adam Bateup on phone : or by email: AdamBateup@eurofins.com

Results will be delivered electronically via email to Emma Walsh - Emma.Walsh@senversa.com.au.

Note: A copy of these results will also be delivered to the general Senversa Pty Ltd NSW email address.





email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne 6 Monterey Road Dandenong South VIC 3175

+61 3 8564 5000

NATA# 1261

Site# 1254

Geelong 19/8 Lewalan Street Grovedale VIC 3216 +61 3 8564 5000 NATA# 1261 Site# 25403

Sydney 179 Magowar Road Girraween NSW 2145 +61 2 9900 8400 NATA# 1261 Site# 18217

Canberra Mitchell ACT 2911 +61 2 6113 8091 NATA# 1261 Site# 25466

Brisbane Newcastle Unit 1.2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Murarrie Mayfield West QLD 4172 NSW 2304 T: +61 7 3902 4600 +61 2 4968 8448 NATA# 1261 NATA# 1261 Site# 20794 Site# 25079 & 25289

Perth 46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 91 05 0159 898

Auckland Auckland (Asb) 35 O'Rorke Road Unit C1/4 Pacific Rise, 43 Detroit Drive Penrose, Mount Wellington, Auckland 1061 Auckland 1061 +64 9 526 4551 +64 9 525 0568 IANZ# 1327 IANZ# 1308

Received:

Priority:

Contact Name:

Due:

Eurofins ARL Pty Ltd Eurofins Environment Testing NZ Ltd

NZBN: 9429046024954

Rolleston, IANZ# 1290

Feb 19, 2024

Emma Walsh

Feb 12, 2024 10:00 AM

Christchurch Tauranga 1277 Cameron Road. Gate Pa, Christchurch 7675 Tauranga 3112 +64 3 343 5201 +64 9 525 0568 IANZ# 1402

Company Name:

Address:

web: www.eurofins.com.au

Senversa Pty Ltd NSW

Level 24, 1 Market Street SYDNEY

NSW 2000

Project Name:

WETHERILL PARK WME

Project ID:

S20102

Order No.: Report #:

Phone:

1067666 02 9994 8016

03 9606 0074 Fax:

Eurofins Analytical Services Manager: Adam Bateup

5 Day

| | | Sa | mple Detail | | | Eurofins Suite B7 | Eurofins Suite B19A: Total N (TKN, NOx), Total P | |
|--|-----------------|--------------|------------------|--------|---------------|-------------------|--|--|
| Melbourne Laboratory - NATA # 1261 Site # 1254 | | | | | | | | |
| Sydr | ney Laboratory | NATA # 1261 | Site # 18217 | • | | Х | Х | |
| Exte | rnal Laboratory | | | | | | | |
| No | Sample ID | Sample Date | Sampling Time | Matrix | LAB ID | | | |
| 1 | QC203 | Feb 09, 2024 | | Water | S24-Fe0028297 | Х | Х | |
| Test | Counts | | | | | 1 | 1 | |



Senversa Pty Ltd NSW Level 24, 1 Market Street SYDNEY NSW 2000





NATA Accredited Accreditation Number 1261 Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: Emma Walsh

Report 1067666-W

Project name WETHERILL PARK WME

Project ID S20102
Received Date Feb 12, 2024

| Client Sample ID | | | QC203 |
|--|-----------|------|---------------|
| Sample Matrix | | | Water |
| Eurofins Sample No. | | | S24-Fe0028297 |
| Date Sampled | | | Feb 09, 2024 |
| Test/Reference | LOR | Unit | |
| Total Recoverable Hydrocarbons | <u>'</u> | ' | |
| TRH C6-C9 | 0.02 | mg/L | < 0.02 |
| TRH C10-C14 | 0.05 | mg/L | < 0.05 |
| TRH C15-C28 | 0.1 | mg/L | < 0.1 |
| TRH C29-C36 | 0.1 | mg/L | < 0.1 |
| TRH C10-C36 (Total) | 0.1 | mg/L | < 0.1 |
| TRH C6-C10 | 0.02 | mg/L | < 0.02 |
| TRH C6-C10 less BTEX (F1)N04 | 0.02 | mg/L | < 0.02 |
| TRH >C10-C16 | 0.05 | mg/L | < 0.05 |
| TRH >C10-C16 less Naphthalene (F2)N01 | 0.05 | mg/L | < 0.05 |
| TRH >C16-C34 | 0.1 | mg/L | < 0.1 |
| TRH >C34-C40 | 0.1 | mg/L | < 0.1 |
| TRH >C10-C40 (total)* | 0.1 | mg/L | < 0.1 |
| ВТЕХ | · | , | |
| Benzene | 0.001 | mg/L | < 0.001 |
| Toluene | 0.001 | mg/L | < 0.001 |
| Ethylbenzene | 0.001 | mg/L | < 0.001 |
| m&p-Xylenes | 0.002 | mg/L | < 0.002 |
| o-Xylene | 0.001 | mg/L | < 0.001 |
| Xylenes - Total* | 0.003 | mg/L | < 0.003 |
| 4-Bromofluorobenzene (surr.) | 1 | % | 73 |
| Total Recoverable Hydrocarbons - 2013 NEPM | Fractions | | |
| Naphthalene ^{N02} | 0.01 | mg/L | < 0.01 |
| Polycyclic Aromatic Hydrocarbons | | | |
| Acenaphthene | 0.001 | mg/L | < 0.001 |
| Acenaphthylene | 0.001 | mg/L | < 0.001 |
| Anthracene | 0.001 | mg/L | < 0.001 |
| Benz(a)anthracene | 0.001 | mg/L | < 0.001 |
| Benzo(a)pyrene | 0.001 | mg/L | < 0.001 |
| Benzo(b&j)fluoranthene ^{N07} | 0.001 | mg/L | < 0.001 |
| Benzo(g.h.i)perylene | 0.001 | mg/L | < 0.001 |
| Benzo(k)fluoranthene | 0.001 | mg/L | < 0.001 |
| Chrysene | 0.001 | mg/L | < 0.001 |
| Dibenz(a.h)anthracene | 0.001 | mg/L | < 0.001 |
| Fluoranthene | 0.001 | mg/L | < 0.001 |
| Fluorene | 0.001 | mg/L | < 0.001 |



| Client Sample ID Sample Matrix | | | QC203 Water |
|-----------------------------------|--------|------|----------------|
| Eurofins Sample No. | | | S24-Fe0028297 |
| Date Sampled | | | Feb 09, 2024 |
| Test/Reference | LOR | Unit | |
| Polycyclic Aromatic Hydrocarbons | | | |
| Indeno(1.2.3-cd)pyrene | 0.001 | mg/L | < 0.001 |
| Naphthalene | 0.001 | mg/L | < 0.001 |
| Phenanthrene | 0.001 | mg/L | < 0.001 |
| Pyrene | 0.001 | mg/L | < 0.001 |
| Total PAH* | 0.001 | mg/L | < 0.001 |
| 2-Fluorobiphenyl (surr.) | 1 | % | 78 |
| p-Terphenyl-d14 (surr.) | 1 | % | 134 |
| | | | |
| Nitrate & Nitrite (as N) | 0.05 | mg/L | 0.11 |
| Nitrate (as N) | 0.02 | mg/L | 0.10 |
| Nitrite (as N) | 0.02 | mg/L | < 0.02 |
| Phosphate total (as P) | 0.01 | mg/L | 0.05 |
| Total Kjeldahl Nitrogen (as N) | 0.2 | mg/L | 1.3 |
| Total Nitrogen (as N)* | 0.2 | mg/L | 1.4 |
| Heavy Metals | | | |
| Arsenic | 0.001 | mg/L | 0.011 |
| Cadmium | 0.0002 | mg/L | < 0.0002 |
| Chromium | 0.001 | mg/L | 0.005 |
| Copper | 0.001 | mg/L | 0.006 |
| Lead | 0.001 | mg/L | 0.005 |
| Mercury | 0.0001 | mg/L | < 0.0001 |
| Nickel | 0.001 | mg/L | 0.16 |
| Zinc | 0.005 | mg/L | 0.18 |



Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

| Description | Testing Site | Extracted | Holding Time |
|---|--------------|--------------|---------------------|
| Total Recoverable Hydrocarbons - 1999 NEPM Fractions | Sydney | Feb 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Feb 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Feb 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| BTEX | Sydney | Feb 17, 2024 | 14 Days |
| - Method: LTM-ORG-2010 BTEX and Volatile TRH | | | |
| Polycyclic Aromatic Hydrocarbons | Sydney | Feb 17, 2024 | 7 Days |
| - Method: LTM-ORG-2130 PAH and Phenols in Soil and Water | | | |
| Metals M8 | Sydney | Feb 17, 2024 | 28 Days |
| - Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS | | | |
| Total Nitrogen Set (as N) | | | |
| Nitrate & Nitrite (as N) | Melbourne | Feb 21, 2024 | 28 Days |
| - Method: LTM-INO-4450 Determination of Nitrogen Species by Discrete Analyser | | | |
| Nitrate (as N) | Melbourne | Feb 21, 2024 | 28 Days |
| - Method: LTM-INO-4450 Determination of Nitrogen Species by Discrete Analyser | | | |
| Nitrite (as N) | Melbourne | Feb 21, 2024 | 2 Days |
| - Method: LTM-INO-4450 Determination of Nitrogen Species by Discrete Analyser | | | |
| Total Kjeldahl Nitrogen (as N) | Melbourne | Feb 21, 2024 | 28 Days |
| - Method: APHA 4500-Norg B,D Total Kjeldahl Nitrogen by FIA | | | |
| Eurofins Suite B19A: Total N (TKN, NOx), Total P | | | |
| Phosphate total (as P) | Sydney | Feb 17, 2024 | 28 Days |
| - Method: E052 Total Phosphate (as P) | | | |



email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne 6 Monterey Road Dandenong South VIC 3175 +61 3 8564 5000 NATA# 1261

Geelong 19/8 Lewalan Street Grovedale VIC 3216 +61 3 8564 5000 NATA# 1261 Site# 25403

Canberra Sydney 179 Magowar Road Girraween Mitchell NSW 2145 ACT 2911 +61 2 9900 8400 +61 2 6113 8091 NATA# 1261 NATA# 1261 Site# 18217 Site# 25466

Brisbane Newcastle Unit 1.2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Murarrie Mayfield West QLD 4172 NSW 2304 T: +61 7 3902 4600 +61 2 4968 8448 NATA# 1261 NATA# 1261 Site# 20794

Site# 25079 & 25289

Perth 46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 91 05 0159 898

Auckland Auckland (Asb) 35 O'Rorke Road Unit C1/4 Pacific Rise. 43 Detroit Drive Penrose, Auckland 1061 +64 9 526 4551 IANZ# 1327

Due:

Received:

Priority:

Contact Name:

Eurofins ARL Pty Ltd Eurofins Environment Testing NZ Ltd

NZBN: 9429046024954

Mount Wellington, Auckland 1061 +64 9 525 0568 IANZ# 1308

Christchurch Tauranga 1277 Cameron Road. Rolleston, Gate Pa, Christchurch 7675 Tauranga 3112 +64 3 343 5201 +64 9 525 0568 IANZ# 1290 IANZ# 1402

Feb 12, 2024 10:00 AM

Feb 19, 2024

Emma Walsh

Company Name:

Address:

web: www.eurofins.com.au

Senversa Pty Ltd NSW Level 24, 1 Market Street

SYDNEY

Site# 1254

NSW 2000

Project Name:

WETHERILL PARK WME

Project ID: S20102 Order No.: Report #:

Phone:

1067666 02 9994 8016

03 9606 0074 Fax:

Eurofins Analytical Services Manager: Adam Bateup

5 Day

| | | Sa | mple Detail | | | Eurofins Suite B7 | Eurofins Suite B19A: Total N (TKN, NOx), Total P |
|------|-----------------|----------------|------------------|--------|---------------|-------------------|---|
| Melb | ourne Laborato | ry - NATA # 12 | 61 Site # 12 | 54 | | | Х |
| Sydr | ey Laboratory | - NATA # 1261 | Site # 18217 | , | | Х | Х |
| Exte | rnal Laboratory | | | | | | |
| No | Sample ID | Sample Date | Sampling Time | Matrix | LAB ID | | |
| 1 | QC203 | Feb 09, 2024 | | Water | S24-Fe0028297 | Х | Х |
| Test | Counts | | | | | 1 | 1 |



Internal Quality Control Review and Glossary

General

- 1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request
- 2. All soil/sediment/solid results are reported on a dry weight basis unless otherwise stated.
- 3. All biota/food results are reported on a wet weight basis on the edible portion unless otherwise stated.
- 4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
- Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences
- Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds
- 7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
- 8. Samples were analysed on an 'as received' basis.
- 9. Information identified in this report with blue colour indicates data provided by customers that may have an impact on the results.
- 10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the date of sampling; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is 7 days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram mg/L: milligrams per litre ppm: parts per million μg/L: micrograms per litre ppb: parts per billion %: Percentage

org/100 mL: Organisms per 100 millilitres NTU: Nephelometric Turbidity Units MPN/100 mL: Most Probable Number of organisms per 100 millilitres

Colour: Pt-Co Units CFU: Colony forming unit

Terms

APHA American Public Health Association CEC Cation Exchange Capacity COC Chain of Custody

CP Client Parent - QC was performed on samples pertaining to this report CRM Certified Reference Material (ISO17034) - reported as percent recovery.

Dry Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis

Duplicate A second piece of analysis from the same sample and reported in the same units as the result to show comparison.

LOR Limit of Reporting.

LCS Laboratory Control Sample - reported as percent recovery.

Method Blank In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within. NCP

RPD Relative Percent Difference between two Duplicate pieces of analysis SPIKE Addition of the analyte to the sample and reported as percentage recovery

SRA Sample Receipt Advice

The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria Surr - Surrogate

Tributyltin oxide (bis-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits. TRTO

TCI P Toxicity Characteristic Leaching Procedure TEQ Toxic Equivalency Quotient or Total Equivalence

QSM US Department of Defense Quality Systems Manual Version 5.4

US EPA United States Environmental Protection Agency

WA DWER Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR: No Limit

Results between 10-20 times the LOR: RPD must lie between 0-50% Results >20 times the LOR: RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 - 150%, VOC recoveries 70 - 130%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 5.4, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

- 1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided
- 2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
- 3. pH and Free Chlorine analysed in the laboratory Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time. Analysis will begin as soon as possible after sample receipt.
- 4. Recovery Data (Spikes & Surrogates) where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
- 5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
- 6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data



Quality Control Results

| Test | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|--|--------------|----------|----------------------|----------------|--------------------|
| Method Blank | | | | | |
| Total Recoverable Hydrocarbons | | | | | |
| TRH C6-C9 | mg/L | < 0.02 | 0.02 | Pass | |
| TRH C10-C14 | mg/L | < 0.05 | 0.05 | Pass | |
| TRH C15-C28 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C29-C36 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C6-C10 | mg/L | < 0.02 | 0.02 | Pass | |
| TRH >C10-C16 | mg/L | < 0.05 | 0.05 | Pass | |
| TRH >C16-C34 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH >C34-C40 | mg/L | < 0.1 | 0.1 | Pass | |
| Method Blank | | | | | |
| BTEX | | | | | |
| Benzene | mg/L | < 0.001 | 0.001 | Pass | |
| Toluene | mg/L | < 0.001 | 0.001 | Pass | |
| Ethylbenzene | mg/L | < 0.001 | 0.001 | Pass | |
| m&p-Xylenes | mg/L | < 0.002 | 0.002 | Pass | |
| o-Xylene | mg/L | < 0.001 | 0.001 | Pass | |
| Xylenes - Total* | mg/L | < 0.003 | 0.003 | Pass | |
| Method Blank | 1 3 | | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | 3 | | | | |
| Naphthalene | mg/L | < 0.01 | 0.01 | Pass | |
| Method Blank | - 1 - 3 | | | | |
| Polycyclic Aromatic Hydrocarbons | | | | | |
| Acenaphthene | mg/L | < 0.001 | 0.001 | Pass | |
| Acenaphthylene | mg/L | < 0.001 | 0.001 | Pass | |
| Anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Benz(a)anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(a)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(b&j)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(g.h.i)perylene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(k)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Chrysene | mg/L | < 0.001 | 0.001 | Pass | |
| Dibenz(a.h)anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluorene | mg/L | < 0.001 | 0.001 | Pass | |
| Indeno(1.2.3-cd)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Naphthalene | mg/L | < 0.001 | 0.001 | Pass | |
| Phenanthrene | mg/L | < 0.001 | 0.001 | Pass | |
| Pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Method Blank | | | | | |
| Nitrate & Nitrite (as N) | mg/L | < 0.05 | 0.05 | Pass | |
| Nitrite (as N) | mg/L | < 0.02 | 0.02 | Pass | |
| Phosphate total (as P) | mg/L | < 0.01 | 0.01 | Pass | |
| Total Kjeldahl Nitrogen (as N) | mg/L | < 0.2 | 0.2 | Pass | |
| Method Blank | ,g, <u>_</u> | | | | |
| Heavy Metals | | | | | |
| Arsenic | mg/L | < 0.001 | 0.001 | Pass | |
| Cadmium | mg/L | < 0.0002 | 0.0002 | Pass | |
| Chromium | mg/L | < 0.001 | 0.001 | Pass | |
| Copper | mg/L | < 0.001 | 0.001 | Pass | |
| Lead | mg/L | < 0.001 | 0.001 | Pass | |
| | III9/⊏ | \ 0.001 | 0.001 | 1 1 433 | 1 |



| Test | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|---|-------|----------|----------------------|----------------|--------------------|
| Nickel | mg/L | < 0.001 | 0.001 | Pass | |
| Zinc | mg/L | < 0.005 | 0.005 | Pass | |
| LCS - % Recovery | | | | | |
| Total Recoverable Hydrocarbons | | | | | |
| TRH C6-C9 | % | 91 | 70-130 | Pass | |
| TRH C10-C14 | % | 96 | 70-130 | Pass | |
| TRH C6-C10 | % | 91 | 70-130 | Pass | |
| TRH >C10-C16 | % | 92 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| BTEX | | | | | |
| Benzene | % | 116 | 70-130 | Pass | |
| Toluene | % | 111 | 70-130 | Pass | |
| Ethylbenzene | % | 109 | 70-130 | Pass | |
| m&p-Xylenes | % | 111 | 70-130 | Pass | |
| o-Xylene | % | 107 | 70-130 | Pass | |
| Xylenes - Total* | % | 109 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Frac | tions | | | | |
| Naphthalene | % | 91 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Polycyclic Aromatic Hydrocarbons | | | | | |
| Acenaphthene | % | 100 | 70-130 | Pass | |
| Acenaphthylene | % | 96 | 70-130 | Pass | |
| Anthracene | % | 97 | 70-130 | Pass | |
| Benz(a)anthracene | % | 102 | 70-130 | Pass | |
| Benzo(a)pyrene | % | 117 | 70-130 | Pass | |
| Benzo(b&j)fluoranthene | % | 98 | 70-130 | Pass | |
| Benzo(g.h.i)perylene | % | 129 | 70-130 | Pass | |
| Benzo(k)fluoranthene | % | 120 | 70-130 | Pass | |
| Chrysene | % | 125 | 70-130 | Pass | |
| Dibenz(a.h)anthracene | % | 118 | 70-130 | Pass | |
| Fluoranthene | % | 116 | 70-130 | Pass | |
| Fluorene | % | 105 | 70-130 | Pass | |
| Indeno(1.2.3-cd)pyrene | % | 122 | 70-130 | Pass | |
| Naphthalene | % | 81 | 70-130 | Pass | |
| Phenanthrene | % | 98 | 70-130 | Pass | |
| Pyrene | % | 115 | 70-130 | Pass | |
| LCS - % Recovery | | | · | | |
| Nitrate & Nitrite (as N) | % | 117 | 70-130 | Pass | |
| Nitrite (as N) | % | 104 | 70-130 | Pass | |
| Phosphate total (as P) | % | 113 | 70-130 | Pass | |
| Total Kjeldahl Nitrogen (as N) | % | 107 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Heavy Metals | | | | | |
| Arsenic | % | 101 | 80-120 | Pass | |
| Cadmium | % | 100 | 80-120 | Pass | |
| Chromium | % | 102 | 80-120 | Pass | |
| Copper | % | 103 | 80-120 | Pass | |
| Lead | % | 102 | 80-120 | Pass | |
| Mercury | % | 101 | 80-120 | Pass | |
| Nickel | % | 102 | 80-120 | Pass | |
| Zinc | % | 101 | 80-120 | Pass | |



| Test | Lab Sample ID | QA Source | Units | Result 1 | | | Acceptance Limits | Pass Limits | Qualifying Code |
|---|--|--|---|---|--|--|--|---|--------------------|
| Spike - % Recovery | | | | | | | | | |
| Total Recoverable Hydrocarbons | | | | Result 1 | | | | | |
| TRH C6-C9 | S24-Fe0035298 | NCP | % | 84 | | | 70-130 | Pass | |
| TRH C10-C14 | S24-Fe0029342 | NCP | % | 88 | | | 70-130 | Pass | |
| TRH C6-C10 | S24-Fe0035298 | NCP | % | 82 | | | 70-130 | Pass | |
| TRH >C10-C16 | S24-Fe0029342 | NCP | % | 86 | | | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | | | |
| BTEX | | | | Result 1 | | | | | |
| Benzene | S24-Fe0035298 | NCP | % | 115 | | | 70-130 | Pass | |
| Toluene | S24-Fe0035298 | NCP | % | 93 | | | 70-130 | Pass | |
| Ethylbenzene | S24-Fe0035298 | NCP | % | 101 | | | 70-130 | Pass | |
| m&p-Xylenes | S24-Fe0035298 | NCP | % | 98 | | | 70-130 | Pass | |
| o-Xylene | S24-Fe0035298 | NCP | % | 99 | | | 70-130 | Pass | |
| Xylenes - Total* | S24-Fe0035298 | NCP | % | 99 | | | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | | | |
| Total Recoverable Hydrocarbons | - 2013 NEPM Fract | ions | | Result 1 | | | | | |
| Naphthalene | S24-Fe0035298 | NCP | % | 79 | | | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | | | |
| | | | | Result 1 | | | | | |
| Nitrate & Nitrite (as N) | L24-Fe0048715 | NCP | % | 112 | | | 70-130 | Pass | |
| Nitrite (as N) | L24-Fe0048715 | NCP | % | 108 | | | 70-130 | Pass | |
| Phosphate total (as P) | S24-Fe0030091 | NCP | % | 110 | | | 70-130 | Pass | |
| Total Kjeldahl Nitrogen (as N) | M24-Fe0051544 | NCP | % | 86 | | | 70-130 | Pass | |
| Spike - % Recovery | | - | | | | | | | |
| Heavy Metals | | | | Result 1 | | | | | |
| Arsenic | R24-Fe0030398 | NCP | % | 102 | | | 75-125 | Pass | |
| Cadmium | R24-Fe0030398 | NCP | % | 101 | | | 75-125 | Pass | |
| Chromium | R24-Fe0030398 | NCP | % | 101 | | | 75-125 | Pass | |
| Copper | R24-Fe0030398 | NCP | % | 102 | | | 75-125 | Pass | |
| Lead | R24-Fe0030398 | NCP | % | 101 | | | 75-125 | Pass | |
| Mercury | R24-Fe0030398 | NCP | % | 100 | | | 75-125 | Pass | |
| Nickel | R24-Fe0030398 | NCP | % | 103 | | | 75-125 | Pass | |
| Zinc | R24-Fe0030398 | NCP | % | 100 | | | 75-125 | Pass | |
| | | QA | | | | | Acceptance | Pass | Qualifying |
| Test | Lab Sample ID | Source | Units | Result 1 | | | Limits | Limits | Code |
| Duplicate | | | | | | | | | |
| Total Recoverable Hydrocarbons | | | | Result 1 | Result 2 | RPD | | | |
| TRH C6-C9 | S24-Fe0026053 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| TRH C10-C14 | S24-Fe0030079 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| TRH C15-C28 | S24-Fe0030079 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH C29-C36 | S24-Fe0030079 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH C6-C10 | 004 5-0000050 | NOD | | | < 0.02 | <1 | 30% | Pass | |
| | S24-Fe0026053 | NCP | mg/L | < 0.02 | < 0.02 | \ 1 | 00,0 | | |
| TRH >C10-C16 | S24-Fe0026053 S24-Fe0030079 | NCP NCP | | < 0.02 < 0.05 | < 0.02 | <1 | 30% | Pass | |
| | | NCP | mg/L | | < 0.05 | | | Pass Pass | |
| TRH >C10-C16 | S24-Fe0030079 | NCP NCP | mg/L mg/L | < 0.05 | | <1 | 30% | | |
| TRH >C10-C16 TRH >C16-C34 | S24-Fe0030079 S24-Fe0030079 | NCP | mg/L | < 0.05 < 0.1 | < 0.05 < 0.1 | <1 <1 | 30% 30% | Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 | S24-Fe0030079 S24-Fe0030079 | NCP NCP | mg/L mg/L | < 0.05 < 0.1 | < 0.05 < 0.1 | <1 <1 | 30% 30% | Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate | S24-Fe0030079 S24-Fe0030079 S24-Fe0030079 | NCP NCP NCP | mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 | < 0.05 < 0.1 < 0.1 | <1 <1 <1 | 30% 30% | Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX | S24-Fe0030079 S24-Fe0030079 | NCP NCP NCP | mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 | < 0.05 < 0.1 < 0.1 | <1 <1 <1 RPD | 30% 30% 30% | Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP | mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 | 30% 30% 30% 30% 30% | Pass Pass Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene Ethylbenzene | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP NCP NCP NCP | mg/L mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 | 30% 30% 30% 30% 30% 30% | Pass Pass Pass Pass Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene Ethylbenzene m&p-Xylenes | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP NCP NCP NCP NCP | mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 < 0.001 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 < 0.001 < 0.002 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 < | 30% 30% 30% 30% 30% 30% 30% | Pass Pass Pass Pass Pass Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene Ethylbenzene m&p-Xylenes o-Xylene | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP NCP NCP NCP NCP NCP | mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 < 0.001 < 0.002 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 < 0.001 < 0.002 < 0.001 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 < | 30% 30% 30% 30% 30% 30% 30% 30% | Pass Pass Pass Pass Pass Pass Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene Ethylbenzene m&p-Xylenes o-Xylene Xylenes - Total* | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP NCP NCP NCP NCP | mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 < 0.001 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 < 0.001 < 0.002 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 < | 30% 30% 30% 30% 30% 30% 30% | Pass Pass Pass Pass Pass Pass Pass | |
| TRH >C10-C16 TRH >C16-C34 TRH >C34-C40 Duplicate BTEX Benzene Toluene Ethylbenzene m&p-Xylenes o-Xylene | \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0030079 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 \$24-Fe0026053 | NCP NCP NCP NCP NCP NCP NCP NCP | mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L | < 0.05 < 0.1 < 0.1 Result 1 < 0.001 < 0.001 < 0.002 < 0.001 | < 0.05 < 0.1 < 0.1 Result 2 < 0.001 < 0.001 < 0.001 < 0.002 < 0.001 | <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 <1 < | 30% 30% 30% 30% 30% 30% 30% 30% | Pass Pass Pass Pass Pass Pass Pass Pass | |



| Duplicate | | | | | | | | | |
|--------------------------------|---------------|-----|------|----------|----------|-----|-----|------|--|
| • | | | | Result 1 | Result 2 | RPD | | | |
| Nitrate & Nitrite (as N) | M24-Fe0051355 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| Nitrite (as N) | M24-Fe0051355 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| Phosphate total (as P) | S24-Fe0043848 | NCP | mg/L | 0.04 | 0.04 | 2.3 | 30% | Pass | |
| Total Kjeldahl Nitrogen (as N) | M24-Fe0049917 | NCP | mg/L | 150 | 110 | 28 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| Heavy Metals | | | | Result 1 | Result 2 | RPD | | | |
| Arsenic | S24-Fe0030077 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Cadmium | S24-Fe0030077 | NCP | mg/L | < 0.0002 | < 0.0002 | <1 | 30% | Pass | |
| Chromium | S24-Fe0030077 | NCP | mg/L | 0.003 | 0.003 | 8.0 | 30% | Pass | |
| Copper | S24-Fe0030077 | NCP | mg/L | 0.011 | 0.011 | 4.5 | 30% | Pass | |
| Lead | S24-Fe0030077 | NCP | mg/L | 0.004 | 0.004 | 2.9 | 30% | Pass | |
| Mercury | S24-Fe0030077 | NCP | mg/L | < 0.0001 | < 0.0001 | <1 | 30% | Pass | |
| Nickel | S24-Fe0030077 | NCP | mg/L | 0.004 | 0.004 | 7.5 | 30% | Pass | |
| Zinc | S24-Fe0030077 | NCP | mg/L | 0.029 | 0.028 | 4.6 | 30% | Pass | |



Comments

Sample Integrity

Custody Seals Intact (if used) N/A Attempt to Chill was evident Yes Sample correctly preserved Yes Appropriate sample containers have been used Yes Sample containers for volatile analysis received with minimal headspace Yes Samples received within HoldingTime Yes Some samples have been subcontracted No

Qualifier Codes/Comments

Code Description

F2 is determined by arithmetically subtracting the "naphthalene" value from the ">C10-C16" value. The naphthalene value used in this calculation is obtained from volatiles (Purge & Trap analysis).

N01

Where we have reported both volatile (P&T GCMS) and semivolatile (GCMS) naphthalene data, results may not be identical. Provided correct sample handling protocols have been followed, any observed differences in results are likely to be due to procedural differences within each methodology. Results determined by both techniques have passed all QAQC acceptance criteria, and are entirely technically valid.

F1 is determined by arithmetically subtracting the "Total BTEX" value from the "C6-C10" value. The "Total BTEX" value is obtained by summing the concentrations of BTEX analytes. The "C6-C10" value is obtained by quantitating against a standard of mixed aromatic/aliphatic analytes. N04

Please note:- These two PAH isomers closely co-elute using the most contemporary analytical methods and both the reported concentration (and the TEQ) apply specifically to the total of the two co-eluting PAHs N07

Authorised by:

N02

Adam Bateup Analytical Services Manager Fang Yee Tan Senior Analyst-Metal Maria Tian Senior Analyst-Organic Mary Makarios Senior Analyst-Inorganic Roopesh Rangarajan Senior Analyst-Organic Roopesh Rangarajan Senior Analyst-Volatile Ryan Phillips Senior Analyst-Inorganic

Glenn Jackson **Managing Director**

Final Report - this report replaces any previously issued Report

- Indicates Not Requested
- * Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

Eurofins shall not be liable for loss, cost, damages or expenses incurred by the client, or any other person or company, resulting from the use of any information or interpretation given in this report. In no case shall Eurofins be liable for consequential damages including, but not limited to, lost profits, damages for failure to meet deadlines and lost production arising from this report. This document shall not be reproduced except in full and relates only to the items tested. Unless indicated otherwise, the tests were performed on the samples as received.

sonversa

Chain of Custody Documentation

| Senversa Pt | | | | Laboratory: | ALS NSW | | | _ | | | | | | malysis | Required | _ | |
|------------------------------|--|-------------------------|------------------------------------|--------------------------------|--|--------------------|---------------|---------------|--------------------------------------|---------------|--|------------------------------------|---------------|-----------|---------------|--------------|---|
| ABN 89 132 | | | | Address: Contact: Phone; | Sample Receipt | | | | | | | | | dialysis | Required | | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. |
| Job Number | n | \$2 | 0102 | Purchase Order: | | | | | | | | | | | | | |
| Project Nam | e: | | Park WME | Quote No: | EN(103\21 | | | | Ş Ç | | | | | | | | |
| Sampled By | : | Rowa | an Faint | Turn Around Time: | Standard | | | | Fe and Mn) | | etals | | | | | | |
| Project Man | ager: | | a Walsh | Page: | | of 1 | | | Fe " | | W W | NX | | | | | ' I |
| | | rowan.faint@: | senversa.com.au senversa.com.au | | | OI I | | | WATER - EG005F Dissolved Metals (| | WATER - W-26 TRH/BTEXN/PAH/8 Metals | WATER - W-18 TRH(C6 - C9)/BTEXN | - EP080 | | | | |
| Email Repor | t To: | | senversa.com.au | Phone/Mobile: | 0420 218 472 | | | | R-E | | TEX: | 8-9 0-8 | ~ - H | Z | | | |
| Lab ID | Sample ID | Sample Information | | - | Container Inform | | | | ATE | 8-TV | ATE H/B | ATE O | WATER BTEXN | and | | HOLD | |
| 2 | MW1 | Water | Date . | Time | Type / Code | Total Bottles | - | | | | _ | ≥ E | § ₽ | ₽ | | 포 | |
| | MVV2 | | 11/07/2024 | | 1xP, 2xVSA, 1xN, 1xUA, 1xSP | 6 | | | X | Х | Х | | | | | | |
| 3 | MVV3 | Water | 11/07/2024 | | 1xP, 2xVSA, 1xN, 1xUA, 1xSP | 6 | | | Х | Х | Х | | | | | | |
| 4 | | Water | 11/07/2024 | | 1xP, 2xVSA, 1xN, 1xUA, 1xSP | - 6 | | | Х | Х | Х | | | | | | |
| - | MW4 | Water | 11/07/2024 | | 1xP, 2xVSA, 1xN, 1xUA, 1xSP | 6 | | | Х | Х | Х | | | | | | |
| 5 | MVV6 | Water | 11/07/2024 | | 1xP, 2xVSA, 1xN, 1xUA, 1xSP | 6 | | | X | X | X | | | | | | *=== |
| 6 | QC104 | Water | 11/07/2024 | | 2xVSA, 1xN, 1xUA, 1xSP | 5 | | | Х | | Х | | | Х | | | |
| _ | QC204 | Water | 11/07/2024 | | 2xVSA, 1xN, 1xUA, 1xSP | 5 | | | Х | | Х | | | Х | | | Please forward to Eurofins |
| -3 | QC304 | Water | 11/07/2024 | | 2xVSA, 1xN, 1xUA, 1xSP | 5 | | | X | | Х | | | Х | | | |
| Je. | QC405 | Water | 11/07/2024 | | VSA | 1 | | | | | | X | | | | | |
| 34 | QC505 | Water | 11/07/2024 | | VSA | 1 | | | | | | ~ | Х | | _ | _ | |
| | | | | | | | $\overline{}$ | | | | - | - | ^ | | _ | + | - |
| | | | | | | | _ | _ | - | | | _ | _ | | _ | - | |
| | | | | | | | - | _ | _ | - | - | - | - | _ | _ | _ | |
| 8 | | | | | | | - | - | | _ | - | - | - | _ | | _ | |
| | | | | | | | - | | | | - | | _ | | _ | | |
| | | | - | | | | - | | _ | | | - | | | | | |
| | | - 7 | | - | | | | | | | | | | | | | |
| - | 1 | 11 | 111 | | | | | | | | | | | | | | |
| | 1// | | | | | | | | | | | | | | | | |
| | | - | | | | | | 孤 | | | | | | | | | |
| | | | | | | | | 9/1/20 | Nº S | | | | | | | 1 | |
| | | | | | | | | 懲 | | | | | | | | | |
| | N. | | 0 | | | | | 100 | | - 4 | | | | | | _ | |
| | | 1/0 | , | | | | | | | | \neg | - | | - | _ | + | |
| | | MD | | | | | \rightarrow | _ | | | | - | - | | _ | | |
| | | | | | | | - | - | _ | - | - | - | - | - | _ | - | |
| | | | | | | | - | - | \rightarrow | \rightarrow | - | - | - | - | _ | + | |
| Total | | | الرزائدي | | | 47 | | | - 3 | | | | | 31 | | | |
| Sampler: I at | test that proper field sampli s were used during the coll | ng procedures in a | ccordance with Se | nversa standard proc | edures and/or project | Sampler Name: | | Rowan | Faint | | Signatur | e: | $\overline{}$ | | | Date: | 11/07/2024 |
| | | ection of these san | ipies. | | | | | | _ | | _ | -1 | 4 | au | | | |
| Relinquished Name/Signati | | Bauma Estat | | 0.1 | Method of Shipment (if applica | able): | | | Receive | | | | | | | | 1-1- |
| Name/Signati Of: | | Rowan Faint Senversa | | Date: Time: | Carrier / Reference #: | | _ | $\overline{}$ | | gnature: | 7 | nenh | | L | | | Date: 11/7/17 |
| Name/Signate | | Octivored. | | Date: | Date/Time: | | | \rightarrow | Of: | | A | 0.0 | 1 | | | | Time: ,715 |
| Of: | | | | Time: | Carrier / Reference #: Date/Time: | | | | Vame/Si Of: | gnature: | _ | en | Jun. | _ | | | Date: 5/7/2 |
| Name/Signatu | ıre; | | | Date: | Carrier / Reference #: | | _ | | | gnature: | n | | | | _ | | Tittle. |
| Of: | | | | Time: | Date/Time: | | | | 26. | | | | | | | | Date: |
| W | fater Container Codes: P = Ur | preserved Plastic; N = | Nitric Acid (HNO ₃) Pr | eserved Plastic; ORC = N | iltric Preserved ORC; SH = Sodium Hy | droxide (NaOH)/Cad | Intium (Co | d) Presei | ved; S = | Sodium I | Hydroxide | Preserve | d Plastic | : STH = 8 | odium thiosul | fate preserv | ved plastic; |

V = VOA Vial Hydochloric Acid (HCI) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H ≈ HCI Preserved Plastic; HS = HCI Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST ≈ Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

1517



EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Geelong Melbourne 6 Monterey Road 19/8 Lewalan Street 179 Magowar Road Unit 1,2 Dacre Street Dandenong South Grovedale VIC 3175 VIC 3216 +61 3 8564 5000 +61 3 8564 5000 NATA# 1261 NATA# 1261 Site# 1254 Site# 25403

Girraween NSW 2145 +61 2 9900 8400 NATA# 1261 Site# 18217

Mitchell ACT 2911 +61 2 6113 8091 NATA# 1261 Site# 25466

Newcastle 1/21 Smallwood Place 1/2 Frost Drive Mayfield West Murarrie QLD 4172 NSW 2304 T: +61 7 3902 4600 +61 2 4968 8448 NATA# 1261 NATA# 1261 Site# 20794 & 2780 Site# 25079

Perth 46-48 Ranksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 91 05 0159 898

Perth ProMicro 46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2561 Site# 2554

ABN: 47 009 120 549

Penrose, Auckland 1061 +64 9 526 4551 IANZ# 1327

NZBN: 9429046024954

35 O'Rorke Road Unit C1/4 Pacific Rise 43 Detroit Drive Mount Wellington, Auckland 1061 +64 9 525 0568 IANZ# 1308

Tauranga 1277 Cameron Road Rolleston, Gate Pa, Christchurch 7675 Tauranga 3112 +64 3 343 5201 +64 9 525 0568 IANZ# 1290 IANZ# 1402

Sample Receipt Advice

Company name: Contact name: Project name: Project ID:

Turnaround time:

Senversa Pty Ltd NSW Emma Walsh WETHERILL PARK WME S20102

5 Day Jul 15, 2024 2:30 PM

Date/Time received 1117968 **Eurofins reference**

Sample Information

A detailed list of analytes logged into our LIMS, is included in the attached summary table.

All samples have been received as described on the above COC.

COC has been completed correctly.

Attempt to chill was evident.

Appropriately preserved sample containers have been used.

All samples were received in good condition.

Samples have been provided with adequate time to commence analysis in accordance with the relevant

Appropriate sample containers have been used.

Sample containers for volatile analysis received with zero headspace.

Split sample sent to requested external lab.

Some samples have been subcontracted.

N/A Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Adam Bateup on phone: or by email: AdamBateup@eurofins.com

Results will be delivered electronically via email to Emma Walsh - Emma.Walsh@senversa.com.au.

Note: A copy of these results will also be delivered to the general Senversa Pty Ltd NSW email address.





email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne Geelong Sydney Canberra Brisbane 6 Monterey Road 19/8 Lewalan Street 179 Magowar Road Unit 1,2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Dandenong South Grovedale Girraween Mitchell Murarrie VIC 3175 VIC 3216 NSW 2145 ACT 2911 QLD 4172 +61 3 8564 5000 +61 2 9900 8400 T: +61 7 3902 4600 +61 3 8564 5000 +61 2 6113 8091 NATA# 1261 NATA# 1261 NATA# 1261 NATA# 1261 NATA# 1261 Site# 20794 & 2780 Site# 1254 Site# 25403 Site# 18217 Site# 25466

ABN: 47 009 120 549

Perth ProMicro 46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2561 Site# 2554

Auckland Auckland (Focus) 35 O'Rorke Road Unit C1/4 Pacific Rise. Penrose, Mount Wellington, Auckland 1061 Auckland 1061 +64 9 526 4551 +64 9 525 0568 IANZ# 1327 IANZ# 1308

NZBN: 9429046024954

Received:

Contact Name:

Priority:

Due:

Christchurch 43 Detroit Drive Rolleston, Christchurch 7675 Tauranga 3112 +64 3 343 5201 IANZ# 1290

Jul 15, 2024 2:30 PM

Jul 22, 2024

Emma Walsh

5 Day

Tauranga 1277 Cameron Road. Gate Pa, +64 9 525 0568 IANZ# 1402

Address

web: www.eurofins.com.au

Company Name: Senversa Pty Ltd NSW Level 24, 1 Market Street

SYDNEY NSW 2000

Project Name: Project ID:

WETHERILL PARK WME

S20102

Order No.: Report #:

Perth

Welshpool

NATA# 2377

Site# 2370

WA 6106

Newcastle

Mayfield West

+61 2 4968 8448

NSW 2304

NATA# 1261

Site# 25079

ABN: 91 05 0159 898

46-48 Banksia Road

+61 8 6253 4444

1117968

Phone: 02 9994 8016 03 9606 0074 Fax:

Eurofins Analytical Services Manager: Adam Bateup

| | | Sa | mple Detail | | | Iron (filtered) | Manganese (filtered) | Total Recoverable Hydrocarbons | Eurofins Suite B7 (filtered metals) |
|------|-----------------|---------------|------------------|--------|---------------|-----------------|----------------------|--------------------------------|-------------------------------------|
| Sydr | ey Laboratory | - NATA # 1261 | Site # 18217 | , | | Χ | Х | Х | Χ |
| Exte | rnal Laboratory | | | | | | | | |
| No | Sample ID | Sample Date | Sampling Time | Matrix | LAB ID | | | | |
| 1 | QC204 | Jul 11, 2024 | | Water | S24-JI0037338 | Х | Х | Х | Х |
| Test | Counts | | | | | 1 | 1 | 1 | 1 |



Senversa Pty Ltd NSW Level 24, 1 Market Street SYDNEY NSW 2000





NATA Accredited Accreditation Number 1261 Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: Emma Walsh

Report 1117968-W

Project name WETHERILL PARK WME

Project ID S20102
Received Date Jul 15, 2024

| Client Sample ID | | | QC204 |
|--|-----------|------|---------------|
| Sample Matrix | | | Water |
| Eurofins Sample No. | | | S24-JI0037338 |
| Date Sampled | | | Jul 11, 2024 |
| Test/Reference | LOR | Unit | |
| Total Recoverable Hydrocarbons | ' | ' | |
| TRH C6-C9 | 0.02 | mg/L | < 0.02 |
| TRH C10-C14 | 0.05 | mg/L | 0.09 |
| TRH C15-C28 | 0.1 | mg/L | 0.4 |
| TRH C29-C36 | 0.1 | mg/L | 0.3 |
| TRH C10-C36 (Total) | 0.1 | mg/L | 0.79 |
| TRH C6-C10* | 0.02 | mg/L | < 0.02 |
| TRH C6-C10 less BTEX (F1)N04 | 0.02 | mg/L | < 0.02 |
| TRH >C10-C16* | 0.05 | mg/L | 0.11 |
| TRH >C10-C16 less Naphthalene (F2)*N01 | 0.05 | mg/L | 0.11 |
| TRH >C16-C34* | 0.1 | mg/L | 0.6 |
| TRH >C34-C40 | 0.1 | mg/L | 0.2 |
| TRH >C10-C40 (total)* | 0.1 | mg/L | 0.91 |
| ВТЕХ | • | | |
| Benzene | 0.001 | mg/L | < 0.001 |
| Toluene | 0.001 | mg/L | < 0.001 |
| Ethylbenzene | 0.001 | mg/L | < 0.001 |
| m&p-Xylenes | 0.002 | mg/L | < 0.002 |
| o-Xylene | 0.001 | mg/L | < 0.001 |
| Xylenes - Total* | 0.003 | mg/L | < 0.003 |
| 4-Bromofluorobenzene (surr.) | 1 | % | 73 |
| Total Recoverable Hydrocarbons - 2013 NEPM | Fractions | - | |
| Naphthalene ^{N02} | 0.01 | mg/L | < 0.01 |
| Polycyclic Aromatic Hydrocarbons | · | | |
| Acenaphthene | 0.001 | mg/L | < 0.001 |
| Acenaphthylene | 0.001 | mg/L | < 0.001 |
| Anthracene | 0.001 | mg/L | < 0.001 |
| Benz(a)anthracene | 0.001 | mg/L | < 0.001 |
| Benzo(a)pyrene | 0.001 | mg/L | < 0.001 |
| Benzo(b&j)fluoranthene ^{N07} | 0.001 | mg/L | < 0.001 |
| Benzo(g.h.i)perylene | 0.001 | mg/L | < 0.001 |
| Benzo(k)fluoranthene | 0.001 | mg/L | < 0.001 |
| Chrysene | 0.001 | mg/L | < 0.001 |
| Dibenz(a.h)anthracene | 0.001 | mg/L | < 0.001 |
| Fluoranthene | 0.001 | mg/L | < 0.001 |
| Fluorene | 0.001 | mg/L | < 0.001 |



| Client Sample ID Sample Matrix | | | QC204 Water |
|-----------------------------------|--------|------|----------------|
| Eurofins Sample No. | | | S24-JI0037338 |
| Date Sampled | | | Jul 11, 2024 |
| Test/Reference | LOR | Unit | |
| Polycyclic Aromatic Hydrocarbons | | | |
| Indeno(1.2.3-cd)pyrene | 0.001 | mg/L | < 0.001 |
| Naphthalene | 0.001 | mg/L | < 0.001 |
| Phenanthrene | 0.001 | mg/L | < 0.001 |
| Pyrene | 0.001 | mg/L | < 0.001 |
| Total PAH* | 0.001 | mg/L | < 0.001 |
| 2-Fluorobiphenyl (surr.) | 1 | % | 78 |
| p-Terphenyl-d14 (surr.) | 1 | % | 109 |
| Nitrate & Nitrite (as N) | 0.05 | mg/L | < 0.05 |
| Nitrate (as N) | 0.02 | mg/L | < 0.02 |
| Nitrite (as N) | 0.02 | mg/L | < 0.02 |
| Phosphate total (as P) | 0.01 | mg/L | 0.03 |
| Total Kjeldahl Nitrogen (as N) | 0.2 | mg/L | 1.0 |
| Total Nitrogen (as N)* | 0.2 | mg/L | 1.0 |
| Heavy Metals | | | |
| Arsenic (filtered) | 0.001 | mg/L | < 0.01 |
| Cadmium (filtered) | 0.0002 | mg/L | < 0.002 |
| Chromium (filtered) | 0.001 | mg/L | < 0.01 |
| Copper (filtered) | 0.001 | mg/L | < 0.01 |
| Iron (filtered) | 0.05 | mg/L | 7.4 |
| Lead (filtered) | 0.001 | mg/L | < 0.01 |
| Manganese (filtered) | 0.005 | mg/L | 7.4 |
| Mercury (filtered) | 0.0001 | mg/L | < 0.001 |
| Nickel (filtered) | 0.001 | mg/L | 0.20 |
| Zinc (filtered) | 0.005 | mg/L | 0.25 |



Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

| Description | Testing Site | Extracted | Holding Time |
|---|--------------|--------------|---------------------|
| Total Recoverable Hydrocarbons - 1999 NEPM Fractions | Sydney | Jul 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Jul 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fractions | Sydney | Jul 17, 2024 | 7 Days |
| - Method: LTM-ORG-2010 TRH C6-C40 | | | |
| BTEX | Sydney | Jul 17, 2024 | 14 Days |
| - Method: LTM-ORG-2010 BTEX and Volatile TRH | | | |
| Polycyclic Aromatic Hydrocarbons | Sydney | Jul 17, 2024 | 7 Days |
| - Method: LTM-ORG-2130 PAH and Phenols in Soil and Water | | | |
| Metals M8 filtered | Sydney | Jul 17, 2024 | 28 Days |
| - Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS | | | |
| Total Nitrogen Set (as N) | | | |
| Nitrate & Nitrite (as N) | Melbourne | Jul 17, 2024 | 28 Days |
| - Method: LTM-INO-4450 Determination of Nitrogen Species by Discrete Analyser | | | |
| Nitrate (as N) | Melbourne | Jul 17, 2024 | 28 Days |
| - Method: LTM-INO-4450 Determination of Nitrogen Species by Discrete Analyser | | | |
| Nitrite (as N) | Melbourne | Jul 17, 2024 | 2 Days |
| - Method: LTM-INO-4450 Nitrogens by Discrete Analyser | | | |
| Total Kjeldahl Nitrogen (as N) | Melbourne | Jul 17, 2024 | 28 Days |
| - Method: APHA 4500-Norg B,D Total Kjeldahl Nitrogen by FIA | | | |
| Phosphate total (as P) | Sydney | Jul 17, 2024 | 28 Days |
| - Method: E052 Total Phosphate (as P) | | | |
| Heavy Metals (filtered) | Sydney | Jul 17, 2024 | 180 Days |
| - Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS | | | |



email: EnviroSales@eurofins.com

Eurofins Environment Testing Australia Pty Ltd

ABN: 50 005 085 521

Melbourne Geelong Sydney Canberra Brisbane 6 Monterey Road 19/8 Lewalan Street 179 Magowar Road Unit 1,2 Dacre Street 1/21 Smallwood Place 1/2 Frost Drive Dandenong South Grovedale Girraween Mitchell Murarrie VIC 3175 VIC 3216 NSW 2145 ACT 2911 QLD 4172 +61 3 8564 5000 +61 2 9900 8400 +61 3 8564 5000 +61 2 6113 8091 T: +61 7 3902 4600 NATA# 1261 NATA# 1261 NATA# 1261 NATA# 1261 NATA# 1261 Site# 20794 & 2780 Site# 1254 Site# 25403 Site# 18217 Site# 25466

ABN: 91 05 0159 898

Perth 46-48 Banksia Road Welshpool WA 6106 +61 8 6253 4444 NATA# 2377 Site# 2370

ABN: 47 009 120 549 NZBN: 9429046024954

> Auckland 35 O'Rorke Road Penrose, Auckland 1061 +64 9 526 4551 IANZ# 1327

Auckland (Focus) Christchurch Unit C1/4 Pacific Rise. 43 Detroit Drive Mount Wellington, Rolleston, Auckland 1061 +64 3 343 5201 +64 9 525 0568 IANZ# 1308 IANZ# 1290

Tauranga 1277 Cameron Road. Gate Pa, Christchurch 7675 Tauranga 3112 +64 9 525 0568 IANZ# 1402

Address

web: www.eurofins.com.au

Company Name: Senversa Pty Ltd NSW Level 24, 1 Market Street

SYDNEY NSW 2000

Project Name: Project ID:

WETHERILL PARK WME

S20102

Order No.: Report #: Phone:

Fax:

Newcastle

Mayfield West

+61 2 4968 8448

NSW 2304

NATA# 1261

Site# 25079

1117968 02 9994 8016 03 9606 0074

Perth ProMicro

+61 8 6253 4444

Welshpool

WA 6106

NATA# 2561

Site# 2554

46-48 Banksia Road

Received: Jul 15, 2024 2:30 PM Jul 22, 2024 Due: Priority: 5 Dav

Contact Name: Emma Walsh

Eurofins Analytical Services Manager: Adam Bateup

| | | Sa | mple Detail | | | Iron (filtered) | Manganese (filtered) | Total Recoverable Hydrocarbons | Eurofins Suite B7 (filtered metals) |
|------|-----------------|--------------|------------------|--------|---------------|-----------------|----------------------|--------------------------------|-------------------------------------|
| Sydr | ney Laboratory | NATA # 1261 | Site # 18217 | • | | Χ | Χ | Х | Χ |
| Exte | rnal Laboratory | | | | | | | | |
| No | Sample ID | Sample Date | Sampling Time | Matrix | LAB ID | | | | |
| 1 | QC204 | Jul 11, 2024 | | Water | S24-JI0037338 | Х | Х | Х | Х |
| Test | Counts | | | | | 1 | 1 | 1 | 1 |



Internal Quality Control Review and Glossary

General

- 1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request
- 2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
- 3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
- 4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
- Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences
- Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
- 7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
- 8. Samples were analysed on an 'as received' basis.
- 9. Information identified in this report with blue colour indicates data provided by customers that may have an impact on the results.
- 10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date: therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days

Units

mg/kg: milligrams per kilogram mg/L: milligrams per litre ppm: parts per million μg/L: micrograms per litre ppb: parts per billion %: Percentage

org/100 mL: Organisms per 100 millilitres NTU: Nephelometric Turbidity Units MPN/100 mL: Most Probable Number of organisms per 100 millilitres

Colour: Pt-Co Units (CU) CFU: Colony Forming Unit

Terms

APHA American Public Health Association CEC Cation Exchange Capacity COC Chain of Custody

Client Parent - QC was performed on samples pertaining to this report CP CRM Certified Reference Material (ISO17034) - reported as percent recovery.

Dry Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis

Duplicate A second piece of analysis from the same sample and reported in the same units as the result to show comparison.

LOR Limit of Reporting.

LCS Laboratory Control Sample - reported as percent recovery.

Method Blank In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water NCP Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.

RPD Relative Percent Difference between two Duplicate pieces of analysis SPIKE Addition of the analyte to the sample and reported as percentage recovery

SRA Sample Receipt Advice

The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria Surr - Surrogate

Tributyltin oxide (bis-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits. TRTO

TCI P Toxicity Characteristic Leaching Procedure TEQ Toxic Equivalency Quotient or Total Equivalence

QSM US Department of Defense Quality Systems Manual Version 6.0

US EPA United States Environmental Protection Agency

WA DWER Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR: No Limit

Results between 10-20 times the LOR: RPD must lie between 0-50% Results >20 times the LOR: RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 - 150%, VOC recoveries 50 - 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

- 1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided
- 2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
- 3. pH and Free Chlorine analysed in the laboratory Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time. Analysis will begin as soon as possible after sample receipt.
- 4. Recovery Data (Spikes & Surrogates) where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
- 5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
- 6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data



Quality Control Results

| Test | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|---|-----------|----------|----------------------|----------------|--------------------|
| Method Blank | · | | | | |
| Total Recoverable Hydrocarbons | | | | | |
| TRH C6-C9 | mg/L | < 0.02 | 0.02 | Pass | |
| TRH C10-C14 | mg/L | < 0.05 | 0.05 | Pass | |
| TRH C15-C28 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C29-C36 | mg/L | < 0.1 | 0.1 | Pass | |
| TRH C6-C10* | mg/L | < 0.02 | 0.02 | Pass | |
| TRH >C10-C16* | mg/L | < 0.05 | 0.05 | Pass | |
| TRH >C16-C34* | mg/L | < 0.1 | 0.1 | Pass | |
| TRH >C34-C40 | mg/L | < 0.1 | 0.1 | Pass | |
| Method Blank | | | | | |
| ВТЕХ | | | | | |
| Benzene | mg/L | < 0.001 | 0.001 | Pass | |
| Toluene | mg/L | < 0.001 | 0.001 | Pass | |
| Ethylbenzene | mg/L | < 0.001 | 0.001 | Pass | |
| m&p-Xylenes | mg/L | < 0.002 | 0.002 | Pass | |
| o-Xylene | mg/L | < 0.001 | 0.001 | Pass | |
| Xylenes - Total* | mg/L | < 0.003 | 0.003 | Pass | |
| Method Blank | | | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fraction | ıs | | | | |
| Naphthalene | mg/L | < 0.01 | 0.01 | Pass | |
| Method Blank | | 1 9.9 . | 1 9.0. | 1 466 | |
| Polycyclic Aromatic Hydrocarbons | | | | | |
| Acenaphthene | mg/L | < 0.001 | 0.001 | Pass | |
| Acenaphthylene | mg/L | < 0.001 | 0.001 | Pass | |
| Anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Benz(a)anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(a)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(b&j)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(g.h.i)perylene | mg/L | < 0.001 | 0.001 | Pass | |
| Benzo(k)fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Chrysene | mg/L | < 0.001 | 0.001 | Pass | |
| Dibenz(a.h)anthracene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluoranthene | mg/L | < 0.001 | 0.001 | Pass | |
| Fluorene | mg/L | < 0.001 | 0.001 | Pass | |
| Indeno(1.2.3-cd)pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Naphthalene | mg/L | < 0.001 | 0.001 | Pass | |
| Phenanthrene | mg/L | < 0.001 | 0.001 | Pass | |
| Pyrene | mg/L | < 0.001 | 0.001 | Pass | |
| Method Blank | ı ıııg/ L | V 0.001 | 0.001 | 1 400 | |
| Nitrate & Nitrite (as N) | mg/L | < 0.05 | 0.05 | Pass | |
| Nitrate (as N) | mg/L | < 0.02 | 0.02 | Pass | |
| Nitrite (as N) | mg/L | < 0.02 | 0.02 | Pass | |
| Phosphate total (as P) | mg/L | < 0.01 | 0.02 | Pass | |
| Total Kjeldahl Nitrogen (as N) | mg/L | < 0.2 | 0.01 | Pass | |
| Method Blank | IIIg/L | \ U.Z | 0.2 | 1 055 | |
| Heavy Metals | | T T | | | |
| Arsenic (filtered) | ma/l | < 0.001 | 0.001 | Pass | |
| Cadmium (filtered) | mg/L | | 0.001 | Pass | |
| · · · · · | mg/L | < 0.0002 | | 1 | |
| Chromium (filtered) | mg/L | < 0.001 | 0.001 | Pass | |
| Copper (filtered) | mg/L | < 0.001 | 0.001 | Pass | |
| Iron (filtered) | mg/L | < 0.05 | 0.05 | Pass | |



| Test | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|---|---------|----------|----------------------|----------------|--------------------|
| Lead (filtered) | mg/L | < 0.001 | 0.001 | Pass | |
| Manganese (filtered) | mg/L | < 0.005 | 0.005 | Pass | |
| Mercury (filtered) | mg/L | < 0.0001 | 0.0001 | Pass | |
| Nickel (filtered) | mg/L | < 0.001 | 0.001 | Pass | |
| Zinc (filtered) | mg/L | < 0.005 | 0.005 | Pass | |
| LCS - % Recovery | | | | | |
| Total Recoverable Hydrocarbons | | | | | |
| TRH C6-C9 | % | 104 | 70-130 | Pass | |
| TRH C10-C14 | % | 77 | 70-130 | Pass | |
| TRH C6-C10* | % | 105 | 70-130 | Pass | |
| TRH >C10-C16* | % | 75 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| BTEX | | | | | |
| Benzene | % | 100 | 70-130 | Pass | |
| Toluene | % | 111 | 70-130 | Pass | |
| Ethylbenzene | % | 109 | 70-130 | Pass | |
| m&p-Xylenes | % | 113 | 70-130 | Pass | |
| o-Xylene | % | 109 | 70-130 | Pass | |
| Xylenes - Total* | % | 112 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Total Recoverable Hydrocarbons - 2013 NEPM Fr | actions | | | | |
| Naphthalene | % | 112 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Polycyclic Aromatic Hydrocarbons | | | | | |
| Acenaphthene | % | 112 | 70-130 | Pass | |
| Acenaphthylene | % | 109 | 70-130 | Pass | |
| Anthracene | % | 118 | 70-130 | Pass | |
| Benz(a)anthracene | % | 107 | 70-130 | Pass | |
| Benzo(a)pyrene | % | 109 | 70-130 | Pass | |
| Benzo(b&j)fluoranthene | % | 109 | 70-130 | Pass | |
| Benzo(g.h.i)perylene | % | 110 | 70-130 | Pass | |
| Benzo(k)fluoranthene | % | 104 | 70-130 | Pass | |
| Chrysene | % | 114 | 70-130 | Pass | |
| Dibenz(a.h)anthracene | % | 111 | 70-130 | Pass | |
| Fluoranthene | % | 117 | 70-130 | Pass | |
| Fluorene | % | 117 | 70-130 | Pass | |
| Indeno(1.2.3-cd)pyrene | % | 109 | 70-130 | Pass | |
| Naphthalene | % | 84 | 70-130 | Pass | |
| Phenanthrene | % | 122 | 70-130 | Pass | |
| Pyrene | % | 117 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Nitrate & Nitrite (as N) | % | 103 | 70-130 | Pass | |
| Nitrite (as N) | % | 106 | 70-130 | Pass | |
| Phosphate total (as P) | % | 105 | 70-130 | Pass | |
| Total Kjeldahl Nitrogen (as N) | % | 76 | 70-130 | Pass | |
| LCS - % Recovery | | | | | |
| Heavy Metals | | | | | |
| Arsenic (filtered) | % | 91 | 80-120 | Pass | |
| Cadmium (filtered) | % | 93 | 80-120 | Pass | |
| Chromium (filtered) | % | 91 | 80-120 | Pass | |
| Copper (filtered) | % | 91 | 80-120 | Pass | |
| Iron (filtered) | % | 89 | 80-120 | Pass | |
| Lead (filtered) | % | 89 | 80-120 | Pass | |
| Manganese (filtered) | % | 92 | 80-120 | Pass | |



| Test | | | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
|--------------------------------|---------------------|--------------|-------|----------|----------------------|----------------|--------------------|
| Mercury (filtered) | | | % | 100 | 80-120 | Pass | |
| Nickel (filtered) | | | % | 92 | 80-120 | Pass | |
| Zinc (filtered) | | | % | 91 | 80-120 | Pass | |
| Test | Lab Sample ID | QA Source | Units | Result 1 | Acceptance Limits | Pass Limits | Qualifying Code |
| Spike - % Recovery | | | | | | | |
| Total Recoverable Hydrocarbons | 5 | | | Result 1 | | | |
| TRH C6-C9 | S24-JI0038805 | NCP | % | 81 | 70-130 | Pass | |
| TRH C10-C14 | S24-JI0037155 | NCP | % | 81 | 70-130 | Pass | |
| TRH C6-C10* | S24-JI0038805 | NCP | % | 81 | 70-130 | Pass | |
| TRH >C10-C16* | S24-JI0037155 | NCP | % | 82 | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | |
| BTEX | | | | Result 1 | | | |
| Benzene | S24-JI0038805 | NCP | % | 77 | 70-130 | Pass | |
| Toluene | S24-JI0038805 | NCP | % | 87 | 70-130 | Pass | |
| Ethylbenzene | S24-JI0038805 | NCP | % | 86 | 70-130 | Pass | |
| m&p-Xylenes | S24-JI0038805 | NCP | % | 92 | 70-130 | Pass | |
| o-Xylene | S24-JI0038805 | NCP | % | 88 | 70-130 | Pass | |
| Xylenes - Total* | S24-JI0038805 | NCP | % | 90 | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | |
| Total Recoverable Hydrocarbons | s - 2013 NEPM Fract | ions | | Result 1 | | | |
| Naphthalene | S24-JI0038805 | NCP | % | 92 | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | |
| Polycyclic Aromatic Hydrocarbo | ns | | | Result 1 | | | |
| Acenaphthene | W24-JI0032492 | NCP | % | 103 | 70-130 | Pass | |
| Acenaphthylene | W24-JI0032492 | NCP | % | 100 | 70-130 | Pass | |
| Anthracene | W24-JI0032492 | NCP | % | 111 | 70-130 | Pass | |
| Benz(a)anthracene | W24-JI0032492 | NCP | % | 99 | 70-130 | Pass | |
| Benzo(a)pyrene | W24-JI0032492 | NCP | % | 102 | 70-130 | Pass | |
| Benzo(b&j)fluoranthene | W24-JI0032492 | NCP | % | 99 | 70-130 | Pass | |
| Benzo(g.h.i)perylene | W24-JI0032492 | NCP | % | 103 | 70-130 | Pass | |
| Benzo(k)fluoranthene | W24-JI0032492 | NCP | % | 102 | 70-130 | Pass | |
| Chrysene | W24-JI0032492 | NCP | % | 105 | 70-130 | Pass | |
| Dibenz(a.h)anthracene | W24-JI0032492 | NCP | % | 102 | 70-130 | Pass | |
| Fluoranthene | W24-JI0032492 | NCP | % | 110 | 70-130 | Pass | |
| Fluorene | W24-JI0032492 | NCP | % | 111 | 70-130 | Pass | |
| Indeno(1.2.3-cd)pyrene | W24-JI0032492 | NCP | % | 103 | 70-130 | Pass | |
| Naphthalene | W24-JI0032492 | NCP | % | 79 | 70-130 | Pass | |
| Phenanthrene | W24-JI0032492 | NCP | % | 116 | 70-130 | Pass | |
| Pyrene | W24-JI0032492 | NCP | % | 111 | 70-130 | Pass | |
| Spike - % Recovery | | | | 1 | | | |
| | | | | Result 1 | | | |
| Nitrate & Nitrite (as N) | M24-JI0040441 | NCP | % | 113 | 70-130 | Pass | |
| Nitrite (as N) | M24-JI0040441 | NCP | % | 100 | 70-130 | Pass | |
| Phosphate total (as P) | S24-JI0037338 | CP | % | 79 | 70-130 | Pass | |
| Total Kjeldahl Nitrogen (as N) | M24-JI0014226 | NCP | % | 98 | 70-130 | Pass | |
| Spike - % Recovery | | | | | | | |
| Heavy Metals | | | | Result 1 | | | |
| Arsenic (filtered) | S24-JI0045319 | NCP | % | 88 | 75-125 | Pass | |
| Cadmium (filtered) | S24-JI0045319 | NCP | % | 95 | 75-125 | Pass | |
| Chromium (filtered) | S24-JI0045319 | NCP | % | 88 | 75-125 | Pass | |
| Copper (filtered) | S24-JI0045319 | NCP | % | 90 | 75-125 | Pass | |
| Iron (filtered) | S24-JI0045319 | NCP | % | 91 | 75-125 | Pass | |
| Lead (filtered) | S24-JI0045319 | NCP | % | 87 | 75-125 | Pass | |
| Manganese (filtered) | S24-JI0045319 | NCP | % | 91 | 75-125 | Pass | |



| Test | Lab Sample ID | QA Source | Units | Result 1 | | | Acceptance Limits | Pass Limits | Qualifying Code |
|-------------------------------------|--------------------------------|--------------|-------|----------|----------|----------|----------------------|----------------|--------------------|
| Mercury (filtered) | S24-JI0045319 | NCP | % | 94 | | | 75-125 | Pass | |
| Nickel (filtered) | S24-JI0045319 | NCP | % | 90 | | | 75-125 | Pass | |
| Zinc (filtered) | S24-JI0045319 | NCP | % | 93 | | | 75-125 | Pass | |
| Test | Lab Sample ID | QA Source | Units | Result 1 | | | Acceptance Limits | Pass Limits | Qualifying Code |
| Duplicate | | | | | | | | | |
| Total Recoverable Hydrocarbons | | | | Result 1 | Result 2 | RPD | | | |
| TRH C6-C9 | S24-JI0038806 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| TRH C10-C14 | S24-JI0037154 | NCP | mg/L | 0.06 | 0.07 | 13 | 30% | Pass | |
| TRH C15-C28 | S24-JI0037154 | NCP | mg/L | 0.3 | 0.2 | 19 | 30% | Pass | |
| TRH C29-C36 | S24-JI0037154 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| TRH C6-C10* | S24-JI0038806 | NCP | mg/L | < 0.02 | < 0.02 | <1 | 30% | Pass | |
| TRH >C10-C16* | S24-JI0037154 | NCP | mg/L | 0.08 | 0.09 | 5.7 | 30% | Pass | |
| TRH >C16-C34* | S24-JI0037154 | NCP | mg/L | 0.3 | 0.2 | 26 | 30% | Pass | |
| TRH >C34-C40 | S24-JI0037154 | NCP | mg/L | < 0.1 | < 0.1 | <1 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| BTEX | | | | Result 1 | Result 2 | RPD | | | |
| Benzene | S24-JI0038806 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Toluene | S24-JI0038806 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Ethylbenzene | S24-JI0038806 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| m&p-Xylenes | S24-JI0038806 | NCP | mg/L | < 0.002 | < 0.002 | <1 | 30% | Pass | |
| o-Xylene | S24-JI0038806 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Xylenes - Total* | S24-JI0038806 | NCP | mg/L | < 0.003 | < 0.003 | <1 | 30% | Pass | |
| Duplicate | | , | J | • | | | • | | |
| Total Recoverable Hydrocarbons | - 2013 NEPM Frac | tions | | Result 1 | Result 2 | RPD | | | |
| Naphthalene | S24-JI0038806 | NCP | mg/L | < 0.01 | < 0.01 | <1 | 30% | Pass | |
| Duplicate | | | | | | | | | |
| Polycyclic Aromatic Hydrocarboi | ns | | | Result 1 | Result 2 | RPD | | | |
| Acenaphthene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Acenaphthylene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Anthracene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Benz(a)anthracene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Benzo(a)pyrene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Benzo(b&i)fluoranthene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Benzo(g.h.i)perylene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Benzo(k)fluoranthene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Chrysene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Dibenz(a.h)anthracene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Fluoranthene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Fluorene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| | N24-JI0037864 | NCP | | < 0.001 | | | 30% | Pass | |
| Indeno(1.2.3-cd)pyrene Naphthalene | N24-JI0037864 N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 <1 | 30% | Pass | |
| Phenanthrene | N24-JI0037864 N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| | | | mg/L | 1 | < 0.001 | <1 | | | |
| Pyrene | N24-JI0037864 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Duplicate | | | | Danista | Decition | DDD | | | |
| Alteria O Alteria (All | DO4 H0007503 | NCD | | Result 1 | Result 2 | RPD | 0001 | D- | |
| Nitrate & Nitrite (as N) | B24-JI0037599 | NCP | mg/L | 0.39 | 0.38 | 3.0 | 30% | Pass | |
| Nitrite (as N) | B24-JI0037599 | NCP | mg/L | 0.25 | 0.25 | <1 | 30% | Pass | |
| Phosphate total (as P) | N24-JI0035143 | NCP | mg/L | 7.7 | 7.7 | <1 | 30% | Pass | |
| Total Kjeldahl Nitrogen (as N) | S24-JI0038669 | NCP | mg/L | 1500 | 1700 | 8.3 | 30% | Pass | |



| Duplicate | | | | | | | | | |
|----------------------|---------------|-----|------|----------|----------|-----|-----|------|-----|
| Heavy Metals | | | | Result 1 | Result 2 | RPD | | | |
| Arsenic (filtered) | S24-JI0045318 | NCP | mg/L | 0.002 | 0.002 | 4.4 | 30% | Pass | |
| Cadmium (filtered) | S24-JI0045318 | NCP | mg/L | < 0.0002 | < 0.0002 | <1 | 30% | Pass | |
| Chromium (filtered) | S24-JI0045318 | NCP | mg/L | 0.12 | 0.12 | 2.2 | 30% | Pass | |
| Copper (filtered) | S24-JI0045318 | NCP | mg/L | 0.002 | 0.002 | 11 | 30% | Pass | |
| Iron (filtered) | S24-JI0045318 | NCP | mg/L | < 0.05 | < 0.05 | <1 | 30% | Pass | |
| Lead (filtered) | S24-JI0045318 | NCP | mg/L | < 0.001 | < 0.001 | <1 | 30% | Pass | |
| Manganese (filtered) | S24-JI0045318 | NCP | mg/L | < 0.005 | < 0.005 | <1 | 30% | Pass | |
| Mercury (filtered) | S24-JI0045318 | NCP | mg/L | < 0.0001 | < 0.0001 | <1 | 30% | Pass | |
| Nickel (filtered) | S24-JI0045318 | NCP | mg/L | 0.002 | 0.001 | 32 | 30% | Fail | Q15 |
| Zinc (filtered) | S24-JI0045318 | NCP | mg/L | 0.005 | < 0.005 | 10 | 30% | Pass | |



Comments

Sample Integrity

Custody Seals Intact (if used) N/A Attempt to Chill was evident Yes Sample correctly preserved Yes Appropriate sample containers have been used Yes Sample containers for volatile analysis received with minimal headspace Yes Samples received within HoldingTime Yes Some samples have been subcontracted No

Qualifier Codes/Comments

Code Description

F2 is determined by arithmetically subtracting the "naphthalene" value from the ">C10-C16" value. The naphthalene value used in this calculation is obtained from volatiles (Purge & Trap analysis). N01

Where we have reported both volatile (P&T GCMS) and semivolatile (GCMS) naphthalene data, results may not be identical. Provided correct sample handling protocols have been followed, any observed differences in results are likely to be due to procedural differences within each methodology. Results determined by both techniques have passed all QAQC acceptance criteria, and are entirely technically valid.

N02

F1 is determined by arithmetically subtracting the "Total BTEX" value from the "C6-C10" value. The "Total BTEX" value is obtained by summing the concentrations of BTEX analytes. The "C6-C10" value is obtained by quantitating against a standard of mixed aromatic/aliphatic analytes. N04

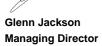
Please note:- These two PAH isomers closely co-elute using the most contemporary analytical methods and both the reported concentration (and the TEQ) apply specifically to the total of the two co-eluting PAHs N07

The RPD reported passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of this report.

Authorised by:

Q15

Nileshni Goundar Analytical Services Manager Caitlin Breeze Senior Analyst-Inorganic Maria Tian Senior Analyst-Organic Mickael Ros Senior Analyst-Metal Roopesh Rangarajan Senior Analyst-Organic Roopesh Rangarajan Senior Analyst-Volatile Ryan Phillips Senior Analyst-Inorganic



Final Report - this report replaces any previously issued Report

- Indicates Not Requested
- * Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here

Eurofins shall not be liable for loss, cost, damages or expenses incurred by the client, or any other person or company, resulting from the use of any information or interpretation given in this report. In no case shall Eurofins be liable for consequential damages including, but not limited to, lost profits, damages for failure to meet deadlines and lost production arising from this report. This document shall not be reproduced except in full and relates only to the items tested. Unless indicated otherwise, the tests were performed on the samples as received.

Appendix F: Mann Kendall Groundwater Trend Analysis

| Location | Chem Name | Unit | Earliest | Latest | number_results | number_detects | minimum | maximum | average | percentile80 | mann_kendall_trend |
|-------------|---|--------------|----------------------|------------------------|----------------|----------------|----------------------|----------------|-------------------|--------------|--------------------|
| Code MW1 | Phenol | μg/L | 8-Feb-23 | 8-Feb-23 | 1 | 0 | <1.0 | <1.0 | 1 | 1 | |
| MW1 | Nitrate (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.01 | <0.1 | 0.035 | | No Trend |
| MW1 | Manganese | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.59 | 4.84 | 2.1525 | | Stable |
| MW1 | Nickel | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.015 | 0.156 | 0.0575 | | No Trend |
| MW1 | Arsenic | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 0.007 | 0.012 | 0.0095 | 0.0114 | Stable |
| MW1 | Chromium | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 0 | < 0.001 | <0.001 | 0.001 | 0.001 | |
| MW1 | Copper | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 2 | < 0.001 | 0.015 | 0.005 | 0.0078 | No Trend |
| MW1 | Zinc | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 0.012 | 0.174 | 0.06175 | 0.0966 | No Trend |
| MW1 | Ammonia (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 0.19 | 0.71 | 0.4675 | 0.578 | Decreasing |
| MW1 | Total BTEX | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | 0 | <1.0 | <1.0 | 1 | 1 | |
| MW1 | >C10-C40 Fraction (Sum) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | 1 | <100.0 | 520.0 | 205 | 268 | No Trend |
| MW1 | C6-C10 Fraction minus BTEX (F1) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <20.0 | <20.0 | 20 | 20 | |
| MW1 | Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.5 | <0.5 | 0.5 | 0.5 | |
| MW2 | Phenol | μg/L | 8-Feb-23 | 8-Feb-23 | 1 | | <1.0 | <1.0 | 1 | 1 | |
| MW2 | Nitrate (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 2 <0.01 | 0.03 | 0.0175 | | Stable |
| MW2 | Manganese | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.96 | 3.28 | 1.75 | | Increasing |
| MW2 | Nickel | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.005 | 0.006 | 0.00575 | | Stable |
| MW2 | Arsenic | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.004 | 0.008 | 0.005 | 0.0056 | |
| MW2 | Chromium | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | < 0.001 | <0.001 | 0.001 | 0.001 | |
| MW2 | Copper | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.001 | 0.011 | 0.0035 | | No Trend |
| MW2 | Zinc | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | < 0.005 | 0.009 | 0.00775 | | Stable Stable |
| MW2 | Ammonia (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.26 | 0.52 | 0.435 | | Stable |
| MW2 | Total BTEX | μg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | <1.0 | <1.0 <100.0 | 100 | 100 | |
| MW2 MW2 | >C10-C40 Fraction (Sum) | μg/L | 8-Feb-23 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | 0 <100.0 0 <20.0 | <20.0 | 20 | 20 | |
| MW2 | C6-C10 Fraction minus BTEX (F1) Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | 0 < 0.5 | <0.5 | 0.5 | 0.5 | |
| MW3 | Phenol | μg/L | 8-Feb-23 | 8-Feb-23 | 1 | | <1.0 | <1.0 | 0.3 | 0.3 | |
| MW3 | Nitrate (as N) | μg/L mg/L | 8-Feb-23 | 11-Jul-24 | 7 | | 3 < 0.01 | 0.1 | 0.028571429 | | No Trend |
| MW3 | Manganese | mg/L | 8-Feb-23 | 11-Jul-24 | 11 | | 5.8 | 7.4 | 6.533636364 | | Increasing |
| MW3 | Nickel | mg/L | 8-Feb-23 | 11-Jul-24 | 12 | | 0.16 | 0.207 | 0.189833333 | | Stable |
| MW3 | Arsenic | mg/L | 8-Feb-23 | 11-Jul-24 | 12 | | 0.002 | 0.011 | 0.00775 | | Stable |
| MW3 | Chromium | mg/L | 8-Feb-23 | 11-Jul-24 | 12 | | 3 < 0.001 | <0.01 | 0.00675 | | Stable |
| MW3 | Copper | mg/L | 8-Feb-23 | 11-Jul-24 | 12 | | 3 < 0.001 | <0.01 | 0.006833333 | | Stable |
| MW3 | Zinc | mg/L | 8-Feb-23 | 11-Jul-24 | 12 | | 0.074 | 0.253 | 0.195416667 | 0.2462 | |
| MW3 | Ammonia (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.22 | 0.29 | 0.27 | | Stable |
| MW3 | Total BTEX | μg/L | 8-Feb-23 | 11-Jul-24 | 8 | 0 | <1.0 | <1.0 | 1 | 1 | |
| MW3 | >C10-C40 Fraction (Sum) | μg/L | 8-Feb-23 | 11-Jul-24 | 12 | 4 | <100.0 | 910.0 | 226.6666667 | 346 | No Trend |
| MW3 | C6-C10 Fraction minus BTEX (F1) | μg/L | 8-Feb-23 | 11-Jul-24 | 12 | 0 | <10.0 | <20.0 | 19.16666667 | 20 | Stable |
| MW3 | Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L | 8-Feb-23 | 11-Jul-24 | 11 | 0 | <0.5 | <1.0 | 0.636363636 | 1 | Stable |
| MW4 | Phenol | μg/L | 8-Feb-23 | 8-Feb-23 | 1 | 0 | <1.0 | <1.0 | 1 | 1 | |
| MW4 | Nitrate (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 1 | 0.01 | <0.01 | 0.01 | 0.01 | |
| MW4 | Manganese | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 4.0 | 6.04 | 5.13 | 5.686 | Stable |
| MW4 | Nickel | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 0.011 | 0.021 | 0.01725 | 0.0204 | Decreasing |
| MW4 | Arsenic | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | 4 | 0.005 | 0.008 | 0.00675 | 0.0074 | Stable |
| MW4 | Chromium | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.001 | <0.001 | 0.001 | 0.001 | |
| MW4 | Copper | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.001 | 0.005 | 0.002 | | No Trend |
| MW4 | Zinc | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | . <0.005 | 0.006 | 0.00525 | 0.0054 | |
| MW4 | Ammonia (as N) | mg/L | 8-Feb-23 | 11-Jul-24 | 4 | | 0.28 | 0.34 | 0.31 | | Decreasing |
| MW4 | Total BTEX | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <1.0 | 2.0 | 1.25 | | Stable |
| MW4 | >C10-C40 Fraction (Sum) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <100.0 | <100.0 | 100 | 100 | |
| MW4 | C6-C10 Fraction minus BTEX (F1) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <20.0 | <20.0 | 20 | 20 | |
| MW4 | Sum of Polycyclic aromatic hydrocarbons (PAH) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | | <0.5 | <0.5 | 0.5 | 0.5 | |
| MW6 | Phenol Nitrate (as Ni) | μg/L | 8-Feb-23 | 8-Feb-23 | 1 | | <1.0 | <1.0 | - | - | Ctable |
| MW6 MW6 | Nitrate (as N) | mg/L | 8-Feb-23 8-Feb-23 | 11-Jul-24 | 4 | | 0.18 <0.01 | 1.93 0.225 | 1.1875 0.08375 | | Stable No Trend |
| | Manganese Nickel | mg/L | 8-Feb-23 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | 2 <0.01 2 <0.001 | | 0.08375 | 0.126 | |
| MW6 MW6 | Arsenic | mg/L mg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | 2 <0.001 2 <0.001 | 0.002 | 0.00125 | | Stable |
| MW6 | Chromium | mg/L mg/L | 8-Feb-23 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | 0.001 | <0.002 | 0.0015 | 0.002 | |
| MW6 | Copper | mg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | . <0.001 | 0.001 | 0.001 | 0.001 | |
| MW6 | Zinc | mg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | . <0.001 | 0.005 | 0.0015 | 0.0018 | |
| MW6 | Ammonia (as N) | mg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | ! <0.005 ! <0.01 | 0.006 | 0.00325 | | No Trend |
| MW6 | Total BTEX | μg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | <1.0 | <1.0 | 0.0323 | 0.048 | |
| | >C10-C40 Fraction (Sum) | μg/L μg/L | 8-Feb-23 | 11-Jul-24 11-Jul-24 | 4 | | <1.0 | <1.0 | 100 | 100 | |
| N/I/A/6 | | μ8/ L | 0-1-60-23 | 11-Jul-24 | 4 | U | , 100.0 | -100.0 | 100 | 100 | |
| MW6 MW6 | C6-C10 Fraction minus BTEX (F1) | μg/L | 8-Feb-23 | 11-Jul-24 | 4 | 0 | <20.0 | <20.0 | 20 | 20 | |

Senversa Pty Ltd

ABN 89 132 231 380 www.senversa.com.au enquiries@senversa.com.au LinkedIn: Senversa

LinkedIn: Senversa Facebook: Senversa





To the extent permissible by law, Senversa shall not be liable for any errors, omissions, defects or misrepresentations, or for any loss or damage suffered by any persons (including for reasons of negligence or otherwise).

©2024 Senversa Pty Ltd



Appendix B – Example Quarterly OEMP Checklist





| • | RECYCLING | AN – WETHERILL PARK | INS | SPECTION | CHECK | KLIST |
|--------|---|--|----------------|--------------------------------------|------------------|--|
| Locat | tion: | reDirect – Wetherill Park | D | ate: | 28.06.2 | 4 |
| Inspe | ection Completed By: | M.Stewart | Si | gnature: | m. P | Stewart |
| 1. Ge | neral Management and m | nitigations \square N/A | | Frequency | Y/N/NA | General Comments |
| 1.2 | Employees and contract trained. | ors have been inducted and are suitab | As required | Y | | |
| 1.3 | Plant and equipment be the start of the day? | ing used is in good working condition | at | Daily | Y | |
| 2. Tra | iffic mitigations \Box N | I/A | Frequency | Y/N/NA | General Comments | |
| 2.1 | Traffic is continually mo | onitored by Operations Coordinator? | | Daily | Υ | |
| 2.2 | Alll car spaces are free to | from obstruction and maintained for upress? | ise | Daily | Y | |
| 2.3 | Vehicles are entering ar | nd leaving the site in forward direction | ١. | Daily | Υ | |
| 2 Air | quality odour and dust r | nitigations N/A | | Frequency | Y/N/NA | General Comments |
| 3. All | 3. Air quality, odour and dust mitigations □ N/A Good dust management procedures are being implemented | | | | T/IV/IVA | General Comments |
| 3.1 | (inside building): Sweeper working and be | | Daily | Y | | |
| 3.2 | Good dust management procedures are implemented (outside the building): Sweeper working and being used? | | | | Υ | |
| 3.3 | Residual waste has been waste bin capacity)? | n transported offsite (check general | | Daily | Υ | |
| | | | | | | |
| 5. Sto | rmwater mitigations | N/A | | Frequency | Y/N/NA | General Comments |
| 5.1 | Are there any spills that | t have been left unattended? | | Daily | N | |
| 5.2 | | s been inspected for any build up of and vegetation within drainage systen | n? | Monthly | Υ | |
| 5.3 | If materials identified in removed? | stormwater drains, has it been | | Monthly | Y | |
| 5.4 | Inflow areas and pit gra | tes have been inspected and clear of | | Monthly | Y | |
| 5.5 | 1 1 | aters, first flush devices and litter and are operating correctly. | | Monthly | Y | |
| 5.6 | Site structires to be regularly checked for erosion and scouring | | | Monthly | Υ | |
| 5.7 | 5.7 Treatment areas and structures will be regularly checked for the build up of litter material | | | | Υ | |
| 5.8 | collected sediment, del | ect internal walls and base. Remove ar oris, litter and vegetation. Inspect and lowing any removal of objects. Ensure e upon refitment. | | Quarterly (Mar, Jun, Sep, Dec) | Y | Lift grate, brush out lip for grate and down walls remove debris replace grate |
| 5.9 | Have all drainage struct | cures been inspected noting any | Bi- | | | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 1 of 4 | | | |

annually

(Jun, Dec)

Inspected no action required

dilapidation, if so have repairs been carried out?





| 5.10 | Rainwater tank – has tank been checked for evidence of litter and functioning properly | Bi- annually (Jun, Dec) | Y | Check Basket – no litter |
|---------|---|-------------------------------|--------|-------------------------------------|
| 5.11 | Rainwater tank – has tank been checked for evidence of access by pests (birds, insects, mosquito larvae ect.) | Bi- annually (Jun, Dec) | Υ | Empty tank inspect no sign of pests |
| 5.12 | Rainwater tank – has structural integrity of tank been inspected? Note any dilapidation or repairs required / completed. | Bi- annually (Jun, Dec) | Y | No repairs required |
| 5.13 | The sediment chamber of the Ecoceptor will be reqgularly checked and cleaned and any damaged covers replaced. | Bi- annually (Jun, Dec) | Y | Checked no action required |
| 6. Ver | min and pest management mitigations N/A | Frequency | Y/N/NA | General Comments |
| 6.1 | Drainage sumps and catch drains will be inspected daily and cleaned regularly to prevent providing a habitat for pests. | Ongoing | Υ | |
| 6.2 | Has the site been inspected for windblown litter? Any identified litter must be removed and disposed appropriately. | Ongoing | Υ | |
| 6.3 | All overhead structures and internal roofs are visually inspected weekly to ensure they are kept clean. | Ongoing | Y | |
| 7. Pol | lution management mitigations | Frequency | Y/N/NA | General Comments |
| 7.1 | Are all dangerous goods stored appropriately according to their ADG classes and compatibility? | Daily | Υ | |
| 7.2 | Has training on the pollution incident response management plan been provided in toolbox? | As required | Υ | |
| 8. Fire | e management mitigations | Frequency | Y/N/NA | General Comments |
| 8.1 | Fire extinguishers are positioned at readily accessible points, including on mobile plant | Daily | Y | |
| 9. Noi | se and vibration mitigations \square N/A | Frequency | Y/N/NA | General Comments |
| 9.1 | Are defective plant parked up and not being used? | As required | Υ | |
| 10. W | aste management mitigations | Frequency | Y/N/NA | General Comments |
| 10.1 | All waste stored on site onsite is permitted by the EPL? | Daily | Y | |
| 10.2 | The total amount of waste stored at the premises is under EPL Authorised Amount? | Daily | Y | |
| 10.3 | The total amount of waste received daily is being recorded via the weighbridges in place? | Daily | Y | |
| | ooding mitigations | Frequency | Y/N/NA | General Comments |
| 11. Flo | - NA | | | |
| 11. Flo | Inspection and maintenance of the Flood Emergency Kit will be undertaken as required to ensure all components are present and in operating condition. | Bi- annually (Jun, Dec) | Υ | Fully stocked and in good condition |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 2 of 4 | | |





| 12. Bi | odiversity 🗆 N/A | Frequency | Y/N/NA | General Comments |
|--------|---|--------------------------------------|--------|------------------|
| 12.1 | Weed treatment will occur alongside maintenance of landscaping within subject site. This supports compliance with the NSW Biosecurity Act 2015. | Quarterly (Mar, Jun, Sep, Dec) | Y | |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 3 of 4 | | |





| Action Plan - to be | transferred as a 'Ha | zard Report' | | |
|---|-----------------------|-----------------|----------------------|-----------------|
| Actions required | Action assigned to | Date assigned | Date to be completed | Signature |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| Storage and Reference | Inspection Comple | eted By | | Date |
| To be reviewed at Site Meeting. | | | | |
| Workplace inspection checklists must be complete the end of each day. | ddaily, stored in the | site file and u | ploaded to Dat | astation before |

| Document Title: WORKPLACE INSPECTION CHECKLIST - PERIODIC | | | | | | | |
|---|--------------|----------|--------------|-----------------------|--------------|--|--|
| Approved By: | Date Issued: | Version: | Review Date: | Author: | Page Number: | | |
| Environmental Manager | 21/06/2022 | 1.0 | 21/06/2025 | Environmental Manager | 4 of 4 | | |



Appendix C – Appendix D – Community Complaints



| Complaint No | Category | Date Received | Property | Detail | Follow Up Actions |
|-----------------|----------|------------------|----------|--------|-------------------|
| NIL | - | - | - | - | - |



Blank Page